

## Effects of Culture Conditions of Immobilized Recombinant *Escherichia coli* on Cyclodextrin Glucanotransferase (CGTase) Excretion and Cell Stability

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### ABSTRACT

The targeting of recombinant proteins for excretion into culture medium presents significant advantages over cytoplasmic expression. However, during the excretion of recombinant protein, caution must be taken in order to avoid cell lysis due to pressure build-up through overproduction of the expressed recombinant protein in the periplasmic space. In the present study, recombinant *Escherichia coli* expressing cyclodextrin glucanotransferase (CGTase) was immobilized by adsorption and entrapment in a porous hollow fiber membrane. The effects of culture conditions (post induction time, agitation rate and pH) on CGTase excretion, cell lysis and plasmid stability of immobilized cells were studied. The optimum post induction time, agitation rate and pH were found to be 24 h, 200 rpm and pH 9, respectively. The immobilized cells exhibited a 2.8–4.6-fold increase in CGTase excretion, a 16–95% reduction of cell lysis and a 323–464% increase in plasmid stability compared with free cells. Hence, immobilizing *E. coli* using a porous hollow fiber membrane proved to be valuable for the excretion of a recombinant protein and increased cell viability.

**KEYWORDS:** Immobilized cell; free cell; CGTase excretion; cell lysis; plasmid stability; porous hollow fiber membrane

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