

**BIOLOGICAL PRODUCTION OF METHANOL BY USING PECTIN
METHYLESTERASE (PME)**

ERNIE ZAYANI BINTI KAMARUZAMAN

**BACHELOR OF CHEMICAL ENGINEERING (BIOTECHNOLOGY)
UNIVERSITI MALAYSIA PAHANG**

©ERNIE ZAYANI BINTI KAMARUZAMAN (2015)

Thesis Access Form

No _____ Location _____

Author :

Title :

Status of access OPEN / RESTRICTED / CONFIDENTIAL

Moratorium period: _____ years, ending _____ / _____ 20 _____

Conditions of access proved by (CAPITALS): **ASSOC. PROF. IR DR NORAZWINA BINTI ZAINOL**

Supervisor (Signature).....

Faculty:

Author's Declaration: *I agree the following conditions:*

OPEN access work shall be made available in the Universiti Malaysia Pahang only and not allowed to reproduce for any purposes.

The statement itself shall apply to ALL copies:

This copy has been supplied on the understanding that it is copyright material and that no quotation from the thesis may be published without proper acknowledgement.

Restricted/confidential work: All access and any photocopying shall be strictly subject to written permission from the University Head of Department and any external sponsor, if any.

Author's signature.....Date:

users declaration: for signature during any Moratorium period (Not Open work):

I undertake to uphold the above conditions:

Date	Name (CAPITALS)	Signature	Address

**BIOLOGICAL PRODUCTION OF METHANOL BY USING PECTIN
METHYLESTERASE (PME)**

ERNIE ZAYANI BINTI KAMARUZAMAN

Thesis submitted in partial fulfilment of the requirements
for the award of the degree of
Bachelor of Chemical Engineering (Biotechnology)

**Faculty of Chemical & Natural Resources Engineering
UNIVERSITI MALAYSIA PAHANG**

JUNE 2015

©ERNIE ZAYANI BINTI KAMARUZAMAN (2015)

SUPERVISOR'S DECLARATION

We hereby declare that we have checked this thesis and in our opinion, this thesis is adequate in terms of scope and quality for the award of the degree of Bachelor of Chemical Engineering (Biotechnology).

Signature :
Name of main supervisor : IR DR. NORAZWINA BINTI ZAINOL
Position : ASSOCIATE PROFESSOR
Date : 5 JUNE 2015

STUDENT'S DECLARATION

I hereby declare that the work in this thesis is my own except for quotations and summaries which have been duly acknowledged. The thesis has not been accepted for any degree and is not concurrently submitted for award of other degree.

Signature :

Name : ERNIE ZAYANI BINTI KAMARUZAMAN

ID Number : KE11033

Date : JUNE 2015

Dedication

To my lovely family for always be there for me through thick and thin.

To all my friends for always support me and help me when needed.

To my kind supervisor for always guide and being understanding person.

ACKNOWLEDGEMENT

I would like to thanks the following people and organisations;

- My supervisors Associate Professor Ir. Dr.Norazwina Bt Zainol for her guidance through an effective well-arranged weekly meeting.
- Miss Siti Natrah Bt Ismail for her guidance on the laboratory work.
- All my friends for their supports.

ABSTRACT

Methanol contains only one carbon atom (CH_3OH) is the simplest alcohol that also known as wood alcohol. It is a colourless liquid, volatile, flammable and poisonous in properties. It is widely used in several industries such as plastic industry. It also been used as an original feedstock to make polymer. This research study was focus on the production of methanol by using pectin methylesterase (PME). This research study consists of two objectives. The first objective of this research was to produce methanol by using pectin methylesterase (PME). The second objective was to analyse the factors that affecting the methanol production from pectin methylesterase (PME). The raw material, lime peels waste and the substrate, pectin solution were prepared. The extractions of PME from lime peels were done. PME and pectin were mixed for enzymatic reaction process. The experiment for the preliminary study was performed. Preliminary study was performed with five factors which were pH, temperature, concentration of NaCl, agitation and enzymatic reaction time to know the highest production of methanol. Design experiment by using Design Expert software was done after preliminary experiment finished. The analysis to know the production of methanol was performed by using high performance liquid chromatography (HPLC). The result of preliminary experiment by using HPLC showed that the highest concentration produced was at pH 7 and temperature of 70 °C with 5 hours enzymatic reaction time and 3 M of concentration of NaCl. The results of 18 runs sample from Design Expert software were analysed by using Response Surface Methodology (RSM) through Fractional Factorial Design (FFD). The results showed that the range yield of methanol was between 0.3 – 2.2 g/g. The main effect analysis showed that concentration of NaCl had contributed the most in methanol production with percentage contribution of 45.36%. The result of interaction between factors showed that the interaction between factor A (pH) and factor B (temperature) at pH 5 and 70 °C produced the highest methanol with 13.22 % contribution. The value of R^2 for the regression analysis was 0.9904. Hence, the methanol was successfully produced from pectin methylesterase (PME) by using lime peels waste.

ABSTRAK

Metanol mengandungi satu karbon atom (CH_3OH) adalah alkohol yang paling mudah yang juga dikenali sebagai alkohol kayu. Ia adalah cecair yang tidak berwarna, tidak menentu, mudah terbakar dan beracun dalam tanah. Ia digunakan secara meluas dalam beberapa industri seperti industri plastik. Ia juga telah digunakan sebagai bahan mentah asal untuk membuat polimer. Kajian penyelidikan ini adalah fokus kepada pengeluaran metanol dengan menggunakan "pectin methylesterase" (PME). Kajian penyelidikan ini terdiri daripada dua objektif. Objektif pertama kajian ini adalah untuk menghasilkan metanol dengan menggunakan (PME). Objektif kedua adalah untuk menganalisis faktor-faktor yang memberi kesan kepada pengeluaran metanol daripada (PME). Bahan mentah iaitu sisa kulit limau dan substrat iaitu cecair pektin telah disediakan. Pengekstrakan PME dari kulit limau telah dilakukan. PME dan pektin bercampur untuk proses tindak balas enzim. Percubaan untuk kajian awal dilakukan. Kajian awal telah dilakukan dengan lima faktor iaitu pH, suhu, kepekatan NaCl, goncangan dan masa tindak balas enzim untuk mengetahui pengeluaran tertinggi metanol. Eksperimen reka bentuk dengan menggunakan perisian "Design Expert" dilakukan selepas percubaan awal selesai. Analisis untuk mengetahui pengeluaran metanol telah dilakukan dengan menggunakan "High Performance Liquid Chromatography" (HPLC). Hasil eksperimen awal dengan menggunakan HPLC menunjukkan bahawa kepekatan tertinggi yang dikeluarkan adalah pada pH 7 dan suhu 70°C dengan 5 jam masa tindak balas enzim dan 3 M daripada kepekatan NaCl. Keputusan daripada 18 sampel berlangsung dari perisian "Design Expert" dianalisis dengan menggunakan "Response Surface Methodology" (RSM) melalui pecahan "Full Factorial Design" (FFD). Hasil kajian menunjukkan bahawa hasil julat metanol adalah antara 0.3-2.2 g / g. Analisis kesan utama menunjukkan bahawa kepekatan NaCl telah menyumbang pengeluaran tertinggi metanol dengan sumbangan peratusan 45.36%. Hasil daripada interaksi antara faktor menunjukkan bahawa interaksi antara faktor A (pH) dan faktor B (suhu) pada pH 5 dan 70°C menghasilkan metanol tertinggi dengan peratusan sebanyak 13.22%. Nilai R^2 untuk analisis regresi adalah 0.9904. Oleh itu, metanol telah berjaya dihasilkan daripada (PME) dengan menggunakan sisa kulit limau.

TABLE OF CONTENTS

SUPERVISOR'S DECLARATION	V
STUDENT'S DECLARATION	VII
DEDICATION.....	VII
ACKNOWLEDGEMENT	VIII
ABSTRACT.....	IXX
ABSTRAK.....	X
TABLE OF CONTENTS.....	XII
LIST OF FIGURES	XIII
LIST OF TABLES.....	XIV
LIST OF ABBREVIATIONS.....	XIV
1 INTRODUCTION	166
1.1 Motivation and statement of problem	166-17
1.2 Objectives.....	18
1.3 Scope of this research.....	18
1.4 Main contribution of this work	19
1.5 Organisation of this thesis	19
2 LITERATURE REVIEW	21
2.1 Overview	21-22
2.2 Methanol	23-24
2.3 Previous study on methanol production.....	24-25
2.4 Pectin as a substrate	25-26
2.5 Pectin methylesterase (PME) fromextraction of lime peels.....	26-27
2.6 Response surface methodology (RSM).....	27-28
2.7 Screening using Fractional Factorial Design (FFD)	28-29
2.8 Factors used in the methanol enzymatic reaction	29
2.8.1 pH.....	29-30
2.8.2 Temperature.....	30
2.8.3 Agitation.....	30
2.8.4 Enzymatic reaction time.....	31
2.8.5 Concentration of NaCl.....	31

3	MATERIALS AND METHODS.....	32
3.1	Overview	32-33
3.2	Chemical and raw material.....	34
3.3	Extraction of PME from lime peels	34
3.4	Enzymatic reaction.....	35
3.5	Preliminary study	35
3.6	Design experiment.....	36-37
3.7	Analysis of methanol by using HPLC.....	37
4	RESULTS AND DISCUSSION.....	38
4.1	Introduction	38
4.2	Preliminary	38-41
4.3	ANOVA & Regression Analysis	42-44
4.4	Main effect analysis	45-46
4.5	Interaction between factors	47-49
4.6	Comparison of methanol production with other reseachers.....	50-51
5	CONCLUSION & RECCOMENDATION	52
5.1	Conclusion.....	52-53
5.2	Future work ressomendation	53
6	REFERENCES	54-58

LIST OF FIGURES

Figure 3.1: Flow chart process of the experiment	33
Figure 4.1: Graph of concentration of methanol versus pH	40
Figure 4.2: Graph of concentration of methanol versus temperature	40
Figure 4.3: Graph of concentration of methanol versus NaCl	41
Figure 4.4: Graph of concentration of methanol versus time	41
Figure 4.5: The percentage distribution of each main factor and their interaction.....	46
Figure 4.6: The interaction graph between factor A, pH and factor B, temperature	48
Figure 4.7: The interaction graph between factor A, pH and factor C, time	49
Figure 4.8: The interaction graph between factor A, pH and factor E, agitation	49

LIST OF TABLES

Table 3.1: Process variables and levels for FFD.....	36
Table 3.2: Table factor of design experiment for screening.....	37
Table 4.1: Data from ANOVA.....	43-44
Table 4.2: Table factors of design experiment for factorial analysis.....	44
Table 4.3: Comparison for methanol production from agricultural source.....	51

LIST OF ABBREVIATIONS

PME	Pectin methylesterase
FFD	Fractional factorial design
RSM	Response surface method
DM	Degree of methylation
HM	High-methoxyl
UMP	Universiti Malaysia Pahang
CO ₂	Carbon dioxide
NaCl	Sodium chloride
CO	Carbon monoxide
H ₂ O	Water
H ₂	Hydrogen
HPLC	High Performance Liquid Chromatography
U.S.	United States
RPM	Rotational per minute
mL	Millilitre
M	Molar
w/v	Weight per volume
ANOVA	Analysis of Variance
NaOH	Sodium hydroxide
HCl	Hydrochloride acid
°C	Degree Celcius
Rpm	Revolution per minute
%	Percentage

1. INTRODUCTION

1.1 *Motivation and statement of problem*

Methanol was the simplest alcohol that contained only one carbon atom (CH_3OH). It was known as wood alcohol. Methanol was a colourless liquid, volatile, flammable and poisonous. It can be produced from different feedstock resources, predominately from natural gas and coal. Methanol was used in several industries such as in plastic industry. It also acts as an original feedstock to make polymer. Methanol also had been proved to be as fuel. When it was blended with gasoline in internal combustion engines, 85% of it was methanol and other 15% was gasoline. It also can be produced as pure methanol. Besides that, methanol also served as a raw material to produced chemical products for example formaldehyde, acetic acid, polymers, paints, adhesives, construction material and synthetic chemicals. (Luzia *et al.*, 2011)

Pectin methylesterase (PME) involved in de-esterification of pectin that released methanol and acidic acid. It is a heterogeneous group of enzyme complex that involved in pectin hydrolysis and composed of pectin esterase. The applications of PME were in food industry, textile, wines, pulp and paper industry (Sameer *et al.*, 2013). This enzyme catalysed the hydrolysis of the methyl ester group from pectin. It found in plants and also in pathogenic fungi and bacteria (Gayen S., & Ghosh U., 2011). PME had been purified and characterised from several species of citrus fruits such as orange and lemon (John & Tove, 2002). According to John & Tove (2002), citrus fruits were commercially used for juice extraction. Because of the high content in pectin, it also been used in production of methanol.

In this study, PME were be extracted from the citrus fruit that was lime peels waste. Lime peels were being mixed with pectin solution for enzymatic reaction process. Based on the production of methanol, the best factors were being investigated further in order to increase methanol production. The factors that were varied are temperature, pH, fermentation time, concentration of NaCl and agitation.

Fractional analysis by using two levels Fractional Factorial Designs (FFD) was studied in this research. Two level Fractional Factorial Designs (FFD) are popular experimental design and commonly used in engineering analysis (Don, 2013). In addition, FFD allows possible consideration of multi factors and it can determine the most relevant factors from all of the outcomes. Recently, FFD was the analysis that used to investigate the effect of tested independent variables to the response within the investigation range (Khalil *et al.*, 2011). Hence, FFD was a technique to determine the influence of several variables on the response and also estimating the overall main factor effects and interaction of different factors (Golshani *et al.*, 2013). According to Jawad *et al.*, (2013), FFD sign had been used to study the effect of independent variables and the level of selected factors that been chosen for preliminary experiments.

Usually, methanol was produced from synthesis gas where the main gas used is carbon monoxide and hydrogen. In order to produce methanol in typical plant, natural gas feedstock had to convert into a synthesis gas stream that consist of CO, CO₂, H₂O and H₂. It was usually accomplished by the catalytic reforming of feed gas and stream. Hence, by this synthesis gas method, the methanol produced was highly exothermic and taking place over a catalyst bed at moderate temperature. This condition will generate more energy of electricity and this were increased the capital cost. In this research study, a lime peels were used as the medium to release PME where PME was needed in this research for the production of methanol. They were chosen because it was agricultural or agro-industrial wastes that were abundant, renewable and inexpensive energy source that available in Malaysia. Hence, by PME that extracted from lime peels waste, it provides cost effective and eco-friendly method for the production of methanol on large scale (Patil & Chaundhari, 2010).

1.2 Objectives

The following were the objectives of this research:

- To produce methanol by using pectin methylesterase (PME) from lime peels waste and pectin solution as substrate
- To analyse the factor that affecting the methanol production from pectin methylesterase (PME)

1.3 Scope of this research

This research study was about the biological production of methanol by using pectin methylesterase (PME). Firstly, substrate which is pectin solution was prepared and the PME was extracted from lime peels waste. The fruits were cut into halves and take out the peels. Immediately the peels were treated to extract the pectin enzyme. Then, the preliminary studies were started by varying the factors. Production of methanol based on enzymatic reaction process was analysed by using High Performance Liquid Chromatography (HPLC). Hence, both objectives were completed. In this research study, it was interested to apply optimization tools like Response Surface Methodology (RSM) to improve the production of methanol via Fractional Factorial Design (FFD). FFD can provide clear understanding about the interactions involved between the process variables during the production process (Salleh, H. *et al.*, 2011). The screening process and analysing factors by using FFD that was taking into account were temperature, pH, enzymatic reaction time, agitation and concentration of NaCl.

1.4 Main contribution of this work

The following were the contributions of this work. Through this research, it can enrich the knowledge about the production of methanol by using pectin methylesterase (PME) as the raw material. To choose and used pectin methylesterase (PME) as the raw material in the production of methanol, it was the best method since it used agricultural waste as the raw material that was peels from lime where it were low cost and eco-friendly. It also can reduce the exponential increased of greenhouse effect by the polluting action of the industrial and transport sector (Luzia *et al.*, 2011).

1.5 Organisation of this thesis

The structure of the reminder of the thesis is outlined as follow:

Chapter 2 provides a description of the applications and general briefing about the raw material which is pectin methylesterase (PME) and the production. A general description on characteristics of the methanol, as well as the factors that was used in this research study as the parameter to run the experiment in preliminary study was also reported. This chapter also provides a brief discussion of the pectin solution and pectin methylesterase (PME) as the enzyme that were used in this study to produce methanol. A summary of the previous experimental work on the production of methanol by using agricultural waste was also presented. A brief discussion on the screening methods for methanol production and the analysis that was used during the screening was also provided.

Chapter 3 gives an overview of the material and methods that were used in this research. The flow diagram about the process of the experiment was also presented. This chapter also reported about the method of substrate preparation of pectin methylesterase (PME) which is from lime peels. The experimental design by using Response Surface Methodology (RSM) through Fractional Factorial Design (FFD) and the screening factors by using Design Expert software also discussed in this chapter. A brief discussion about the enzymatic reaction process, the preliminary study and also design experiment by five factors was also provided. The review about the analysis of the methanol by using High Performance Liquid Chromatography (HPLC) also presented in this chapter.

Chapter 4 was devoted to an enzymatic reaction process between pectin methylesterase (PME) from the lime peels waste as the raw material and pectin solution as a substrate to produce methanol. In this chapter, result and discussion for the experimental study were presented. There were results for preliminary study and results from Design Expert software that consist of ANOVA and regression analysis, main effect analysis, and interaction between factors. This chapter also discussed about comparison of methanol production with other researcher.

Chapter 5 draws together a summary of the thesis and recommendation which might be derived in this work.

2. LITERATURE REVIEW

2.1 Overview

According to Okonko *et al.*, (2009), technology enhancement and human development contributed to the continuous increased in the worldwide energy demand. Bhattacharyya *et al.*, (2008) stated there were three categories of energy sources that were fossil fuel, renewable and nuclear energy. The examples of fossil fuels were coal, petroleum and natural gas that were non-renewable energy sources that will be depleted in the next few years. The renewable energy sources included solar, wind, hydroelectric, biomass and geothermal energy whereas nuclear energy was derived from fission and fusion reactions (Gullu & Demirbas, 2001). Fossil fuel sources depletion had increased the need to reduce the consumption of fossil fuels. However, the depletion was not the only current concerned with fossil fuel use but the environmental degradation is. The burning of fossil fuels and the waste products that produced from it had created an imbalance in the atmospheric carbon dioxide (CO₂) levels, which had become the major contributor to global warming. While the municipal solid wastes from human and animal activities had also contributed to the environmental degradation. Therefore, it had been suggested that this waste should be recycled or converted into energy (Mastro & Mistretta, 2004).

Biomass was considered to be the renewable energy source with the highest potential to contribute to the energy needs of modern society for both developed and developing economics worldwide (Bridgewater, 2003). According to the 2001 report by the International Energy Agency (IEA), biomass currently contributed about 10.8 % of the world energy supplies including waste while other regenerative energy sources such as hydropower, wind, geothermal and solar were contributed about 3% only (Corradetti & Desideri, 2007). Hall *et al.*, (1992) supported that biomass currently supplied the highest proportion of regenerative energy among all regenerative resources with the percentage contribution of biomass as energy sources more than 35% in many developing countries such as tropical Africa.

In 2013, approximately 6.2 billion tons of carbon was emitted into the atmosphere as CO₂ and approximately 40% of this was emitted during the production of electricity. A survey from the U.S. Department of Energy revealed that the consumption of electricity increased significantly every year by 44% from 2006 to 2030 (Achmad *et al.*, 2011). Leduc *et al.*, (2011) stated that by 2050, road transportation was expected to be the largest contributor to greenhouse emission. In Europe, the renewable energy target for 2010 was approximately 5.75% of the transport fuels sold, and this target were likely increased to 10% in 2020. If this trend continues increased, the renewable energy target for the transport fuels sold should reached 27% by 2025.

The worldwide demand of methanol in 2013 was expected to reach 65 million metric tons which driven in a large part by the resurgence of the global housing market and this demand was increased because of the cleaner energy (Methanol Institute). In addition, according to the research from Methanol Institute, the methanol industry will spans the entire world with the worldwide production for Asia, North, South America, Europe, Africa and Middle East was about 33 billion gallons. It also reported that each day more than 100, 000 tons of methanols were used as a chemical feedstock or transportation fuel. They also stated that methanol was a truly global commodity where each day there is more than 80,000 metric tons of methanol will ship from one continent to another. This was due to the high demand of methanol that makes methanol production increased from year to year. In addition, the total cost of methanol production by using biomass was cheaper than the cost of methanol production by natural gas. Thus, biomass processing was the most cost-effective processes that had been developed for the production of methanol from renewable source (Shamsul *et al.*, 2014).

Lastly, the rapid growth of chemical technologies and industries that contributed to air and environmental pollution required some limitations to prevent the excessive emission of carbon dioxide into the atmosphere. These problems was leads some researcher to studies about the production of methanol from biomass.

2.2 *Methanol*

Methanol CH_3OH was a group of alcohol that widely used chemical as a common solvent in organic synthesis. It was the simplest organic liquid hydrogen carrier that acts as a hydrogen storage compound. Methanol also was a liquid transportation fuel that can be produced from fossil or renewable domestic resources. It was an attractive automotive fuel because of its physical and chemical characteristics. In United States, it was the most commonly used as a chemical feedstock, extractant, or solvent for producing methyl tertiary butyl ether (MTBE), an octane-enhancing gasoline additive. It also can be used in net which is 100% pure form as a gasoline substitute, or in gasoline blends. Methanol that produced from biomass had the most potential as a biofuel for power generation because it is distributed form of energy production. Fuel-grade methanol was a clean and efficient alternative fuel that can be used in power industry application for gas turbines. While for the transport sector, methanol acts as a superior to gasoline because it can burns at low temperature. Hence, because of the low volatility, methanol reduced the risk of an explosion or flash fire. The fires can be easily extinguished with water because its characteristics that less flammable than gasoline. This makes methanol more advantageous than hydrogen due to the problems associated with hydrogen storage (Shamsul *et al.*, 2014).

Demirbas, A. *et al.*, (2011) stated that methanol can be used as one possible replacement for conventional gasoline and diesel fuel. It was a promised renewable fuel that had lower carbon emissions compared to conventional fuel. The used of methanol could also reduce carbon emissions by motor vehicles by up to 81% and up to 32% for the carbon emission if the methanol were used to generate electricity. Methanol was industrially important chemical that acts as a raw material used in many chemical processes (Trop *et al.*, 2014). It also mainly used as a feedstock during the production of bulk chemical for example acetic acid and aldehyde (Anita, 2014). According to Hamelinck, (2002), methanol that produced from biomass was a promising carbon neutral fuel. This was because methanol that produced were clean and emitting none of the air pollutant that were SO_x , NO_x , VOS or dust. Form the research also stated that methanol that were derived from the grown biomass can be greenhouse gas neutral from the overall energy chain. This statement was supported by Kumabe *et al.*, (2008) that the

fossil fuel emission during fuel processing had prompted the search for renewable sources that emit zero or low pollution. The use of bio-methanol from biomass was more advantageous than fossil products because of its low pollution emission and raw material availability. Hence, biomass was a renewable energy source that can potentially replace fossil fuels because of the characteristics of this alcohol were identical to those fossil fuels.

The National Renewable Energy Laboratory from U.S. Department of Energy reported that in U.S. industry, it produced approximately 4.7 billion litres of methanol annually. About 38% of these methanols were used in the transportation sector, mostly in the production of MTBE. Most of the methanol produced in the United States today was made from natural gas but methanol also can be produced from other feedstock including coal, biomass and residual oil. Cost-effective, efficient, and environmentally sound processes for producing methanol from biomass make it being pursued by both government and industry research. While in China, since methanol was introduced in 2008, the amount of E85 used in China had included the blending of more than 1 billion US gallons of methanol into fuel (Shamsul *et al.*, 2014). This reported showed that methanol was one of the biofuel that had a potential market in the world in future especially when it produced from renewable energy that was biomass.

2.3 *Previous study on methanol production*

Since a few decades ago, there were many research studies about production of methanol. There had many researchers already doing their research about production of methanol especially by using biogas, natural gas and coal via gasification process also methanol production from CO₂. Nowadays there also had many previous studies about production of methanol by using raw material that do not harm environment and can save a lot of cost which was biomass. Trop *et al.*, (2014) was done the research study about producing methanol from a mixture of torrefied biomass and coal where this method was the so-called torrefaction of biomass. Torrefaction was also known as mild pyrolysis where the process was exposed to 200°C – 300°C within anaerobic environment where this method were used high temperature. Hamelinck & Faaij (2006) reported about previous work on methanol by

syngas process that includes CO and hydrogen. By this process, syngas is converted into methanol by a catalytic process based on copper oxide, zinc oxide or chromium oxide catalyst. In this process, distillation was used to remove water generated during methanol synthesis. Hence, previous research about production of methanol also was done by using natural gas. This production process can be replaced by biogas because biogas contains a large share of CO₂ compared to natural gas. Production of methanol by natural gas process was used waste anaerobic digestion (Kralj & Kralj, 2009). Bhattacharyya *et al.*, (2008) reported that methanol was also produced from the breakdown of methyl esters or the combination of ether with the methoxyl groups of uranic acid that was produced by the decomposition of pectin-like material plant. Xu *et al.*, (2011) had presented a novel approach for high efficient conversion of the CO₂-rich bio-syngas into the CO-rich bio-syngas carried out using biomass char and nickel catalyst, that successfully applied for production of bio methanol by using bio oil. Other than that, there are many researches already produced methanol by using biomass from agricultural waste as a raw material such as sugarcane bagasse, rice straw, rice husk, rice bran, wheat straw and etc. which had high potential in the future.

2.4 Pectin as a substrate

Within numerous enzymes, pectin was one of the best enzymes that can be used in methanol production. Pectin was one of the categories of complex group of associated natural polysaccharides from the primary cell walls and an intracellular region that found in higher plants. It was an important compound for food and pharmaceutical industries in drug delivery systems. The importance of the compound was related to its unique properties and the fact that it was biodegradable. The main raw materials from which commercial pectin was extracted from agricultural by-products, which were citrus peel and apple pomace (Lucyna, L. *et al.*, 2013). Wilkins *et al.*, (2007) stated that almost half of these citrus fruit was squeezed to juice, and the other remainder including peel, segments membranes and other by-product were considered as citrus waste. These citrus wastes can be dried and used as a raw material for pectin extraction and can be used as the substrate for methanol production (Mamma *et al.*, 2008).

According to Wang *et al.*, (2002), the dominant component of pectin were galacturonic acid with neutral sugars primarily galactose, arabinose, rhamnose, and xylose. Pectin was used as ingredients in various fields, including medicinal, pharmaceutical, cosmetics and food industries, for its gelling, stiffening and stabilizing properties. It was abundant in fruit and vegetable. Examples of fruits that had high in pectin were citrus, grape, and plum, peach and apple. According to Stephen (1995), the dominant feature of pectin was a linear chain of α -(1, 4)-linked D-galacturonic acid units where varying proportions of the acid group was present as methoxyl (methyl) esters. Generally, galacturonic acid units compose more than 65% of pectin structure where this important structure was the esterification of galacturonic acid residues with production of methanol.

2.5 Pectin methylesterase (PME) from extraction of lime peels

Pectin methylesterase (PME) was the first enzyme that can acting on pectin that was a major component of plant cell wall which it catalyses the de-methylesterification of galacturonic acid units of pectin and generating free carboxyl groups and releasing proton which methanol (Giovane, A. *et al.*, 2004). PME removed methyl groups from pectic component of cell wall during fruit ripening which then can be depolymerised by polygalacturonase and this process will decreased the intercellular adhesively and tissues rigidity (Assis *et al.*, 2001). Mustapha, N. *et al.*, (2011) stated that de-esterifying enzyme pectin methylesterase (PME) was catalysed the released of methanol and depolymerising enzymes belonging to subclasses that was polygalacturonases. It hydrolysed α -(1, 4) – glycosidic bonds between two non-esterified galacturonic acid residue. These enzymes will act on pectin that acts as a substrate whose degree of esterification was below 55 – 60%. The percentage of carboxyl group that esterified with methanol was called as a degree of methylation (DM). If the DM was more than 50% higher, the pectin was called as high-methoxyl (HM) pectin. But if the DM was less than 50% the pectin was called as low-methoxyl (LM) pectin. Pectin can be de-esterified to produce methanol as it is been abundant in methoxyl side groups. De-esterification of pectin was carried out by a chemical method that was hydrolysatation catalysed by using enzyme. The enzyme that been used in de-esterification of pectin was called pectin methylesterase (PME). Pectin methylesterase (PME) was involved in catalysed the de-

esterification of pectin. It was produced carboxyl group as well as methanol as products. It also presented in all higher plants but commonly pectin methylesterase (PME) was abundant in citrus fruits (ŞİMŞEK & YEMENİCİOĞLU, 2010).

Pectin methylesterase (PME) also catalysed the hydrolysis of the methyl ester group from pectin. The fast and high specific catalysis of pectin methylesterase (PME) make them the economically alternative for hydrolysis of pectin. Pectin methylesterase (PME) was significant to the citrus industry due to the establishment of PME as a causative agent for juice clarification and gelatine of frozen concentrates (Gayen & Ghosh, 2011). According to Nielsen & Christensen (2002), pectin methylesterase (PME) was purified from several fruit that come from citrus family such as lemon, oranges and lime. It was proved that citrus fruits were commercially used in extraction of juices because of the high content of pectin in the fruit. Therefore, detection of a large quantity of pectin in a fruit alone is not in itself enough to qualified that fruit had a higher pectin where pectin were almost exclusively found from citrus peel or apple pomace where both by-product is from juice manufacturing (Thakur *et al.*, 1997). Hence, citrus peel contains about 20-30% of pectin that make it suitable to act as enzyme and de-esterified with pectin to produce methanol (May, 1990).

2.6 Response surface methodology (RSM)

Response surface methodology (RSM) was the most popular optimisation method used in recent years. It was developed by Box and collaborators in the 50s century. There were so many works based on the application of RSM in chemical and biochemical process. One of the optimisation techniques in the application of RSM was the selection of independent variables of major effects on the experiment of methanol production through screening studies (Bezerra, M. A. *et al.*, 2008). The purpose of RSM was to obtain a predicted model and this model can be useful for screening of enzymatic reaction process condition for methanol production. Therefore, in this research study, RSM was used as a screening method in order to increase the yield of methanol production without increased the cost.

RSM was a collection of statistical and mathematical techniques useful for developing, improving and optimising processes in which a response of interest was influenced by several variables and the objective of this method was to screening this response. Hence, it also can be used to define the relationships between the response and the independent variables. RSM had important application in the design, development and formulation of new products, as well as in the improvement of existing product design. It defines the effect of the independent variables, alone or interaction in the process of methanol production. It was possible to separate an optimisation study using RSM into three stages. The first stage is the preliminary work in which the determination of the independent parameters and their levels are carried out. The second stage is the selection of the experimental design and the prediction and verification of the model equation. The last one is the response surface plot and contour plot of the response as a function of the independent parameters and determination of optimum points (Boyaci, H. I. & Bas, D., 2007).

2.7 Screening using Fractional Factorial Design (FFD)

Don (2013) stated that Fractional Factorial Design (FFD) was a popular experimental design method that used for two levels. It was one of the most frequently method that applied fractional design in engineering field. This method made possible to consider multitudinous factors and can identify the most important and relevant factors from the long list of analysis during the enzymatic reaction of methanol production. Khalil *et al.*, (2011) stated that recently, FFD was the method in the analysis that used to investigate the effect of tested independent variables to the response within the investigation range during the experimental analysis. Hence, it was also a technique to determine the influenced of several variables on the response and also to estimating the overall main factor effects and interaction of different factors in methanol production (Golshani *et al.*, 2013). According to Jawad *et al.*, (2013), FFD sign has been used to study the effect of independent variables and the level of selected factors that been chosen for preliminary experiments during methanol production. Xie *et al.*, (2003) stated that Factorial Design was a closed-ended system for process optimisation where in this method, level of factors or parameters were independently varied, each factor at two or more levels. This effects that can be attributed to the factors and their interactions were assessed with maximum efficiency for methanol production in factorial design. Moreover, it allow for the

estimation of the effects of each factor and interaction. The most commonly used fractional factorials in medium improvement experiments were two factorial designs (denoted by 2^n when there are n factors). These designs were the smallest capable of providing detailed information on factor interaction.

2.8 Factors used in methanol enzymatic reaction

During this research study, there must be a few factors that will contribute in order to make sure the methanol can be produced at the end of this research. Hence, in this research study five factors were figured out which these factors can affect the production of methanol. All five factors that had been measured to produce methanol were pH, temperature, agitation, enzymatic reaction time and concentration of NaCl. These factors were the medium that had been used in order to see the successfully of the methanol production in this research study.

2.8.1 pH

pH was a measured of the concentration of hydrogen ions in the solution. The higher the hydrogen ion concentration the lower the pH. Most enzymes function efficiently over a narrow pH ranges. A change in pH above or below this ranges will reduces the rate of enzymatic reaction of that enzyme. Kurita *et al.*, (2008) stated that the PME activity increased with the increasing of the pH and the PME activity was higher at neutral pH. The viscosity of the PME was increased gradually from pH 5.8 to pH 7.0 but not after pH 7.6 where after the pH 7.6, the enzyme begins to be denatured. Enzyme denatured was known as the enzyme that start loses their functional shape, particularly the shape of the active site, such that the substrate was no longer fit to it. Hence, according to Carbonell *et al.*, (2006), optimum pH where the most favourable pH value that the enzyme was most active for the PME activity for all assayed cultivars was at pH 7.8. But according to Rodriguez-Lopez *et al.*, (2013), the highest PME activity was obtained at highest pH that is pH 8.0. As regards of the de-esterification of pectin, high pH would exert a greater influence on the pectin structure and this condition let the pectin become less sensitive to β -elimination that affect the production of

methanol (Kurita *et al.*, 2008). Spagna *et al.*, (2003) reported that optimum pH for the PME extraction was at pH 7. Hence, Amaral *et al.*, (2005) and Arotupin *et al.*, (2008) was reported in their research that these pH range also affect the PME activity in the production of methanol. In general, PME was found to have an optimum pH ranging from 7.5 to 9.0.

2.8.2 Temperature

Like most chemical reactions, the rate of an enzyme reaction increase as the temperature was raised. In this research study, temperature was one of the important factors that affect the production of methanol. According to Carbonell *et al.*, (2006), Atkin & Rouse (1953) had studied about the PME inactivation at different temperatures and found out that PME enzyme was break near 70° C. This means that if the temperature was more than 70°C the enzyme were be denatured. Kurita *et al.*, (2008), study about the effect of temperature in extraction of pectin. Three different temperatures was used in that method which is 50°C, 65°C and 80°C and found that at temperature 50°C was produced high pectin during the extraction. All these statement was supported by Tijsskens *et al.*, (1999) and Amaral *et al.*, (2005) in their research that PME had high activity. PME at maximum rate was at the temperature between 50 and 60° C while the enzyme will lose its activity at the temperature higher that 70° C. Therefore, this statement showed that methanol was less produced or cannot be produced if the temperature higher than 70°C.

2.8.3 Agitation

In this research study, agitation was one of the factors that need to study because between with agitation and without agitation, there were given a different kind of result towards the PME activity and production of methanol. Although this factor is one of the factor that still new in the research but still had a few researcher that discussed it as one of the important factor that will affect the production of methanol. Garcia-Castello

et al., (2012) reported in previous research agitation speed at 175 rpm was the best condition of speed for the pectin methylesterase (PME) activity.

2.8.4 Enzymatic reaction time

Time was one of the factors that also important in this research study in order to produce methanol especially in enzymatic reaction. It was because time will showed at which point of time will be the highest production of enzyme to react with other solution to give the best result of production. According to Rodriguez-Lopez *et al.*, (2013), the extraction time for the PME activity to react and produced methanol was significant in the range 20 – 120 minutes. Hence, from the research the finding of the highest PME was reported at time 100 minutes where at this time, the highest methanol was produced. Garcia-Castello *et al.*, (2012) in the research study stated and fixed that the extraction time of the PME enzymes between 0.33 - 2 hours and between that ranges, the result that PME activity was highest at 90 minutes where it was a suitable time for the reaction with pectin and then produced a highest methanol.

2.8.5 Concentration of NaCl

In this research study, concentration of NaCl was used as an extracting solution in order to produced methanol because according to Contreras-Esquivel *et al.*, (1999), concentration of NaCl in the extracting solution showed a significant effect on the PME activity measured. This PME when react with pectin solution were produced methanol. The effect of NaCl concentration as extracting solution of PME activity from lime peels showed that the enzyme activity of PME was increased substantially in lime peels when NaCl solutions was used as extractant to extract PME from lime peels. PME from lime peels were extracted with NaCl solutions and in the production of methanol, different concentration between 0.5-3.0 M were used. PME activity was referring as the activity where carboxylic groups were released during the extracting process (Rodriguez-Lopez *et al.*, 2013).

3. MATERIALS AND METHODS

3.1 Overview

This chapter was discussed about the material and methods conducted in the experimental work. This chapter explained the enzymatic reaction process and also the analysis of the methanol production. The subchapter from this chapter was covered about preparation raw material, extraction of PME, enzymatic reaction, preliminary study, design experiment, analysis of methanol by using High Performance Liquid Chromatography (HPLC) and also summary about all the method. These methodologies were being used thoroughly in this study of research in order to get analysis of the methanol production. Figure 3.1 showed the flow of the process to conduct the experimental work.

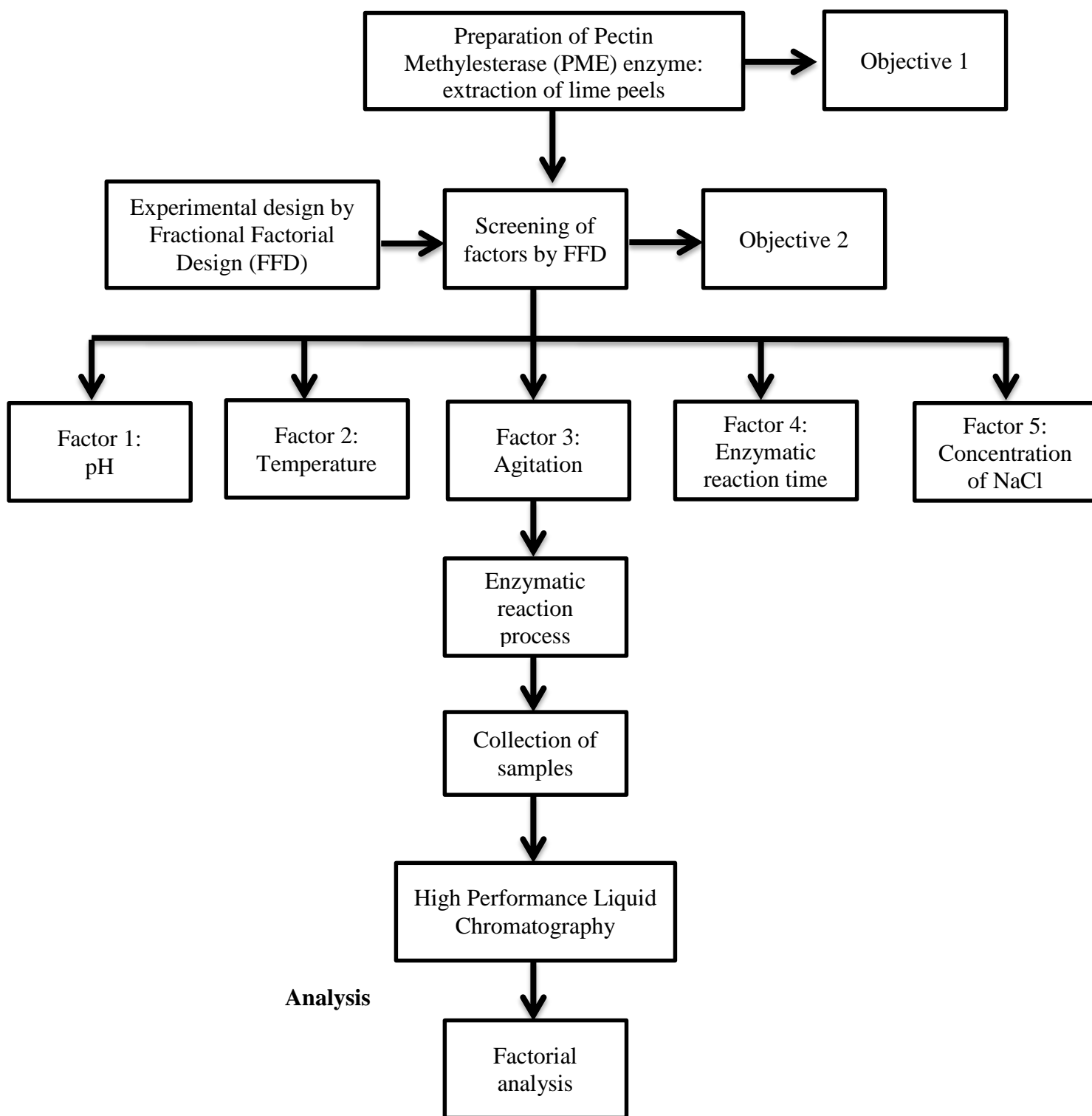


Figure 3.1: Flow chart process of the experiment

3.2 *Chemical and raw material*

The raw material that was used in this research was pectin methylesterase (PME) that was extracted from lime peels. Lime was bought at the nearer supermarket in Gambang, Kuantan, Pahang Malaysia. It was used to obtain PME in the production of methanol. After that, lime peels were washed carefully with tap water and stored in the refrigerator under 4°C to ensure the lime freshness until to be used for the experiment (Rodriguez-Lopez *et al.*, 2013). During the experiment, the limes were cut into halves and take out the peels. After that the peels were immediately extract with NaCl solution by using domestic blender for 1 min. The solutions of lime peels were filtered before it can use for next process (Contreras-Esquivel *et al.*, 1999). Hence, for the chemical, there were five types of chemical used in this research study. In analysing the production of methanol by using HPLC, acetonitrile was used as the mobile phase with the ratio concentration of 55% CAN: 45% H₂O (Salleh, H. *et al.*, 2011). While for the control adjustment of pH, sodium hydroxide (NaOH) and hydrochloric acid (HCl) were used. Sodium chloride (NaCl) was used in the extraction of pectin methylesterase (PME) from lime peels while methanol pure was used during analysis by using HPLC in order to get the standard curve. The standard curve was used as to calculate the production yield of methanol.

3.3 *Extraction of PME from lime peels*

The lime peels were suspended in a 1.0M of NaCl solution where the ratio of the peels to the extractant of concentration of NaCl was 1:4 (w/w). Then the peels that already mixed with 1.0 M NaCl solution were homogenised quickly for 1 min in a blender. The homogenates of peels were filtered with a filter paper and centrifuged at 5800 rpm for 10 minutes. 10 mL of supernatant was put in the shake flask and mixed with 90 mL of pectin solution that acts as a substrate to get 100 mL of mixture solution. The pectin solution were prepared by mixed (1% w/w) of pectin with 0.3 M of NaCl solution (Contreras-Esquivel *et al.*, 1999).

3.4 Enzymatic reaction

For the methanol production, PME was mixed with pectin solution in the shake flask and undergo enzymatic reaction process based on the factors that were chosen. Then it was incubated in the incubator shaker (SI500) for 24 hours. The factors that were manipulated during enzymatic reaction process were temperature, pH, agitation, enzymatic reaction time and also the concentration of NaCl. After that, the results was analysed by using High Performance Liquid Chromatography (HPLC).

3.5 Preliminary study

The preliminary study was conducted in order to determine the best condition between the factors that has been selected during this research. This preliminary study also was done to examine the best experimental factors and narrow the corresponding ranges that had been used in the experiment. For preliminary experiment, the factors that were used were pH, agitation, temperature, enzymatic reaction time and also concentration of NaCl. After the preliminary experiment, the ranges for every factor were chosen and proceed with the screening study. In this research study, one factor at a time was used during preliminary experiment. The range for every factor was showed in Table 3.1. The range for pH was between 3 to 8 while for temperature the range was between 20 to 70° C. For enzymatic reaction time, the range had been set was between 30 minutes until 12 hours. The range for concentration of NaCl was between 0.5 M to 3 M and for agitation it had been set between with agitation and without agitation. Hence, from Table 3.1 also, it showed the process variables and levels for the factor that had been chosen for the FFD.

3.6 Design experiment

The experiment for this research study was done in the analytical laboratory in the chemical engineering and chemical resources laboratory. The screening study was done by using Response Surface Methodology (RSM) through Fractional Factorial Design (FFD) using Design Expert software (Version 8.0.6, State-Ease). This study was done to screen five factors that were studied in the methanol production during preliminary study. Five different factors tested were temperature, pH, enzymatic reaction time, agitation and concentration of NaCl. The levels of selected factors were stated according to preliminary study as shown in Table 3.1. From Table 3.2, value response had been determined and the data was analysed by using Design Expert to screen the highest production for methanol.

Table 3.1: Process variables and levels for FFD

Process variables	Unit	Low	High
Temperature	Degree	20	70
pH		3	8
Enzymatic reaction time	Hours	0.5	12
Agitation	Rpm	No	Yes
Concentration of NaCl	Molar	0.5	3.0

Table 3.2: Table factors of design experiment for screening.

Run	Factor 1: temperature degree	Factor 2: pH	Factor 3: time hour	Factor 4: concentration NaCl; M	Factor 5: agitation rpm
3	25	5	1	1	Yes
18	25	5	1	5	No
8	25	9	1	5	Yes
9	25	9	1	1	No
7	25	5	6	1	No
12	25	5	6	5	Yes
13	25	9	6	1	Yes
17	25	9	6	5	No
5	47.5	7	3.5	3	Yes
6	47.5	7	3.5	3	No
1	70	9	6	1	No
2	70	9	1	5	No
14	70	9	1	1	Yes
15	70	9	6	5	Yes
4	70	5	1	1	Yes
11	70	5	1	1	No
10	70	5	6	1	Yes
16	70	5	6	5	No

3.7 Analysis of methanol by using HPLC

After completed the enzymatic reaction process, the samples were collected for further analysis in order to determine the content of methanol that has been produced. Firstly, analysis was run by using High Liquid Performance Chromatography (HPLC). The mobile phase that was used during the analysis was acetone nitrile. HPLC was a common technique to quantify methanol production. According to Voragen *et al.*, (1986), to determine the degree of methylation and acetylation of pectin, first was to analyse the supernatant by HPLC after the pectin was saponification and precipitated. This has been proved by Kuo *et al.*, (2002) where the analysis of methanol in Chinese liquor medicine by HPLC has been reported by them. Then, Fractional Factorial Design (FFD) method was used to identify the most important and relevant factors from the analysis.

4. RESULT & DISCUSSION

4.1 Introduction

This chapter discusses the outcome of this research study that related to the objectives and scopes. The topics that covered in this chapter were ANOVA and regression analysis, main effect analysis, interaction between factors and comparison of methanol production with other researchers. In this research work, the analysis that was used to measure the concentration of methanol content was High Performance Liquid Chromatography (HPLC). The recent study was conducted to investigate the production of methanol using pectin methylesterase (PME) from lime peels waste and pectin solution as a substrate. From the research work, it showed that all the factors used gave different effect to the methanol production. The interaction between all the factors was analysed by using Design Expert software.

4.2 Preliminary

Preliminary study was done to determine the best condition to conduct further experiment. This preliminary study was done by using mixed of pectin solution and lime peels waste. From the preliminary the highest concentration of methanol was at pH 7 while for the factor temperature, 70°C recorded the highest concentration of methanol. From the analysis, 3 M NaCl produces the highest concentration of methanol while for the factor of enzymatic reaction time, at 5 hours produced the highest concentration of methanol followed by at 3 hours. Hence, from this preliminary result, the range for the factors for screening experiment can be decided by using Design Expert software.

During the experiment for preliminary study, a total 18 run was done with different factor and from the result the ranges for every factor was chosen to proceed the experiment with screening. The result for the preliminary study was showed in the graphs below. From Figure 4.1, the graph showed that the concentrations of methanol were increased at pH 3, pH 4, pH 6 and pH 7. Hence from these values, the ranges for the screening were between pH 5-9. The higher concentration of methanol is at pH 7, this was because the PME present in plants was preferred to produce more at that pH 7. This statement was supported by Benen *et al*, 2003

and Jayani *et al*, 2015. They stated in their research that pectin methylesterase (PME) that produced from plant was the enzyme that had optimum pH ranges between 6 to 8. Gonzalez & Rosso (2011) in their research reported that the higher activity of pectin methylesterase was at pH 4 to 4.5.

Hence, from the Figure 4.2, the graph of concentration of methanol versus temperature only showed increase with the concentration at temperature 20°C, 30°C, 60°C and 70°C where the highest production was at 70°C. The ranges for the screening was set between 50 - 70°C while from Figure 4.3, the methanol only produced at 1.5 M, 2.5 M and 3 M of concentration of NaCl. The highest of the concentration of methanol was at 3 M of concentration and the range for the concentration of NaCl was set at 1 – 5 M. From Figure 4.2, the graph showed that temperature was the factor that produced higher concentration of methanol which is 57.669 g/l at temperature 70° C. Hence, from Figure 4.3, concentration of NaCl was the factor that produced least methanol that was 5.183 g/l at 3M.

Lastly, from Figure 4.4, it showed the graph of concentration of methanol versus enzymatic reaction time. From the graph, the methanol only produced during 2 hours, 3 hours and 5 hours of enzymatic reaction time. The highest production was at 5 hours of time. Hence, the ranges for the screening experiment were 1 to 6 hours of time. From the graph also, it showed the highest R² value was 0.9958. It makes enzymatic reaction time as one of the important factor that must be take place in the research. This was because reaction time was the factor that needed for the enzyme to reacts with the substrate that was pectin to produce methanol.

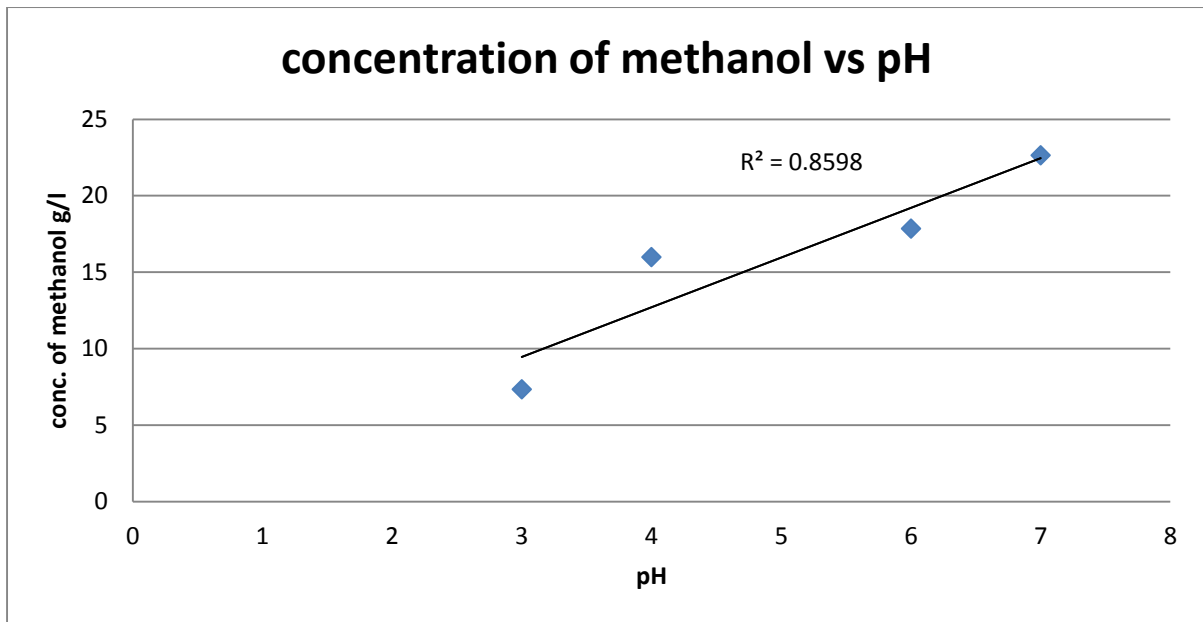


Figure 4.1: Graph of concentration of methanol versus pH

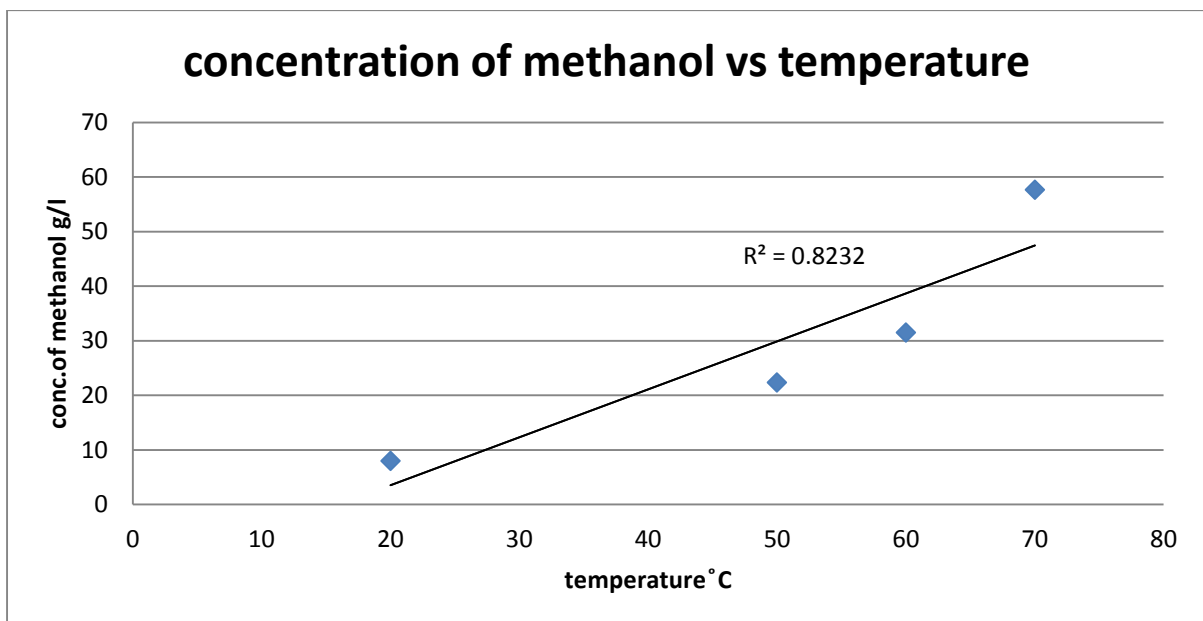


Figure 4.2: Graph of concentration of methanol versus temperature

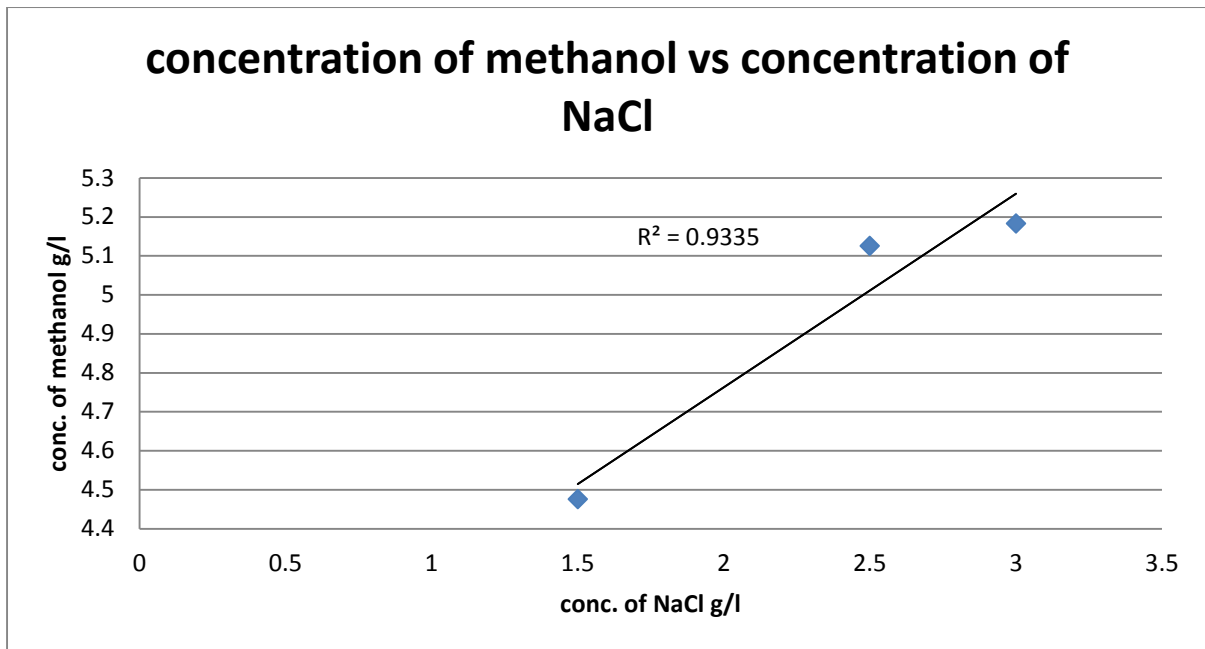


Figure 4.3: Graph of concentration of methanol versus concentration of NaCl

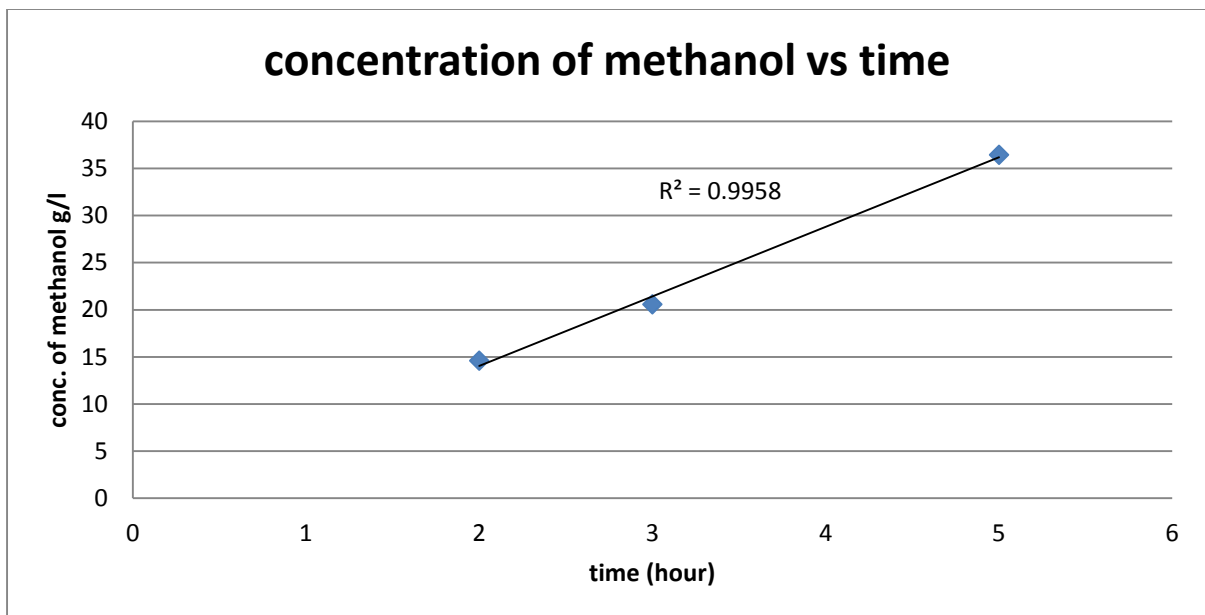


Figure 4.4: Graph of concentration of methanol versus time

4.3 ANOVA & Regression Analysis

In this research study, five factors were assumed to be influence on methanol production by enzymatic reaction of pectin solution and lime peels waste where all the factors were used as a response. The five factors that include were concentration of NaCl, pH, enzymatic reaction time, temperature and agitation. The results of 18 runs factorial design of methanol production were analysed by analysis of variance (ANOVA) and a regression model where it developed to describe the relationship between the selected factors. ANOVA, based on the definition was allowed one to examine whether the variability due to a particular experimental, or combination of factors, that was statistically significant compared to the measured variability due to random sources. Thus ANOVA can be used to examine differences between groups. Hence, ANOVA and its relative can accommodate many types of experimental designs (Pavlidis, P., 2003).

From this research study, ANOVA results showed that pH (A), temperature (B), enzymatic reaction time (C), concentration of NaCl (D) and agitation (E) affected the methanol production. The regression model for methanol production was expressed in the equation 1.

The quadratic equation of regression model for methanol production:

$$\begin{aligned} \text{Methanol} = & +0.65 + 0.025A + 0.049B + 0.019C + 0.22D + 0.054E - 0.12AB + \\ & 0.094AC + 0.061AD + 0.094AE + 0.053BD + 0.045BE - 0.064CD \end{aligned} \quad (1)$$

R^2 was a statistical measured to know of how close the data to the fitted regression line. It was also known as the coefficient of determination or the coefficient of multiple determinations for multiple regressions. Annuar *et al.*, (2008) stated that the R^2 value usually used to determine whether the model was correctly represented the data or not where if the value of the R^2 was near to one, so the regression model was correct. Therefore, from this analysis, the R^2 value for methanol production was 0.9904.

From the Table 4.1, the model showed that the F-value is 25.69 which implies that the model was significant for the research. From the F-value, there was only 1.08% chance that the model F-value was not significant where it was possibly because of the noise occurred. Hence, from the Table also, we can see whether the model was significant or not by looking at the value of $\text{prob} > F$. If the value of $\text{prob} > F$ was less than 0.05, then it showed that the model term was significant while if the value was greater than 0.1, then it indicates that the model terms were not significant. In this case, parameter A and interaction of AB, AC, AD, AE and CD were significant model term because of all their $\text{prob} > F$ was less than 0.05.

Hence, from the table factor of the design experiment for factorial analysis, it showed that the highest yield of methanol production was at pH 9 with temperature of 25°C and 5 M of NaCl with agitation and 1 hour of enzymatic reaction time. The value of the highest yield production of methanol was 2.241480 g/g while the lowest production yield of methanol produced were at pH 9 with 25°C temperature and 1 M of concentration of NaCl without agitation and 1 hour of enzymatic reaction time. The production yield was 0.283838 g/g.

Table 4.1: Data from ANOVA

Source	Sum of square	Df	Mean square	F value	P-value $\text{prob} > F$	
Model	1.56	12	0.13	25.69	0.0108	Significant
A- pH	9.793E ⁻³	1	9.793E ⁻³	1.93	0.2584	
B- temperature	0.039	1	0.039	7.74	in0.0688	
C- time	5.723E ⁻³	1	5.723E ⁻³	1.13	0.3656	
D- conc. of NaCl	0.76	1	0.76	149.94	0.0012	
E- agitation	0.047	1	0.047	9.23	0.0560	
AB	0.22	1	0.22	43.71	0.0070	
AC	0.14	1	0.14	28.15	0.0131	
AD	0.059	1	0.059	11.57	0.0424	
AE	0.14	1	0.14	28.11	0.0131	
BD	0.045	1	0.045	8.85	0.0588	

BE	0.032	1	0.032	6.28	0.0873	
CD	0.066	1	0.066	13.12	0.0362	
Curvature	0.098	2	0.049	9.65	0.0493	Significant
Residual	0.015	3	5.062E ⁻³			
Cor total	1.67	17				

Table 4.2: Table factors of design experiment for factorial analysis

Run	Factor 1 A: pH	Factor 2 B: temperature degree	Factor 3 C: time hour	Factor 4 D: concentration NaCl; M	Factor 5 E: agitation rpm	Yield of methanol g/g
9	9	25	1	1	No	0.283838
3	5	25	1	1	Yes	0.350148
1	9	70	6	1	No	0.564350
7	5	25	6	1	No	0.590826
12	5	25	6	5	Yes	0.874664
14	9	70	1	1	Yes	0.909808
10	5	70	6	1	Yes	1.253106
2	9	70	1	5	No	1.337556
16	5	70	6	5	No	1.355628
13	9	25	6	1	Yes	1.391876
11	5	70	1	1	No	1.528086
5	7	47.5	3.5	3	Yes	1.716906
6	7	47.5	3.5	3	No	1.770520
18	5	25	1	5	No	1.828090
17	9	25	6	5	No	2.003880
15	9	70	6	5	Yes	2.019740
4	5	70	1	5	Yes	2.180300
8	9	25	1	5	Yes	2.241480

4.4 Main effect analysis

From the Figure 4.5, it showed that factor D that was concentration of NaCl had been contributed the most in methanol production with the percentage contribution of 45.36%. According to Contreras-Esquivel *et al.*, (1999), concentration of NaCl in the extracting solution showed a significant effect on the PME activity measured. Rodriguez-Lopaz *et al.*, (2013) stated that the effect of NaCl concentration on the extracting solution of PME activity from lime peels showed that the enzyme activity of PME were increase substantially in lime peels when NaCl solutions were used as extractant to extract PME from the lime peels during the extracting process.

The second factor that contributed to the production of methanol was agitation with the percentage contribution of 2.79%. This research study was done with agitation and without agitation. However, this factor was something new to be discussed. However, Garcia-Costella *et al.*, (2012) reported about factor of agitation in previous research and stated that agitation speed at 175 rpm was the best condition for the highest pectin methylesterase (PME) activity.

Temperature contributed of 2.34% to methanol production. The reaction for enzymatic reaction was increased with increasing of temperature if the inactivation of enzyme is not considered (Wang, Q., 2006). Merrill and Weeks (1945) claimed that the viscosity of pectin solution decreased rapidly and irreversibly when the pectin solution was heated. This is because of the degradation of pectin.

Enzymatic reaction time contributed the least percentage to the methanol production with 0.34% followed by pH with 0.59% contribution. From the result it showed that both of the factors were not the most affecting factors in the methanol production. For a single effect factor, although time consuming and pH are usually important in enzymatic reaction, but for this process these two factors had showed to be less important compared to the other factors. However, enzymatic reaction time and pH contributed the most when interact.

	Term	Stdized Effects	Sum of Squares	% Contribution
	Intercept			
M	A-pH	0.049	9.793E-003	0.59
M	B-temp	0.099	0.039	2.34
M	C-time	-0.038	5.723E-003	0.34
M	D-conc.NaCl	0.44	0.76	45.36
M	E-agitation	0.11	0.047	2.79
M	AB	-0.24	0.22	13.22
M	AC	0.19	0.14	8.51
M	AD	0.12	0.059	3.50
M	AE	0.19	0.14	8.50
O	BC	-0.058	0.013	0.79
M	BD	-0.11	0.045	2.68
M	BE	0.089	0.032	1.90
M	CD	-0.13	0.066	3.97
O	CE	0.020	1.605E-003	0.096
O	DE	-9.176E-003	3.368E-004	0.020

Figure 4.5: The percentage distribution of each main factor and their interaction. Terms: A, pH; B, temperature; C, time; D, concentration of NaCl; E, agitation

4.5 Interaction between factors

Interaction effects were representing the combined effects of factors on the dependent measure. When interaction effects were presented, the impact of one factor depends on the level of the other factors. The interactions between the factors were likely improved the production of methanol. Some of the factors interaction may contribute more than the main factors alone. This showed that pectin methylesterase (PME) enzyme preferred to produce more during interaction rather than alone factor. There were three interaction discovered in this research study.

In Figure 4.6, the first interaction between factor A and factor B showed that there was an interaction that produced higher methanol yield between pH 5 and temperature at 70°C. But at the same temperature and different pH which was at pH 9, it produced lower methanol yield. Hence, in the same interaction but at temperature 25°C and at pH 5 it produced lower methanol while at pH 9 it produced high methanol. The production methanol was different because of the system had a different behaviour at different pH. From this interaction it showed that enzyme in this reaction preferred high temperature and acidic condition. This was because temperature was the factor that contributes the third highest in the production of methanol. Other than that, during preliminary study, temperature was the factor that showed highest production of methanol. Atkin & Rouse (1953) had stated in research study about PME inactivation and found that PME was broken at temperature more than 70°C. This statement means that highest production of methanol can be produced at not more than 70°C.

The second interaction can be seen in Figure 4.7. It showed that the interaction between factor A and factor C during 1 hour reaction time at pH 5 produced high methanol yield. However, the production of methanol was lower at pH 9 but at the same period of reaction time. During 6 hours of enzymatic reaction time at pH 5, the methanol production was the lowest compared to pH 9 at same enzymatic reaction time. Hence, the highest contribution of enzymatic reaction time for the production of methanol in this interaction was during 1 hour compared to 6 hour. According to Rodriguez-Lopez *et al.*, (2013), the extraction time for the PME activity was significant in the range 20 – 120 minutes. From this statement, it showed that the result is significant.

The third interaction was represented in Figure 4.8. It showed the interaction between factor E and factor A at pH 5 without agitation, the production of methanol was higher than at pH 9 without agitation. However, the production of methanol at pH 5 with agitation was lower than the production of methanol at pH 9 with agitation. The high contribution of agitation for methanol production had been explained by Garcia-Castello *et al.*, (2012). Agitation speed at 175 rpm was the best condition for pectin methylesterase (PME) activity to produce methanol. This factor was still something new to discuss especially the interaction with other factors.

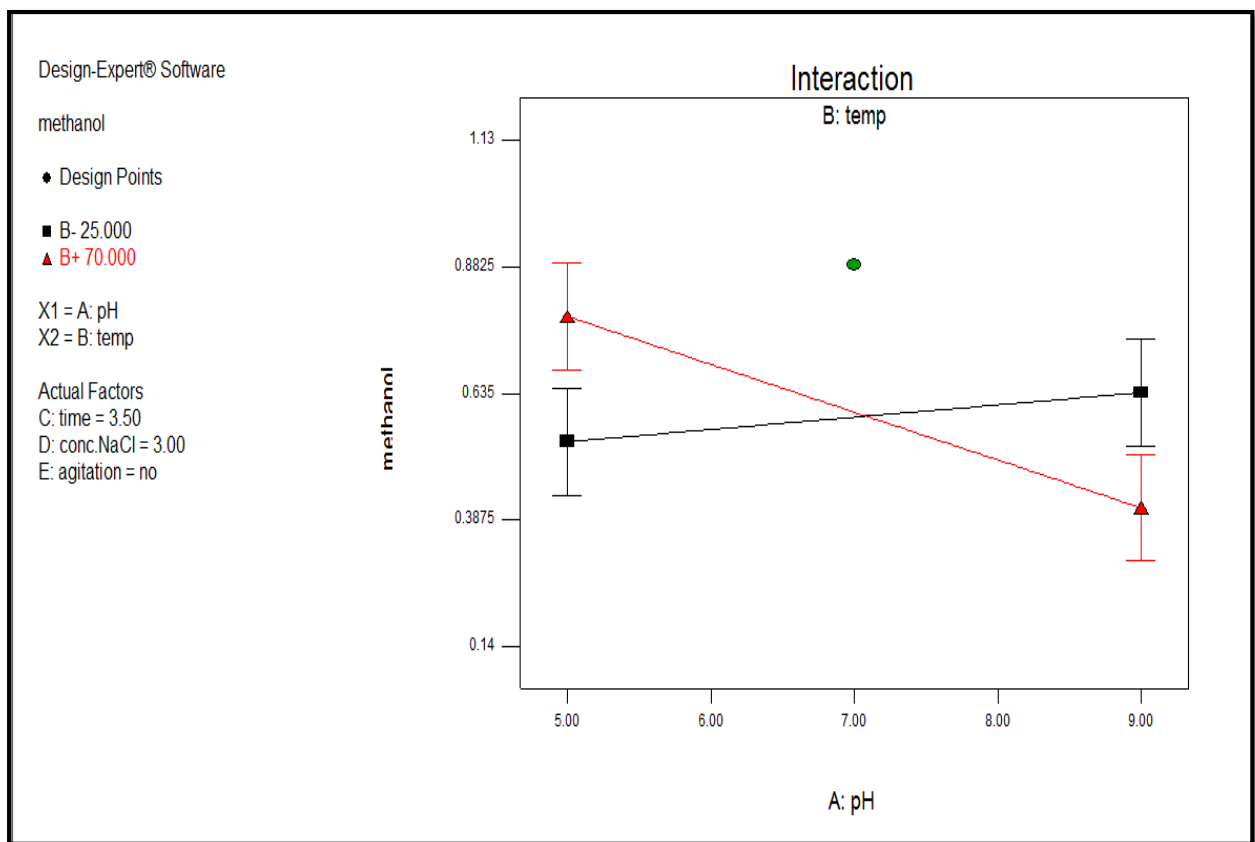


Figure 4.6: The interaction graph between factor A, pH and factor B, temperature

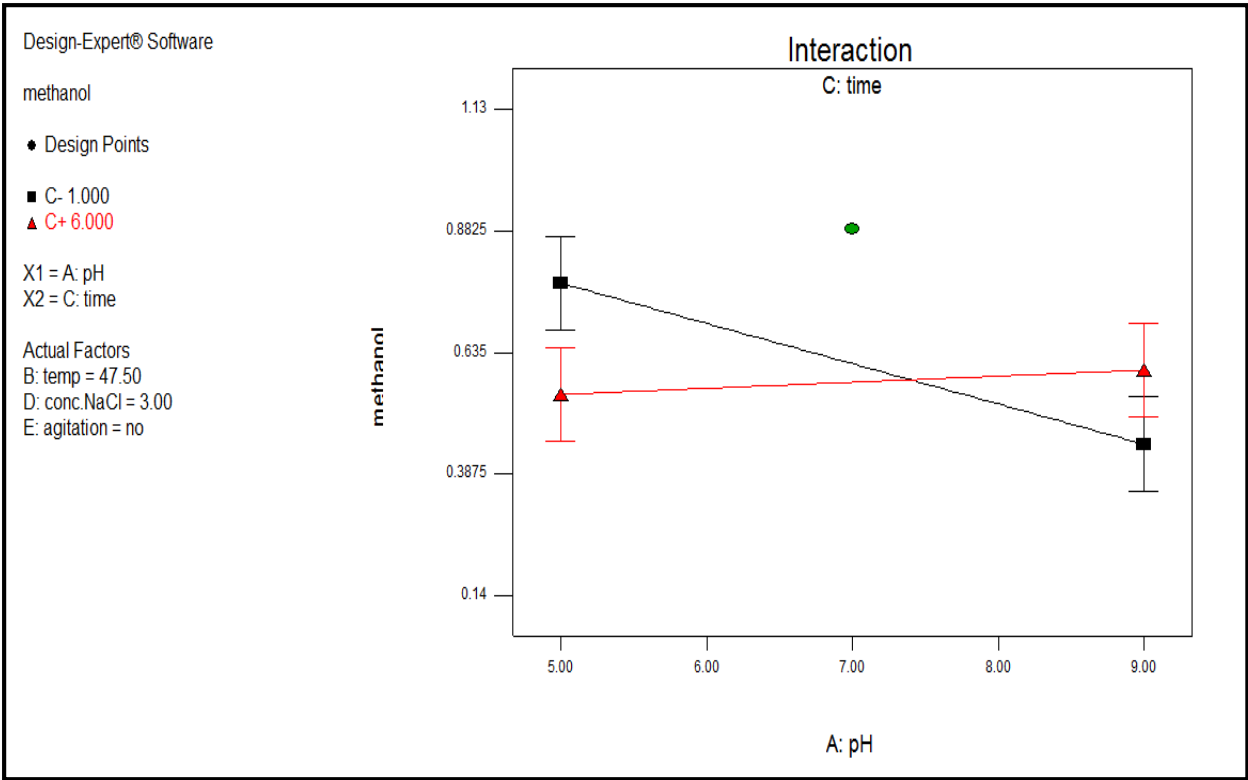


Figure 4.7: The interaction graph between factor A, pH and factor C, time

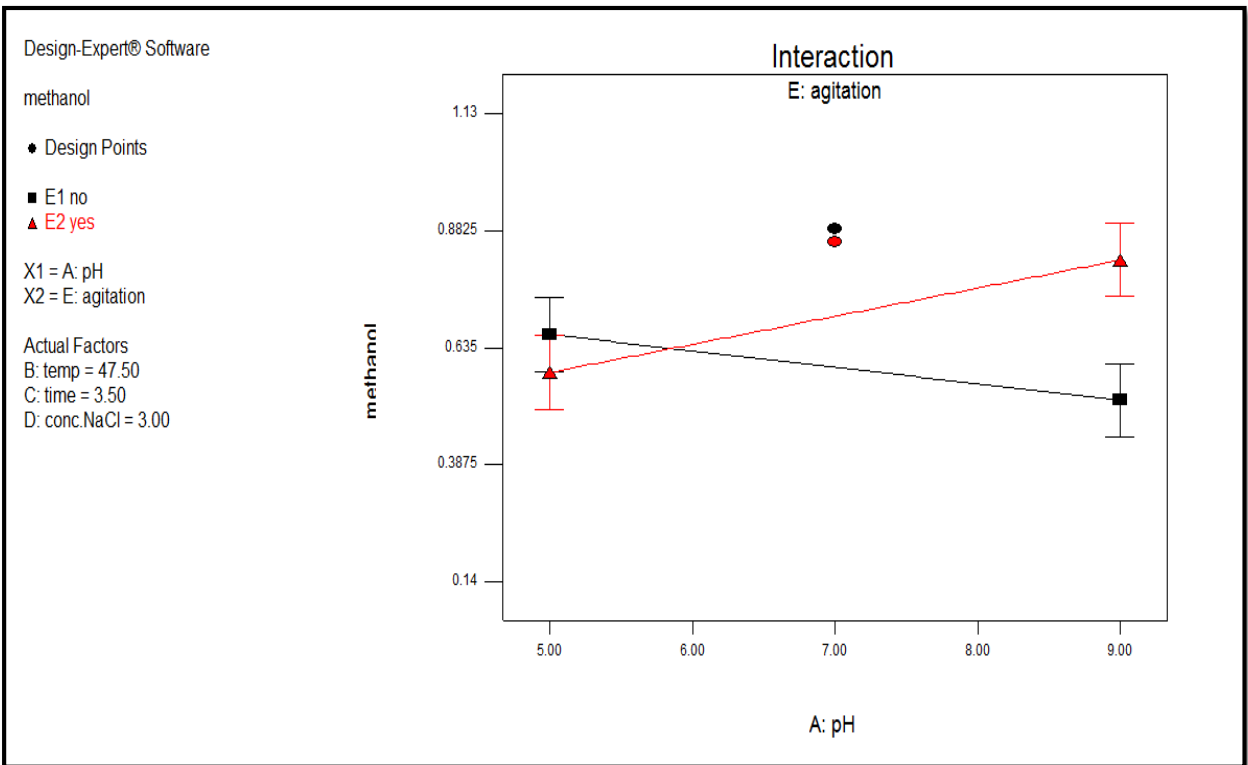


Figure 4.8: The interaction graph between factor A, pH and factor E, agitation

4.6 Comparison of methanol production with other researchers

Over the years, there were many type of biomass and processes that been used in order to produce methanol including agricultural waste, forestry waste, fishery waste, livestock and poultry waste and also sewage sludge. Jawad *et al.*, (2013) stated that based on depending the availability of the substrate, a lot of agricultural waste can be applied in bioprocess industry as an alternative way to replaced costly raw material. Biomass can be used as an alternative because it was a source of sustainable energy. Hence, it was also environmental friendly and had a significant economic potential. In addition, it also created energy security through the production. This waste biomass consists of animal, agricultural forestry and industry types of waste. Hence, there was also had different processed in production of methanol by using biomass such as pyrolysis, gasification and biosynthesis method (Shamsul *et al.*, 2014).

Methanol production by enzymatic reaction process was affected by several factors such as temperature, pH, agitation, concentration of NaCl and enzymatic reaction time. However, there might be having differences in methanol production in this research study with some other researchers due to the selection of variance factors used.

According to Rouse & Atkins (1954), high PME activity can be detected in lime. This supported by Kurita *et al.*, (2008) where citrus fruit such as lime and orange is known for its high amounts of pectin methylesterase (PME) where this PME enzyme can be de-esterification to form methoxyl group which is methanol.

Nielsen & Christenten (2002) reported from the previous research for the activity of PME from the extraction of orange, lemon, lime, grapefruit and clementine fruit. From the extraction of these fruits, it showed that the specific activity of PME was highest in orange and lime which is about 17.8 wt% and 15.6 wt%. Extracts from the separated peels and fruit flesh showed that the level of PME activity was higher in peels extract rather than extract from fruit flesh.

Kamarudin *et al.*, (2013) stated that in comparison between forestry waste, agricultural waste and animal waste, the methanol yield in agricultural waste showed the highest percentage between 0.5-5.93 wt.% rather than forestry waste about 0.1-4.36 wt.% and animal waste that showed methanol yield around 0.4-1.2 wt.%.

As Shamsul *et al.*, (2014) stated, the greatest contribution to biomass energy was from agricultural waste that comprised about 64 wt. % of the total energy demand. Reno *et al.*, (2011) claimed that sugarcane bagasse and corncobs have the potential to produce alcohol components. Hence, Nakagawa *et al.*, (2007) also tested various types of agricultural waste biomass, such as bran, straw and husks from rice and found that rice bran produced high methanol yield about 55 wt% whereas rice straw exhibited methanol yield about 36wt % and rice husks about 39 wt%. Chia (2011) supported it by doing a research using rice husks, sugarcane bagasse, corncobs and nutshell samples to examine the presence of alcohol and other component and proved that sugarcane bagasse contain 5.93 wt.% bio-methanol while corncob produced 0.67 wt.% of bio-methanol by weight.

Table 4.3: Comparison for methanol production from agricultural source

Substrate	Methanol production (wt %)	References
Agricultural waste	0.5 – 5.93	Kamaruddin <i>et al.</i> , (2013)
Forestry waste	0.1 – 4.36	
Sugarcane bagasse	5.93	Chia (2011)
Orange	17.8	Nielsen & Christenten (2002)
Lime	15.6	
Rice bran	55	Nakagawa <i>et al.</i> , (2007)
Rice straw	36	
Rice husk	39	
This work	2.24	-

5. CONCLUSION & RECOMMENDATIONS

This chapter were written to summarize all the results and discussion of the data that presented in chapter 4. A recommendation for been used in further study was also suggested for biological production of methanol by using pectin methylesterase (PME).

5.1 Conclusion

This research study was focused on the production of methanol by using pectin methylesterase (PME) and analysing the factors that affecting the methanol production from pectin methylesterase (PME).

The RSM study to improve the production of methanol via Fractional Factorial Design (FFD) was used to determine the highest condition for the methanol production by using pectin methylesterase (PME). The ANOVA and regression analysis showed significant result towards all the five factors that had been used through this research. The quadratic model of regression analysis for methanol production was used to predict all the response and the R^2 value for methanol production recorded was 0.9904.

The main effect analysis showed that factor D showed the highest percentage contribution for methanol production with 45.36% compared the other factor. While factor C showed the least percentage of contribution for methanol production with 0.34%.

Interaction between factors was represented the combined effects of factors on the dependent measured. It were likely improved the production of methanol because some of the factor may contributed more that the main factor alone. From the result of the FFD analysis, interaction between factor A at pH7 and factor B at temperature 70° C showed the highest methanol production with percentage contribution was 13.22%.

From this research study, it can be concluded that pectin methylesterase (PME) that come from lime peels waste can be potentially promising source in methanol production. Through this research study also, it can be concluded that all the factors that were chosen give effect and play important role in production of methanol. This finding are important to showed that PME from lime peels waste can be used as a cheap substrate in order to reduce the cost in production of methanol. The results that been recorded through this research showed that production of methanol from biomass can lead to the eco-friendly that also can be alternative energy to replace non-renewable energy and fulfilled the increasing in the worldwide energy demand in the future. Future studies were needed to be done for extra proof on the usage of PME from lime peels waste in methanol production.

5.2 *Future work recommendations*

The research carried out in this project was to produce methanol by using pectin methylesterase (PME) where this methanol will be very important chemical in the future and also to analyse the factors that affecting the production of methanol. From the results and conclusion obtained, it was recommended to conduct a research study of experiment for further improved research in this field of study. The following recommendations are:

1. Study on the validation study in order to determine the best condition by using PME to get the highest amount of methanol.
2. The optimization study by using the best condition of factors that had been recorded through this research study in order to obtained the highest production of methanol.

6. REFERENCES

Achmad, F., Kamaruddin, S. K., Daun, W. R. W., Majlan, E. H. Passive direct methanol fuel cells for portable electronic devices. *Appl Energy* 2011;88:1681-9.

Amaral, S. H. D., Assis, S. A. D., Oliveira, O. M. M. D. F. (2005). Partial purification and characterization of pectin methylesterase from orange (*Citrus sinensis*) CV Pera-Rio. *J. Food Biochem.* 29:367-380.

Annur, M. S. M., Tan, I. K. P., & Ramachandran, K. B. (2008), "A Kinetic Model for Growth and Biosynthesis of Medium-Chain-Length Poly-(3-Hydroxyalkanoates) in *Pseudomonas putida*," *Brazilian Journal of Chemical Engineering*, Vol. 25, pp. 217 – 228.

Arotupin, D. J., Akinyosoye, F. A., Onifade, A. K. (2008). Purification and characterization of pectin methylesterase from *Aspergillus reapeans* isolated from cultivated soil. *Afr. J. Biotechnol.* 7:1991-1998.

Assis, S. A., Lima, D. C., Faria Oliveira, O. M. M. (2001). Activity of pectinmethylesterase, pectin content and vitamin C in acerola fruit at various stages of fruit development. *Food Chem.* 74:133-137.

Aw, L., Vasiliou, E. G., Georgakopoulos, D. G., Yialouris, C. P., & Georgiou, C. A. (2013). Rapid Enzymatic Method for Pectin Methyl Esters Determination, 2013.

Aziah, N., & Bhat, R. (2012). Mineral composition and pasting properties of banana pseudo-stem flour from *Musa acuminata* X *balbisiana* cv . Awak grown locally in Perak , Malaysia, 19(4), 1479–1485.

Baş, D., & Boyacı, İ. H. (2007). Modeling and optimization I: Usability of response surface methodology. *Journal of Food Engineering*, 78(3), 836–845.

doi:10.1016/j.jfoodeng.2005.11.024

Benen, J. A. E.,; Van Alabeek, G. J. W. M; Voragen, A. G. J. Handbook of food enzymology. New York: Basel, 2003.

- Bezerra, M. A., Santelli, R. E., Oliveira, E. P., Villar, L. S., & Escaleira, L. A. (2008). Response surface methodology (RSM) as a tool for optimization in analytical chemistry. *Talanta*, 76(5), 965–77. doi:10.1016/j.talanta.2008.05.019
- Bhattacharyya, J. K., Kumar, S., Devotta, S. Studies on acidification in two-phase biomethanation process of municipal solid waste. *Waste Manag* 2008;28:164-9.
- Bridgewater, A. V. (Ed). (2001). *Progress in Thermochemical Biomass Conversion*. Blackwell, Oxford, UK.
- Carbonell, J. V., Contreras, P., Carbonell, L., & Navarro, J. L. (2005). Pectin methylesterase activity in juices from mandarins, oranges and hybrids. *European Food Research and Technology*, 222(1-2), 83–87. doi:10.1007/s00217-005-0043-9
- Chaitanya, M., & Jaya, K. (2014). *Journal of Chemical , Biological and Physical Sciences* Production of pectinase enzyme under solid state fermentation using banana pseudostem by *Aspergillus niger* NCIM 616, 4(4), 3900–3908.
- Chia SK. Production of bio-methanol from agricultural waste by pyrolysis method. Malaysia: Department of Chemical and Process Engineering National University of Malaysia; 2011 [Thesis].
- Contreras-esquivel, J. C., Correa-robles, C., Aguilar, C. N., & Rodríguez, J. (1999). Pectinesterase extraction from Mexican lime (*Citrus aurantifolia* Swingle) and prickly pear (*Opuntia ® cus indica* L .) peels, 65, 0–3.
- Corradetti, A., Desideri, U. (2007). Should biomass be used for power generation or hydrogen production. *Journal of Engineering for Gas Turbines and Power* 129, 629-636.
- Don, M. (2013). Screening of significant factors in collagen extraction from hybrid *Clarias* sp . using a statistical tool, 20(4), 1913–1920.
- Garcia-castello, E. M., Mayor, L., Alcaraz, N., Gras, M. L., Argüelles, A., & Brotóns, D. V.-. (2012). Orange Solid Waste Valorization : Optimization of Pectinase Extraction and Enzymatic Treatment of Orange Press Liquor, 29, 823–828. doi:10.3303/CET1229138

- Golshani, T., Jorjani, E., Chehreh, C. S., Shafaei, S. Z., & Heidari, N. Y. (2013). International Journal of Mining Science and Technology Modeling and process optimization for microbial desulfurization of coal by using a two-level full factorial design. *International Journal of Mining Science and Technology*, 23(2), 261–265. doi:10.1016/j.ijmst.2013.04.009
- Gonzalez, S. L., & Rosso. N. D., (2011). Determination of pectin methylesterase activity in commercial pectinases and study of the inactivation kinetics through two potentiometric procedures. 31(2): 412-417.
- Giovane, a, Servillo, L., Balestrieri, C., Raiola, a, D'Avino, R., Tamburrini, M., ... Camardella, L. (2004). Pectin methylesterase inhibitor. *Biochimica et Biophysica Acta*, 1696(2), 245–52. doi:10.1016/j.bbapap.2003.08.011
- Güllü, D., Demirbas, A. Biomass to methanol via pyrolytic process. *Energy Convers Manag* 2001;42:1349-56.
- Hall, D. O., Rosillo-Calle, F., de Groot, P. (1992). Biomass energy lessons from case studies in developing countries. *Energy Policy*, 62-73.
- Hamelinck, C. N. (2002). Future prospects for production of methanol and hydrogen from biomass, 111, 1–22.
- Hamelinck, C. N., Faaij, A. P. C. (2008). Production of methanol from biomass. Utrecht University. NWS-E-2006-387.
- Hasyierah, N., Salleh, M., Zulkali, M., Daud, M., Arbain, D., Ahmad, M. S., ... Ismail, K. (2011). Optimization of alkaline hydrolysis of paddy straw for ferulic acid extraction. *Industrial Crops & Products*, 34(3), 1635–1640. doi:10.1016/j.indcrop.2011.06.010
- Jawad, A. H., Alkarkhi, A. F. M., & Norulaini, N. A. N. (2013). Production of the lactic acid from mango peel waste – Factorial experiment. *Journal of King Saud University - Science*, 25(1), 39–45. doi:10.1016/j.jksus.2012.04.001
- Jayani, R. S.; Saxena, S.; Gupta, R. Microbial pectinolytic enzymes: A review. *Process Biochemistry*, v. 40, p. 2931-2944, 2005. <http://dx.doi.org/10.1016/j.-rocbio.2005.03.026>

- Kamarudin, S. K., Shamsul, N. S., Ghani, J. A., Chia, S. K., Liew, H. S., & Samsudin, A. S. (2013). Bioresource Technology Production of methanol from biomass waste via pyrolysis. *Bioresource Technology*, *129*, 463–468. doi:10.1016/j.biortech.2012.11.016
- Khalil, M., Darusman, L. K., & Syafitri, U. D. (2011). Application of fractional factorial design to optimize extraction method of artemisinin from *Artemisia annua*, *37*, 219–224. doi:10.2306/scienceasia1513-1874.2011.37.219
- Kova, A. (2014). Journal of Natural Gas Science and Engineering Selecting different raw materials for methanol production using an MINLP model, *19*, 221–227. doi:10.1016/j.jngse.2014.05.008.
- Kumabe, K., Fujimoto, S., Yanagida, T., Ogata, M., Fukuda, T., Yabe, A., et al. Environmental and economic analysis of methanol production process via biomass gasification. *Fuel* 2008;87:1442-7.
- Kurita, O., Fujiwara, T., & Yamazaki, E. (2008). Characterization of the pectin extracted from citrus peel in the presence of citric acid. *Carbohydrate Polymers*, *74*(3), 725–730. doi:10.1016/j.carbpol.2008.04.033
- Kralj, A. K. (2009). Methanol production from biogas. Third International Conference on Energy Environment – Biomedicine.
- Leduc, S., Lundgren, J., Franklin, O., Dotzauer, E. Location of a biomass based methanol production plant: a dynamic problem in northern Sweden. *Appl Energy* 2010;87:68-75.
- Luiza, M., Renó, G., Eduardo, E., Lora, S., Carlos, J., Palacio, E., ... Almazan, O. (2011). A LCA (life cycle assessment) of the methanol production from sugarcane bagasse. *Energy*, *36*(6), 3716–3726. doi:10.1016/j.energy.2010.12.010.
- Mamma, D., Kourtoglou, E., Christakopoulos, P. (2008). Fungal multienzyme production on industrial by-products of the citrus-processing industry. *Bioresour. Technol.* *99*, 2373-2383.
- Mastro, F. L., Mistretta, M. Cogeneration from thermal treatment of selected municipal solid wastes. A stoichiometric model building for the case study on Palermo. *Waste Manag* 2004;24:309-17.

Merrill, R. C., & Weeks, M. (1945). The thermal degradation of pectin. *Journal of the American Chemical Society*, 67, 2244-2247.

Mussatto, S. I., & Teixeira, J. A. (2010). Lignocellulose as raw material in fermentation processes, 897–907.

Nakagawa, H., Harada, T., Inchinose, T., Takeno, T., Matsumoto, S., Kobayashi, M. Bio-methanol production and CO₂ emission reduction from forage grasses, trees, and crop residue. *JARQ Jpn Agric Res Q* 2007;41 : 173-80.

Nielsen, J. E., & Christensen, M. I. E. (2002). Distribution of pectin methyl esterase and acetylerase in the genus *Citrus* visualized by tissue prints and chromatography, 162, 799–807.

Of, A., & Chemical, T. H. E. (2010). Analysis of the chemical composition and morphological structure of banana pseudo-stem, 5, 576–585.

Okonko, I. O., Egwame, R. A., Fajobi, E. A., Nkang, A. O., Iheakanwa, C. I., Ogunnusi, T. A., et al. Current trends in bio-fuel production and its use as an alternative energy security. *Environ Agric Food Chemi* 2009;8:1233-60.

Patil, N. P., & Chaudhari, B. L. (2010). Production and purification of pectinase by soil isolate *penicillium* sp and search for better agro-residue for its use, 2(7), 36–42.

Pavlidis, P. (2003). Using ANOVA for gene selection from microarray studies of the nervous system. *Methods*, 31(4), 282–289. doi:10.1016/S1046-2023(03)00157-9

Rodriguez-lopez, A. D., Mayor, L., Galfarsoro, M. M., Martinez-otalo, J., & Garcia-castello, E. M. (2013). Pectinmethylesterase extraction from orange solid wastes : Optimization and comparison between conventional and ultrasound-assisted treatments, 4(9), 45–50.

Rouse, A. H., & Atkins, C. D. (1954). Lemon and lime pectinesterase and pectin. *Proc. Fla. State Hort. Soc.*, 67, 203-206.

Shamsul, N. S., Kamarudin, S. K., Rahman, N. a., & Kofli, N. T. (2014). An overview on the production of bio-methanol as potential renewable energy. *Renewable and Sustainable Energy Reviews*, 33, 578–588. doi:10.1016/j.rser.2014.02.024

Şimşek, Ş., & Yemenicioğlu, A. (2010). Commercially suitable pectin methylesterase from Valencia orange peels, *34*, 109–119. doi:10.3906/tar-0903-7

Spagna, G., Barbagallo, R. N., & Ingallinera, B. (2003). A specific method for determination of pectin esterase in blood oranges, *32*, 174–177.

Tijskens, L. M. M., Rodis, P. S., Hertog, M. L. A. T. M., Proxenia, N., VAN DIJK, C. (1999). Activity of pectin methylesterase during blanching of peaches. *J. Food. Eng.* *39*:167-177.

Trop, P., Anicic, B., & Goricanec, D. (2014). Production of methanol from a mixture of torrefied biomass and coal. *Energy*, 1–8. doi:10.1016/j.energy.2014.05.045.

Wang, Q. Biomethanol conversion from sugar beet pulp; 2006. [Online] Available from: [<http://drum.lib.umd.edu/bitstream/1903/3833/1/umi-umd-3678.pdf>] Revised; March 2015.

Wilkins, P. P., Parish, M. E. (1997). Minimum inhibitory concentrations of antimicrobials against micro-organisms related to citrus juice. *Food Microbial.* *14*, 373-381.

Xu, Y., Ye, T., Qiu, S., Gong, F., Liu, Y. High efficient conversion of CO₂-rich bio-syngas to CO-rich bio-syngas using biomass char: A useful approach for production of bio-methanol from bio-oil. *Bioresour Technol* 2011;*102*:6239-45.

APPENDIX

Table A1: Table of graph of concentration of methanol versus pH

pH	Conc. Of methanol (g/l)
3	7.34
4	15.998
6	17.847
7	22.652

Table A2: Table of graph of concentration of methanol versus pH

Temperature °C	Conc. Of methanol (g/l)
20	7.984
50	22.353
60	31.538
70	57.669

Table A3: Table of graph of concentration of methanol versus pH

NaCl (M)	Conc. Of methanol (g/l)
1.5	4.476
2.5	5.126
3	5.183

Table A4: Table of graph of concentration of methanol versus pH

Time (hours)	Conc. Of methanol (g/l)
2	14.596
3	20.559
5	36.465