Impact of Antimicrobial Agents against Oral Isolates

Muna Jalal^{a,b,*}, Essam A. Makky^a and Mashitah M. Yusoff^a

Abstract---Tooth decay is considered the most widespread infectious disease in the world. Its spread increases with time, and this increase is associated significantly with the change of dietary mode of modern humans. This study aims to isolate and identify the important bacteria related to tooth decay, determine the sensitivity of bacteria in certain types of antimicrobial agents, and study the effect of heavy metals on bacterial isolates. A total of 50 swabs were collected from the mouths of patients from both sexes, with ages ranging from 1-60 years. The patients were advised to consult with dental clinics and specialized centers to isolate and identify the causative agents associated with oral diseases. Results showed that infection rates in younger age groups (1-20 and 20-40) are higher than the elder group (40-60), with percent incidence of 44% and 32%, respectively. Antibiotic sensitivity test against the isolates showed that chloramphenicol had the highest sensitivity effect with 83.2% followed by rifampicin and gentamicin with 81.35%, penicillin G with 64.40%, and streptomycin with 16.94% Also, these differences were found have lower effect for isolates against (10) heavy metals, where it showed resistance to Iron 3.38%, then nickel , aluminum ,copper, lead to 20.33%, 22.03%, 27.11%, 28.81% respectively, also sliver shown 57.62%. And, this similarity were found have sensitive to antimony and chromium 61.01%.while appeared sensitivity to mercury and cadmium by 100, 86.44.

Keywords _Antibiotic; Heavy metals; Bacteria; Tooth.

I. INTRODUCTION

TOOTH decay is one of the most common infectious diseases affecting millions of people globally [1]. One of the occasional factors for the disease is dental biofilm, which is the bacterial charge that forms permanently on the tooth surfaces [2]. Hazard factors include unsuitable salivary flow, low quality of salivary buffer, incomplete fluoride exposure, and increased consumption of sugar [3]. Caries indicate the centralized removal of susceptible dental hard tissues by acidic products from the bacterial fermentation of dietary carbohydrates [4]. Tooth decay is a chronic disease that is slowly developing in people. Tooth decay presents as smooth holes and fissured surfaces on the crown and root of a tooth. According to the World Health Organization, 60-90% of school children worldwide have dental cavities [5]. This decay is the result of the interaction of the oral microflora plaque, the tooth surface, nourishment, and the oral Environment over time, causing destruction of the tooth enamel [6]. Recently, disease incidence for cavities is decreasing in industrialized nations but is increasing in developing nations [7]. The spread of caries is uneven across the population and communities. The highest incidence is in the lower socioeconomic groups, having limited access to adequate oral health care [8]. Despite the decline in incidence of caries, the United States of America is spending 10 billion USD each year on tooth decay treatment [9]. In other industrialized nations, such as the United Kingdom and China, caries prevalence in the past has been over 50% in children. In developing countries, where oral health care is low, caries are increasing in an alarming rate. Previous studies done in Peru, Mexico, the Philippines, and Taiwan found caries in 75-90% of children [10].

Mutants Streptococci, a group of cariogenic bacteria, is associated in the initiation of dental caries [11]. Another group of bacteria that is substantial in the development of caries is Lactobacillus. Lactobacillus does not usually colonize the tooth surface, but is commonly found in the oral cavity including the dorsum of the tongue [1]. Although it could have a significant role in the caries advancement, Lactobacillus is not essential in the initiation of dental caries [12]. Positive association between salivary levels and bacterial caries is relevant to carbohydrate exhaustion. The presence of Streptococcus and Lactobacillus may potentially indicate the occurrence of not only caries but also of carbohydrate consumption [13]. Streptococcus mutans is commonly accepted as one of the most substantial etiologic agents in caries development and has been shown to directly cause caries in germ-free and specific pathogen-free rat models. However, the presence of caries has been found even in the absence of S. mutans. Although a high percentage of S. mutans has been recovered from teeth without caries, S. mutans remains the species that is most associated with caries. In gnotobiotic and specific germ-free rodent models, S. mutans has the potential to generate caries [14]. Despite the various properties in S. mutans that raises its cariogenicity, strong biofilm indicating the presence of dietary sucrose is a stringent component in the development of caries.

Thus, this study aims to isolate and partially identify important bacteria related to tooth decay and diseases of the mouth, determine the susceptibility of bacteria to certain types of antimicrobial agents

Muna Jalal^{a,b,*} ^aFaculty of Industrial Sciences & Technology, Universiti Malaysia Pahang, 26300 Gambang, Kuantan, Pahang, Malaysia

^bAl-Haweeja Institute, Foundation of Technical Education, Kirkuk, Iraq *Corresponding author: E-mail: <u>munajalal2@gmail.com</u>; Phone: 0129607054; fax: +609-5492766

Essam A. Makky^a and Mashitah M. Yusoff^{a : a}Faculty of Industrial Sciences & Technology, Universiti Malaysia Pahang, 26300 Gambang, Kuantan, Pahang, Malaysia.

II. MATERIALS AND METHODS

Isolation of microbial isolates from patients Collection of samples: With the assistance of dentists, specimens in this study have been collected from the dental units in health centers and dental clinics in Gambang, Pahang, Malaysia. Sterile swabs were used for the patients of both genders, with ages ranging from 1–60 years. Collected samples were - transferred to the laboratory of Universiti Malaysia Pahang.

Microbial culture

Samples from the mouth of patients were cultured on nutrient agar plates and were incubated at 37° for 24 h. The samples were then purified and cultured on agar slants. These were kept in the chiller until use

Gram stain

All isolates were partially identified using Gram stain reaction for microscopic examination and identification Antimicrobial activity test using disc diffusion method

Antibiotic sensitivity test

All antibiotics used in this study were from Mast disctm, Mast Diagnostics, Mast group, Mersey side, except for penicillin G, which was from Oxoid, Basingstoke, Hampshire, England. Streptomycin was prepared in the laboratory. Antibiotic discs (amoxicillin 10 µg, neomycin 10 µg, ampicillin 10 µg, tetracycline 10 µg, gentamicin 10 µg, chloramphenicol 110 µg, penicillin G 10 µg, streptomycin 10 μ g, and rifampicin 5 μ g) used Muller–Hinton agar from Hardy According Diagnostics. to the manufacturer's recommendations, the antibiotic discs were prepared and were autoclaved at 12 °C for 15 min. The medium was then cooled to 45-50 °C and poured onto the plates. The antibiotic discs were allowed to set on a level surface to a depth of approximately 4 mm. Inoculums from primary culture plates were prepared by touching 3-5 colonies with a swab and transferring them into a plate. The inoculums were mixed with two drops of sterile distilled water and were spread in three plates. The nine antibiotic discs prepared were placed onto the inoculated plates. Subsequently, they were placed in the chiller for 15 min and were incubated at 37 °C. After an overnight incubation, the diameter of each inhibition zone was measured and recorded in mm [15].

Heavy metals activity test

Prepare concentration: Prepared concentration was prepared by using 10 milligram /liter for the ten heavy metals (i.e,Iron, copper , aluminum, antimony, nickel, lead, Silver,chromium,cadmium and mercury) The stock solution was prepared for concentration . Filter paper disc was used and laden with 25 μ l of heavy metal [16].

III. RESULTS

Patients isolates

In this study, has been obtained from the mouths of 50 patients in different ages and genders with percentage of 54% males and females, as shown bacteria and yeast (59) isolates in the Table 1 and Fig. 1. It has been shown the primary isolation of samples. The impact of age on the infection rates

of the tooth caries showed the age groups of (20-40 years) and (1-20 years) were the most of the infected compared to elder group (40-60 years), as was the incidence of 44% and 32% respectively.

 TABLE 1

 PRIMARY ISOLATION OF SAMPLES AND PERCENTAGES.

Patients Samples & age (year)	Isolate number	Percentage (%)
Single isolate	33	55.93
Mixed isolate	26	44.07
1-20	16	32
20-40	22	44
40-60	12	24



Fig. 1. Percentage of isolates according to age group.

The sensitivity of bacteria to antibiotics

Data represented in Fig. 2 show the percentage sensitivity of bacterial isolates against nine antibiotics, where it showed they were streptomycin and penicillin G 16.94%, 64.40% respectively. While noted the highest sensitivity to antibiotics was chloramphenicol (83.05%), similarity sensitive for Gentamicin and Rifampicin with (81.35%).



Fig. 2. Percentage sensitivity of bacterial isolates antibiotics.

Sensitivity of bacteria to heavy metal

The results as shown in Fig. 3 that the resistance and sensitive percentages of bacterial isolates seven heavy metals, where it showed explain the resistance to heavy metals, Iron 3.38%, then nickel, aluminum ,copper, lead to 20.33%, 22.03%, 27.11%, 28.81% respectively, also sliver shown

57.62%. And, this similarity were found have sensitive to antimony and chromium 61.01%.while appeared sensitivity to mercury and cadmium by 100%, 86.44%



Fig. 3. Percentage of bacterial isolates sensitive of heavy metal.

IV. DISCUSSION

Patients isolates

The study confirmed that children and the younger are more susceptible to mouth infection. This may be due largely to reasons related to immune shortages of the infected people in these age groups as well as consciousness of health or other factors related to nutrition and public health that increase the rates of infection in patients of children and the younger [17] stated that children are more susceptible to the bacteria that cause decay. That necrosis of the infected children appears to have teeth mutans with its different kinds and with high rates. Also, the frequent consumption of sugar play an important role in the infection with emphasis on the role of the mother as a source of transmission of disease from her infected teeth to her baby, where the levels of these bacteria with mothers are similar to those found at their children.

Sensitivity of bacteria to antibiotics

The current study showed that the chloramphenicol is the best of antibiotic in its influence on the bacterial isolates taken from the mouth, followed by the antibiotic Gentamicin and Rifampicin. The antibiotic tetracycline, Ampicillin, Neomycin, Amoxcillin are less effective on the bacterial isolates. One of the results we have noticed that the bacterial isolates showed variation in their resistance to antibiotics of the group of aminoglycosidate, the ratio of sensitive to Neomycin 74.57%. The percentage of sensetive to Gentamicin by 81.35%. The resistance to aminoglycosidate antibiotics increased notably in recent times, Livermore and Winstanley, 2001 studied Relationships between antibiotic and mechanism are also presented to allow full interpretative reading for those testing wide panels of drugs versus isolates. This resistance which is due to the formation of the enzyme by resistant bacteria modifies the antibiotic and thus loses its effectiveness or because of loss of outer membrane proteins, which reduces the permeability of the antibiotic inside the bacterial cell [18]. Evidenced by the results of the current study, the majority of bacterial isolates possessed prescription relatively high resistance to antibiotics represented β -lactam (Ampicillin, Amoxicillin, Pencillin G). The high bacterial resistance to

antibiotics β -lactam due to several mechanisms, most notably the ability to produce enzymes which β -lactamase the broken bind β -lactam, change the permeability barrier intimacy between the antbiotic and locations of the target Penicillin Binding Protein, came our results are compatible with Cherian and Manjunath, 2003 during their study that extended spectrum beta lactamase producing enterobacteriaceae in a tertiary care hospital in Trinidad and Tobago [19]. The results of this study also showed the high resistance shown by the bacterial isolates to streptomycin explains the mechanism of resistance to this antibiotic, Speculation on this mechanism indicates that the binding of the molecule to the 30S subunit interferes with 50S subunit association with the mRNA strand. This result in an unstable ribosomal-mRNA complex, leading to a frameshift mutation and defective protein synthesis leading to cell death. Syal, et al 2013, reported that streptomycin therapeutic concentrations of 10 mg/mL interfere in the Jaffe reaction and acted as non-creatinine chromogen during in their study that referred Streptomycin interference in Jaffe reaction Possible false positive creatinine estimation in excessive dose exposure [20]. The study also shows an increasing resistance to tetracycline, it is believed that this resistance resulted from the presence of plasmids that encode resistance to the antibiotic which moves significantly. Koo and Woo, 2011 during their study that distribution and transferability of tetracycline resistance determinants in E. coli isolated from meat and meat products reported that the high prevalence of tetracycline resistant Escherichia coli in meat may be due to the high transferability of tetracycline determinants [21]. It is noted during the study that the lowest resistance showed by the bacterial isolates was to chloramphenicol, Gentamicin and Rifampicin. It appeared that most of the bacterial isolates were sensitive to these adversaries and may be due to response to the majority of the isolates of these two adversaries to being of limited use antibiotics at the present time in hospitals, leading to increased resistance to antibiotics can be passed as determinants responsible for drug resistance to antibiotics by plasmids.

Sensitivity of bacteria to heavy metal

The results of this study also showed the high resistance shown by the bacterial isolates to iron Guohua Jiang et al studied reduction of iron in scrubbing solution by magnetic iron microspheres immobilized of iron reducing bacteria so reported that magnetic microspheres to immobilize iron reducing bacteria to improve the biological reduction of iron which was one of the key steps in nitrogen oxides removal by the integrated chemical absorption biological reduction process .The immobilized bacteria performed well iron reduction than free bacteria even under unfavorable pH and temperatures[22]. Evidenced by the results of the current study, the majority of bacterial isolates possessed prescription relatively high sensitive to mercury. Mercury is additionally the sole microorganism metal resistance system whose mechanism ends up in large-scale transformation of its target. The mechanisms of different ion to resistances are supported effluent pumps or living thing sequestration. Barkay and Miller (2003) studied bacterial mercury resistance from atoms to ecosystems so reported that one or more proteins apparently involved in transport genes conferring occur on chromosomes, plasmids, and transposons and their operon arrangements can be quite diverse, structural genes, several of which are modular. proteins protects host cells from this toxic metal [23]. The data obtained during this study clearly shows that with sensitive microorganism of cadmium. This may be due largely to reasons related to less concentration from cadmium increase the rates of sensitive isolates. Cohen, et al. (1990) studied that the effect of zinc and cadmium ions on Escherichia coli and they were noted that exposure of E. coli to various concentrations of these ions resulted in an increase of the total protein and the metal binding proteins amount in the cells. The activity of alkaline phosphates was raise in the presence of these ions [24].

ACKNOWLEDGEMENT

The authors gratefully acknowledge University Malaysia Pahang (UMP), Malaysia for the financial supported by grant GRS 140318 that enables the authors to accomplish this work.

REFERENCES

- Wongkamhaeng, K., O. Poachanukoon, and S. Koontongkaew, *Dental caries, cariogenic microorganisms and salivary properties of allergic rhinitis children*. International Journal of Pediatric Otorhinolaryngology, 2014.
- [2] Petersen, P.E., et al., *The global burden of oral diseases and risks to oral health*. Bulletin of the World Health Organization, 2005. 83(9): p. 661-669.
- [3] MejÀre, I., et al., *Caries risk assessment. A systematic review.* Acta Odontologica Scandinavica, 2014(0): p. 1-11.
- [4] Selwitz, R.H., A.I. Ismail, and N.B. Pitts, *Dental caries*. The Lancet, 2007. **369**(9555): p. 51-59.
- [5] Petersen, P.E., World Health Organization global policy for improvement of oral health-World Health Assembly 2007. International dental journal, 2008. 58(3): p. 115-121.
- [6] Lynch, D.J., An analysis of the role of glucan-binding proteins in Streptococcus mutans biofilm architecture and caries development. 2010.
- [7] Chu, C. and E. Lo, Promoting caries arrest in children with silver diamine fluoride: a review. Oral health & preventive dentistry, 2008. 6(4).
- [8] Bowen, W.H., Do we need to be concerned about dental caries in the coming millennium? Critical Reviews in Oral Biology & Medicine, 2002. 13(2): p. 126-131.
- [9] Benjamin, R.M., Oral health: the silent epidemic. Public health reports, 2010. 125(2): p. 158.
- [10] Bagramian, R.A., F. Garcia-Godoy, and A.R. Volpe, *The global increase in dental caries. A pending public health crisis.* Am J Dent, 2009. 22(1): p. 3-8.
- [11] Loesche, W.J., Role of Streptococcus mutans in human dental decay. Microbiological reviews, 1986. 50(4): p. 353.
- [12] Takahashi, N. and B. Nyvad, *The Role of Bacteria in the Caries Process Ecological Perspectives*. Journal of Dental Research, 2011. **90**(3): p. 294-303.
- [13] Van Houte, J., Microbiological predictors of caries risk. Advances in dental research, 1993. 7(2): p. 87-96.
- [14] Takahashi, N. and B. Nyvad, Caries ecology revisited: microbial dynamics and the caries process. Caries research, 2008. 42(6): p. 409-418.
- [15] Vandepitte, J., et al., Basic laboratory procedures in clinical bacteriology. 2003: World Health Organization.
- [16] Bakht, J., et al., Antimicrobial potentials of Eclipta alba by disc diffusion method. African Journal of Biotechnology, 2013. 10(39): p. 7658-7667.
- [17] Rao, G.G., Risk factors for the spread of antibiotic-resistant bacteria. Drugs, 1998. 55(3): p. 323-330.
- [18] Livermore, D.M., T.G. Winstanley, and K.P. Shannon, *Interpretative reading: recognizing the unusual and inferring resistance mechanisms*

from resistance phenotypes. Journal of Antimicrobial Chemotherapy, 2001. **48**(suppl 1): p. 87-102.

- [19] Cherian, B., et al., Extended-spectrum beta-lactamase producing enterobacteriaceae in a tertiary care hospital in Trinidad and Tobago. The West Indian medical journal, 2003. 52(1): p. 31-33.
- [20] Syal, K., A. Srinivasan, and D. Banerjee, Streptomycin interference in Jaffe reaction—possible false positive creatinine estimation in excessive dose exposure. Clinical biochemistry, 2013. 46(1): p. 177-179.
- [21] Koo, H.-J. and G.-J. Woo, Distribution and transferability of tetracycline resistance determinants in< i> Escherichia coli</i> isolated from meat and meat products. International journal of food microbiology, 2011. 145(2): p. 407-413.
- [22] Jing, G., et al., Reduction of Fe (III) EDTAin a NO scrubbing solution by magnetic Fe chitosan microspheres immobilized mixed culture of ironreducing bacteria. Bioresource Technology, 2012. 108: p. 169-175.
- [23] Barkay, T., S.M. Miller, and A.O. Summers, *Bacterial mercury resistance from atoms to ecosystems*. FEMS microbiology reviews, 2003. 27(2-3): p. 355-384.
- [24] Cohen, I., R. Bitan, and Y. Nitzan, *The effect of zinc and cadmium ions on Escherichia coli B.* Microbios, 1990. 68(276-277): p. 157-168.