

**PRODUCTION OF L-ASPARAGINASE
THROUGH BIODEGRADATION OF CHICKEN
BONE WASTES**

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MASTER OF SCIENCE (BIOTECHNOLOGY)

UNIVERSITI MALAYSIA PAHANG



SUPERVISOR'S DECLARATION

We hereby declare that we have checked this thesis and in our opinion, this thesis is adequate in terms of scope and quality for the award of the degree of Master of Science in Biotechnology

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I hereby declare that the work in this thesis is based on my original work except for quotations and citation which have been duly acknowledged. I also declare that it has not been previously or concurrently submitted for any other degree at Universiti Malaysia Pahang or any other institutions.

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Thesis submitted in fulfillment of the requirements
for the award of the degree of
Master of Science (Biotechnology)

Faculty of Industrial Sciences and Technology
UNIVERSITI MALAYSIA PAHANG

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DEDICATION

Dedicated to my beloved family

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LIST OF SYMBOLS

% Percentage

° C Degree Celsius

:

Ratio

*

Highest value

↓ Lower than Control

2nd Second

± Plus-minus sign

Km Michaelis constant

/ per

μ Micro

LIST OF ABBREVIATIONS

| | |
|--------------------|----------------------------------|
| ALL | Acute lymphoblastic leukemia |
| <i>A. niger</i> | <i>Aspergillus niger</i> |
| ATCC | American Type Culture Collection |
| BSA | Bovine serum albumin |
| cm | Centimetre |
| <i>C. albicans</i> | <i>Candida albicans</i> |
| CB | Chicken bone |
| CCB | Cooked chicken bone |
| CFF | Cell free-filtrate |
| CMC | Carboxymethyl cellulose |
| DEAE- Cellulose | Diethylaminoethyl cellulose |
| <i>E. coli</i> | <i>Escherichia coli</i> |
| EDTA | Ethylenediaminetetraacetic |
| FDA | Food and Drug Administration |
| g | Gram |
| h | Hour |

| | |
|------|-----------------------------------|
| IU | International unit |
| kDa | kiloDalton |
| L | Litre |
| mA | milliampere |
| min | Minute |
| mg | Milligram |
| ml | Millilitre |
| mM | Millimolar |
| ms | millisecond |
| M | Molar |
| MTCC | Microbial Type Culture Collection |
| N | Normality |
| NA | Nutrient agar |
| NB | Nutrient broth |
| ND | Not detected |
| nm | Nanometre |
| PDA | Potato dextrose agar |

| | |
|----------------------|-----------------------------------------------------------|
| PEG | Polyethylene glycol |
| pH | Potential hydrogen |
| PS | Phosphate solution |
| RCB | Raw chicken bone |
| rpm | Revolutions per minute |
| <i>S. cerevisiae</i> | <i>Saccharomyces cerevisiae</i> |
| SD | Standard deviation |
| SDS-PAGE | Sodium dodecyl sulfate polyacrylamide gel electrophoresis |
| SEM | Scanning electron microscopy |
| sp. | Species |
| TEM | Transmission electron microscopy |
| <i>T. reesei</i> | <i>Trichoderma Reesei</i> |
| UMP | Universiti Malaysia Pahang |
| viz | <i>Videlicet</i> |
| v/v | volume per volume |
| w/v | weight per volume |
| WHO | World Health Organization |

XRD X-ray powder diffraction

XPS X-ray photoelectron spectroscopy

LIST OF CHEMICAL FORMULAS

| | |
|--------------------------------------------------------------------|----------------------------------|
| CuSO ₄ .5H ₂ O | Copper (II) Sulfate Pentahydrate |
| HCl | Hydrochloric acid |
| KCl | Potassium chloride |
| KH ₂ PO ₄ | Potassium dihydrogen phosphate |
| KNaC ₄ H ₄ O ₆ .4H ₂ O | Potassium sodium tartrate |
| MgCl ₂ | Magnesium chloride |
| MgSO ₄ | Magnesium sulfate |
| NaCl | Sodium chloride |
| Na ₂ CO ₃ | Sodium carbonate |
| NaH ₂ PO ₄ | Monobasic sodium phosphate |
| Na ₂ HPO ₄ | Dibasic sodium phosphate |
| NH ₄ Cl | Ammonium chloride |
| (NH ₄) ₂ SO ₄ | Ammonium sulfate |
| NaNO ₃ | Sodium nitrate |
| NaOH | Sodium hydroxide |

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ABSTRAK

L-asparaginase mendapat permintaan yang tinggi dari pelbagai industri seperti farmaseutikal, makanan mahupun biopenderia. Ini adalah disebabkan oleh keunikian enzim L-asparaginase dalam menghidrolisis asid amino L-asparagina kepada L-asid aspartik dan ammonia. Oleh yang demikian, enzim L-asparaginase dihasilkan secara besar-besaran melalui proses fermentasi mikrob. Kajian ini bertujuan untuk menghasilkan enzim L-asparaginase melalui proses fermentasi mikrob yang ditambah dengan sisa tulang ayam selaku substrat oleh penghasil enzim yang terbaik. Lantaran itu, kajian ini boleh dikategorikan kepada 3 bahagian di mana bahagian pertama merangkumi beberapa eksperimen dalam usaha mengenalpasti penghasil enzim serta substrat yang terbaik manakala bahagian kedua pula mengenalpasti impak proses parameter yang optimum dengan menggunakan pendekatan Satu-Faktor-Pada-Satu-Masa. Selain itu, enzim L-asparaginase juga diteruskan dengan proses penulenan serta pencirian. Enam mililiter (5×10^8 sel/ml) *Escherichia coli* ATCC 10536 merupakan saiz inokulum yang optimum apabila ia diinkubasikan pada suhu 40°C dan pH 9 untuk 2 hari dalam setiap 50 ml media fermentasi. Tambahan pula, pertumbuhan maksimum *Escherichia coli* ATCC 10536 selaku penghasil enzim L-asparaginase apabila ia ditambah dengan 1.0 % w/v kanji, 0.2 % w/v ammonium klorida serta 1.0 % w/v sisa tulang ayam yang telah dimasak selaku sumber karbon, nitrogen serta substrat. Impak proses parameter yang dikaji mendapati bahawa *Escherichia coli* ATCC 10536 merupakan penghasil enzim L-asparaginase yang terbaik dan ia lebih mengemari kaldu nutrient sewaktu proses fermentasi. Enzim L-asparaginase yang terhasil juga diteruskan dengan proses penulenan termasuk pemendakan ammonium sulfat, dialisis serta selulosa DEAE kromatografi turus. Sebanyak 0.42 % enzim L-asparaginase diperoleh di akhir proses penulenan. Di samping itu, proses pencirian juga dilaksanakan pada enzim yang diperoleh selepas proses dialisis dan didapati bahawa 40°C dan pH 8 merupakan suhu inkubasi dan pH yang optimum. Selain itu, Na^+ merupakan ion logam yang terbaik dalam usaha mengawal selia aktiviti enzim manakala EDTA pula ialah perencat enzim. Kesimpulannya, *Escherichia coli* ATCC 10536 merupakan penghasil enzim L-asparaginase yang terbaik apabila media fermentasi ditambah dengan sisa tulang ayam yang telah dimasak yang tinggi kandungan proteinnya.

ABSTRACT

L-asparaginase is highly demanded in pharmaceutical, food and biosensor industry due to its remarkable properties in hydrolysing L-asparagine into aspartic acid and ammonia. Owing to this significant property, L-asparaginase is widely produced through microbial fermentation. This research aims to produce L-asparaginase enzyme through microbial fermentation by the most potent isolate in the presence of chicken bone wastes as the substrate. Therefore, this research can be categorised into 3 parts where the first part was to perform several studies in determining the best enzyme producer and substrate while the second part was the effect of process parameters using One-Factor-at-a-Time (OFAT) method and the third part was the purification and characterisation study of L-asparaginase. Six millilitres (5×10^8 cells/ml) of *Escherichia coli* ATCC 10536 is the optimum inoculum size in 50 ml of pH 9 growth media at 40 °C for 2 days. In addition to that, *Escherichia coli* ATCC 10536 was fully enhanced when it was engaged with 1.0 % w/v of starch, 0.2 % w/v of ammonium chloride and 1 % w/v of cooked chicken bone (CCB) as the carbon source, nitrogen source and substrate respectively. The effect of process parameters studied using OFAT method revealed that *Escherichia coli* ATCC 10536 is the most potent L-asparaginase producer in this research and it preferred nutrient broth as the growth media in producing L-asparaginase. Furthermore, *Escherichia coli* ATCC 10536 L-asparaginase was purified by ammonium sulfate precipitation, dialysis and DEAE-cellulose chromatography. At the end of purification, 0.42 % yield of L-asparaginase was obtained. In addition, the dialysed ammonium sulfate fraction of L-asparaginase was partially characterised which resulted to 40 °C, pH 8, Na⁺ and ethylenediaminetetraacetic acid (EDTA) being the optimum incubation temperature, pH, metal ion and inhibitor respectively. In conclusion, *Escherichia coli* ATCC 10536 is an excellent L-asparaginase producer when the fermentation is supplemented with high protein content of cooked chicken bone as substrate.

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