



DOI URL:

ADVANCED RESEARCH (IJAR)

<http://dx.doi.org/10.21474/IJAR01/xxx>

RESEARCH ARTICLE

**CHARACTERIZATION OF TWO CHALCONE DERIVATIVES ISOLATED FROM FINGER
ROOT WITH NUTRACEUTICAL POTENTIALS**

**IBRAHIM AWAD MOHAMMED*¹, MUHAMMAD NADEEM AKHTAR¹, HUSSAM DAWOOD
ABDULLAH², HAZRULRIZAWATI ABD HAMID¹**

AFFILIATION:

¹ Faculty of Industrial Sciences and Technology, Universiti Malaysia Pahang .26300
Gambang. Malaysia

² Department of Chemistry College of Education Sciences, Tikrit Universiti. Tikrit. Iraq

*Correspondence Author: ibraheemawad11@yahoo.com

Manuscript Info

Manuscript History

Received: xxxxxxxxxxxxxxxxx

Final Accepted: xxxxxxxxxxxxx

Published: xxxxxxxxxxxxxxxxx

Key words:-

Pinostrobin chalcone

Cardamone

UV-Vis photometry

FTIR

¹HNMR

Abstract *Boesenbergia rotunda* is an example of medicinal herbal plants which has been traditionally employed in the treatment of many life-threatening ailments such as diuretic, dysentery, inflammation, aphrodisiac and gastrointestinal disorder. In this study the two chalcone derivatives were isolated from the root of *B. rotunda* using column-thin chromatography and characterized using different spectroscopy methods such as UV-VIS, FTIR, and ¹HNMR. The bioactive compound identified

were pinostrobin chalcone (1) and cardamone (2). These IR spectra, UV-vis photometry analysis and ¹HNMR suggested that the chemical constituent isolated to be a flavonoid derivative, which is similar to the

previous studies. The result of the study suggests that *B. rotunda* rhizome has a potential in drug and nutraceutical applications.

Introduction:-

The increasing demands for chemical diversity in screening characterizations necessitate succinct investigations into the therapeutic activities of herbal plant bio-products. The finger root (*Beosenbergia rotunda*) is an example of herb plants belonging to the family of *Zingiberaceae*. It belongs to the order of Zingiberales herbaceous ground flora plants in the rainforest and is commonly in countries like Thailand, Malaysia, Indonesia and Myanmar (Cheenpracha et al., 2006). There are approximately 150 species of this family with about 23 species found in Peninsular Malaysia (Sukara et al., 2017). The finger-root of *B. rotunda* has been reported for the treatment of peptic-ulcer, colic, oral diseases, urinary disorders, dysentery and inflammation (Chatsumpun et al., 2017). These numerous medicinal benefits are to the presence of some important bioactive components. There is therefore a necessity to isolate these important bioactive constituents at higher purity. In this study, two bioactive constituents from *B. rotunda* were purified and isolated using column chromatography techniques. The extracted metabolites were characterized using FTIR, NMR, and UV-Vis techniques.

Materials and methods:-

Chemical preparation of Pinostrobin Chalcone

The finger roots (*B. rotunda*) were procured from Selangor Darul Ehsan, Malaysia. The plant sample was identified and authenticated by Assoc. Prof. Dr. Muhammad Nadeem Akhtar at the Faculty of Industrial Sciences and Technology, University Malaysia Pahang. The sample was then dried and grounded into smaller size (0.1 mm) using a Grindomix grinder (GM-200 model, Germany). The extraction process was conducted with 300 g of the sample and methanol solvent using Soxhlet apparatus. The product of extraction was placed on water-bath at 65 °C to remove the residual methanol. The residue was then treated with 5% HCl and filtered. The final solution was acidified using HCl(aq) and Meyers reagent/Dragendorff's reagent until precipitation stops. The resulting precipitates was filtered, wash with water and suspended in MeOH-Me₂CO-H₂O (6:2:1) (Isa et al., 2012). A complex mixture was resolved by vacuum column chromatography (VLC) with silica gel 60, packed in slurry with chloroform as the initial solvent and gradually increased the polarity of a solvent. Chromatography separation was performed on pre-coated TLC plates Kieselgel Si 60; 0.25mm (E. Merck, Darmstadt, Germany). The separated spots were detected under UV light (256 & 366 nm). The individual components was purified by preparing TLC on a pre-coated silica-gel plates using MeOH-CHCl₃. Column Chromatography was performed on a pre-coated TLC plates Kieselgel Si-60; 0.25mm (E. Merck, Darmstadt, Germany).

Characterization Techniques

The *B. rotunda* extracts and the isolated compounds were characterized using Fourier transform infrared spectroscopy (FTIR), Nuclear Magnetic Resonance (NMR), UV-Vis and GCMS as discussed in the proceeding sections.

Ultraviolet-Visible photometry analysis

The Ultraviolet-Visible photometry analysis was conducted using the the Genesys 10s UV-Vis spectrometer. Prior to the analysis the solid single pure compound isolated was first diluted with acetone which produced a pale yellowish liquid solution. The liquid solution was then pipetted into the 1-cm path length quartz cuvettes. The preparation of the sample and the analysis was carried out in the dark at 21 ± 1 °C. To prevent the evaporation of the solvent (acetone), the reactor was closed and stirred. The solution was irradiated at the wavelength ranged of 200 to 500 nm. The absorption spectrum was displayed by the spectrometer and the data was obtained. The wavelength with maximum absorbance of the single pure compound was then identified.

Fourier Transform Infrared (FTIR) analysis

Fourier transform infrared spectroscopy (FTIR) was carried out on the extracts to determine the functional groups (Ojalere et al., 2017; Amani et al., 2018). A Thermo-Nicolet spectrometer (iS5 iD7 ATR, Germany) equipped with OMNIC software was employed in the spectrometry analysis. The analysis was executed using the conventional KBr standard procedure with wavenumber ranging from $4000-600\text{ cm}^{-1}$. Under this study, the spectrum of the observed bond and associated group frequencies were compared with the table of expected absorption bands (Carol 2000).

Nuclear Magnetic Resonance Spectroscopy (NMR)

Further confirmations of the structure of the chemical constituent of the isolated compounds were performed using ^1H NMR. The sample was diluted be diluted in deuterated chloroform (CDCl_3) and then transferred to NMR tube. Based on the NMR spectra, the area under each pattern obtained from the integration of the signal was studied to determine and identify the structure of the compound.

Results and discussion

UV-Vis Photometry studies

The pinostrobin chalcone (1) was absorbed by UV light wavelength range of 250-380 nm. The observed wavelength range corresponds to the conjugated α , β -unsaturated carbonyl ($\text{C}=\text{O}$) chromophore as reported [2]. Figure 1 shows the UV-visible spectrum of pinostrobin chalcone with maximum absorption of 340 nm at wavelength, λ_{max} .

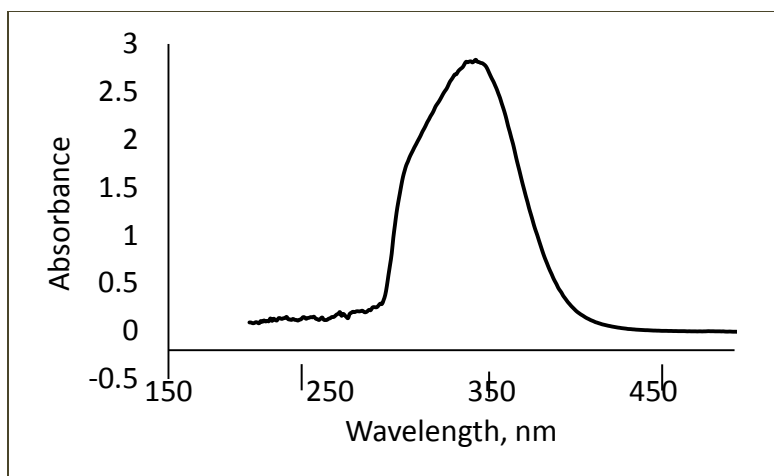


Figure 1 UV spectrum of pure pinostrobin chalcone (1)

However, cardamone (2) absorbed UV light at a wavelength range of 290-380 nm, which corresponds to the conjugated α , β -unsaturated carbonyl (C=O) chromophore [5]. Figure 2 shows the UV-visible spectrum of cardamone (2) with maximum absorption at wavelength, λ_{\max} of 340 nm.

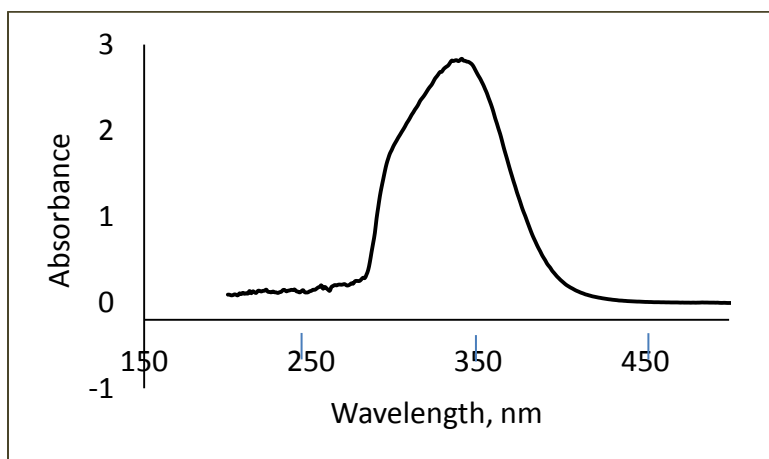


Figure 2 UV spectrum of cardamone (2)

Functional group analysis

Figure 3 shows the IR spectrum of pinostrobin chalcone (1) with an absorption band of 3456 cm^{-1} which corresponds to the hydroxy (O-H). Moreover, the observed absorption band between 2940 and 3092 cm^{-1} corresponds to the aromatic C-H stretching. Absorption band at 1623 cm^{-1} and between 1416 - 1439 cm^{-1} was indicated the presence of carbonyl (C=O) and aromatic ring C=C, respectively. Absorption band appeared in the range of 1158 - 1219 cm^{-1} corresponded to (C-O) [1].

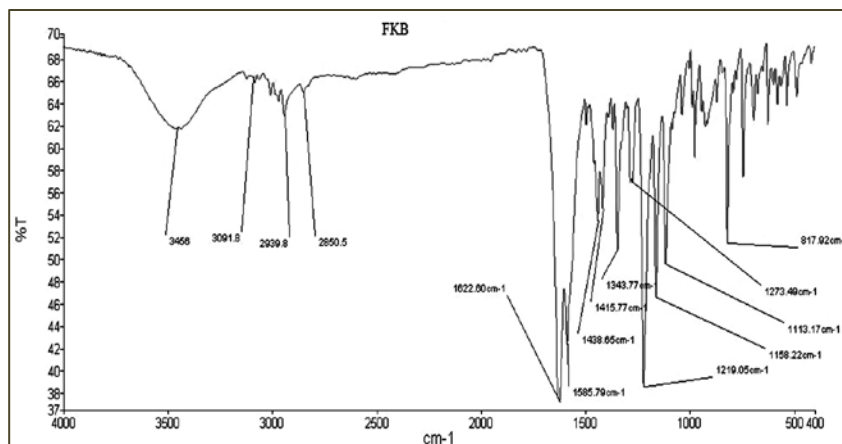


Figure 3 IR spectrum of pure pinostrobin chalcone (1)

IR spectrum of cardamone (2) in Figure 4 shows absorption band at 3456 cm^{-1} which corresponded to hydroxy (O-H). The absorption band between $2940\text{--}3092\text{ cm}^{-1}$ corresponds to the aromatic C-H stretch while the absorption band at 1623 cm^{-1} and between $1416\text{--}1439\text{ cm}^{-1}$ indicated the presence of carbonyl (C=O) and aromatic ring C=C, respectively. Absorption band appeared in the range of $1158\text{--}1219\text{ cm}^{-1}$ corresponded to (C-O) moiety [2].

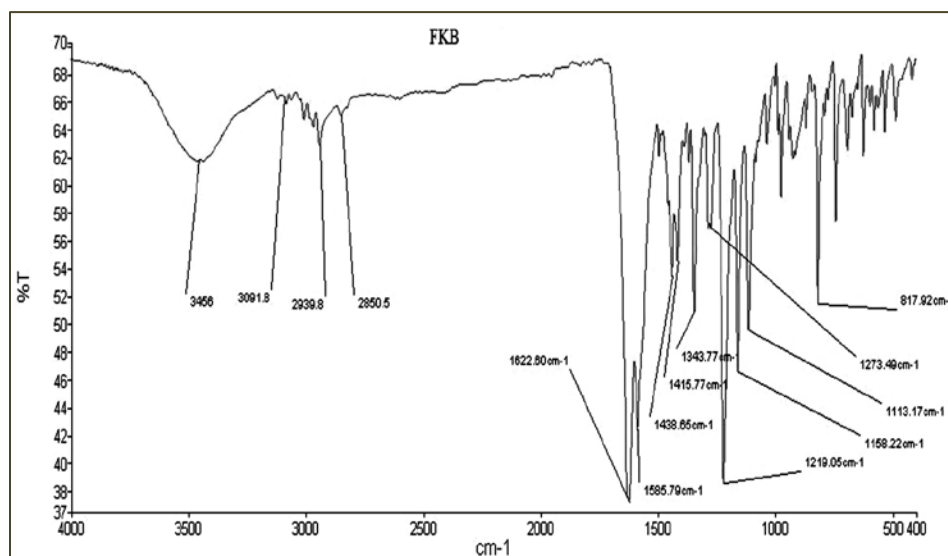


Figure 4 IR spectrum of cardamone (2)

^1H Nuclear Magnetic Resonance (NMR)

The ^1H -NMR spectrum (600 MHz, CDCl_3) of pinostrobin chalcone (1) in Figure 5 shows a singlet at δ 3.84, which was assigned to the methoxy protons at C-4'. Two doublet that appeared at 5.96 (d, $J = 2.46\text{ Hz}$) and 6.11 (d, $J = 2.40\text{ Hz}$) were assigned to C-5' and C-3' protons, respectively.

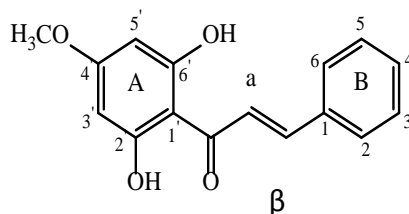


Figure 5 Structure of pure pinostrobin chalcone (2)

A broad multiplet that appeared in the range of 7.38-7.42 was assigned to C-3, C-4 and C-5 protons. Two doublets observed at 7.62 (d, $J = 8.04$ Hz) and 7.60 (d, $J = 8.04$ Hz) were assigned to C-2 and C-6 protons, respectively. A downfield doublet at 7.78 (d, $J = 15.55$ Hz) was assigned to proton at α -carbon and another doublet observed at 7.90 (d, $J = 15.55$ Hz) was assigned to proton at β -carbon, which data was supported by previous work [1,2]. A downfield singlet that appeared at 14.30 was assigned to C-2' hydroxyl proton chelated to carbonyl group. Further details of $^1\text{H-NMR}$ and $^{13}\text{C-NMR}$ spectra are shown in Table 1.

Table 1 NMR data of pure pinostrobin chalcone (2)

Carbon	^{13}C (δ)	^1H (δ)	Multiplicity	Designation
1'	108.14	-	-	-
2'	168.90	-	-	-
3'	93.60	6.11	(d, $J = 2.40$ Hz, 1H)	C3'-H
4'	166.70	-	-	-
5'	90.94	5.96	(d, $J = 2.46$ Hz, 1H)	C5'-H
6'	162.50	-	-	-
1	132.90	-	-	-
2	130.20	7.62	(d, $J = 8.04$ Hz, 1H)	C2-H
3	128.10	7.38-7.42	(m, 3H)	C3-H
4	141.00	7.38-7.42	(m, 3H)	C4-H
5	128.10	7.38-7.42	(m, 3H)	C5-H
6	130.20	7.60	(d, $J = 8.04$ Hz, 1H)	C6-H
A	126.60	7.78	(d, $J = 15.55$ Hz, 1H, H- α)	C α -H
B	142.51	7.90	(d, $J = 15.55$ Hz, 1H, H- β)	C β -H
OCH ₃	55.70	3.84	(s, 3H)	OCH ₃ (C4')
OH	-	14.30	(s, 1H)	OH (C2')
OH(C6')				
C=O	192.22	-	-	-

Moreover, the The $^1\text{H-NMR}$ spectrum (600 MHz, CDCl_3) of cardamone (2) in Figure 6 shows a singlet at δ 3.84, which was assigned to the methoxy protons at C-6' atom.

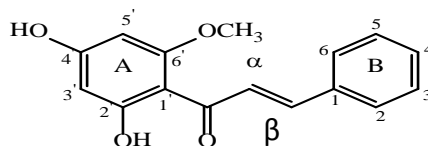


Figure 6 Structure of pure Cardamone (2)

Two doublet that appeared at 5.96 (d, $J = 2.46$ Hz) and 6.11 (d, $J = 2.40$ Hz) were assigned to C-5' and C-3' protons, respectively. A broad multiplet that appeared in the range of 7.38-7.42 was assigned to C-3, C-4 and C-5 protons. Two doublets observed at 7.62 (d, $J = 8.04$ Hz) and 7.60 (d, $J = 8.04$ Hz) were assigned to C-2 and C-6 protons, respectively. A downfield doublet at 7.78 (d, $J = 15.55$ Hz) was assigned to proton at α -carbon and another doublet observed at 7.90 (d, $J = 15.55$ Hz) was assigned to proton at β -carbon, which data is supported by previous work [2,3]. A downfield singlet that appeared at 14.30 was assigned to C-2' hydroxyl proton chelated to carbonyl group. Further details of $^1\text{H-NMR}$ and $^{13}\text{C-NMR}$ spectra are shown in Table 2.

Table 2 NMR data of Cardamone (2)

Carbon	^{13}C (δ)	^1H (δ)	Multiplicity	Designation
1'	108.14	-	-	-
2'	168.90	-	-	-
3'	93.60	6.11	(d, $J = 2.40$ Hz, 1H)	C3'-H
4'	166.70	-	-	-
5'	90.94	5.96	(d, $J = 2.46$ Hz, 1H)	C5'-H
6'	162.50	-	-	-
1	132.90	-	-	-
2	130.20	7.62	(d, $J = 8.04$ Hz, 1H)	C2-H
3	128.10	7.38-7.42	(m, 3H)	C3-H
4	141.00	7.38-7.42	(m, 3H)	C4-H
5	128.10	7.38-7.42	(m, 3H)	C5-H
6	130.20	7.60	(d, $J = 8.04$ Hz, 1H)	C6-H
A	126.60	7.78	(d, $J = 15.55$ Hz, 1H, H- α)	C α -H
B	142.51	7.90	(d, $J = 15.55$ Hz, 1H, H- β)	C β -H
OH (C4')	-	-	-	OH (C4')
OCH ₃ (C6')	55.53	3.92	(s, 3H)	OCH ₃ (C6')
OH (C2')	-	14.30	(s, 1H)	OH (C2')
C=O	193.20	-	-	-

Conclusions

The characterizations of two chalcone derivatives were conducted using UV-VIS, FTIR, and $^1\text{H-NMR}$. The UV-VIS helped in the identification of pinostrobin chalcone (1) and cardamone (2)

by means of its absorption spectra. FTIR revealed the functional groups present to further elucidate and confirm the presence of the two chalcone derivatives in *B. rotunda*. ¹H NMR resolved the magnetical dissimilar proton found in the compound isolated. The result obtained from this study indicated the potential of *B. rotunda* rhizome extracts as drug candidate in therapeutic applications and pharmaceutical industries.

Acknowledgements

We acknowledge the support of the Faculty of Industrial Sciences and Technology, Universiti Malaysia Pahang during the course of this research.

References

- [1] Cheenpracha, S., Karalai, C., Ponglimanont, C., Subhadhirasakul, S., and Tewtrakul, S.(2006).Anti-HIV-1 protease activity of compounds from *Boesenbergia pandurata*, J. Bioorg. & Med. Chem. 14: 1710–1714.
- [2] Sukari, M.A., Ching, A.Y., Lian, G.E., Rahmani, M., Khalid, K. (2017). Cytotoxic Constituents from *Boesenbergia pandurata* (Roxb.) Schltr. Nat. Prod. Sci., 13(2): 110-113.
- [3] Chatsumpun, N., Sritularak, B., Likhitwitayawuid, K. (2017). New Biflavonoids with α -Glucosidase and Pancreatic Lipase Inhibitory Activities from *Boesenbergia rotunda*. Molecules. 22(11):862.
- [4] Isa, N.M., Abdelwahab, S.I., Mohan, S., Abdul, A.B., Sukari, M,A., Taha, M.M., Syam, S., Narrima, P., Cheah, S.C., Ahmad, S., Mustafa M.R.(2012). In vitro anti-inflammatory, cytotoxic and antioxidant activities of *boesenbergia A*, a chalcone isolated from *Boesenbergia rotunda* (L.) (fingerroot). Brazilian J. Med. & Bio. Res., 45(6): 524-530.
- [5] Olalere, O.A., Abdurahman, N.H., Alara, O.R. (2017). Extraction, radical scavenging activities and physicochemical fingerprints of black pepper (*Piper nigrum*) extract. J. Food Measur. Charac., 11(4):2195–201.
- [6] Amani, A., Olalere O.A., AbdElhafiz E. A., Abdurahman H.N., Yunus, R.M., Ghada, M.I., Nassereldeen, A.K. (2018). Comparative Analysis of Polyphenolic and Antioxidant Constituents in Dried Seedlings and Seedless *Acacia nilotica* Fruits. J. Analy. & Testing, 12,

Corresponding Author:- Ibrahim Awad Mohammed

Address:- Faculty of Industrial Sciences and Technology, Universiti Malaysia Pahang .26300 Gambang. Malaysia