

**PRODUCTION OF RENEWABLE GLUCOSE
FROM OIL PALM FROND BAGASSE BY
USING SACCHARISEB C6 THROUGH
ENZYMATIC HYDROLYSIS**

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STUDENT'S DECLARATION

I hereby declare that the work in this thesis is based on my original work except for quotations and citations which have been duly acknowledged. I also declare that it has not been previously or concurrently submitted for any other degree at Universiti Malaysia Pahang or any other institutions.

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LIST OF SYMBOLS

A ₁	Weight of crucibles
A ₂	Initial weight of OPF bagasse
A ₃	Final weight of crucibles and ash
E ₁	Weight of fiberglass filter
E ₂	Weight of wet biomass added to extraction cell
E ₃	Weight of oven-dry extracted biomass and filter
E ₄	Volume of water extract
E _{5, Glu}	Glucose content in HPLC standard
E ₆	Concentration of monomer sugar after acid hydrolysis
E ₇	Concentration of monomer sugar before acid hydrolysis
E ₈	Water extract samples solutions using HPLC
EE	Ethanol extractives
ET	Total extractives
H ₂	Mass of monomer sugar after acid hydrolysis
H ₅	Concentration of monomer sugars after acid hydrolysis
I _{am}	Peak intensity of the amorphous phase
I ₀₀₂	Peak intensity of the 002 crystal plane
L ₁	Weight of filter paper prior to filtration
L ₂	Final weight of the residue and filter paper
M	Mass
MC _{TWB}	Moisture content
MW _{EXT}	Moisture content of oven-dry extracted biomass and filter
SC	Structural carbohydrate
SS _{Glu}	Glucose soluble sugar
V	Volume
W	Weight

LIST OF ABBREVIATIONS

AFEX	Ammonia fiber explosion
ANOVA	Analysis of variance
CCD	Central composite design
DOE	Design of experiments
DP	Degree of polymerization
EFB	Empty fruit bunch
FTIR	Fourier transform infrared
HMF	5-hydroxymethyl-furfural
HPLC	High performance liquid chromatography
LCB	Lignocellulosic biomass
LHW	Liquid hot water
MPOB	Malaysian palm oil board
NREL	National renewable energy laboratory
OFAT	One-factor-at-a-time
OPF	Oil palm frond
OPT	Oil palm tree
PPS	Palm pressed fiber
RSM	Response surface methodology
SEM	Scanning electron microscopy
XRD	X-ray diffraction

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ABSTRAK

Biojisim lignoselulosa (LCB) adalah biojisim yang paling banyak diperbaharui daripada sumber-sumber yang berpotensi tinggi untuk pengeluaran pelbagai jenis produk yang berfaedah. Terkini, kelapa sawit telah dikenal pasti sebagai sumber LCB yang paling berpotensi dan boleh digunakan untuk penghasilan gula. Biasanya, sisa buangan daripada tanaman kelapa sawit digunakan sebagai makanan haiwan tetapi ia bukanlah kaedah yang terbaik dari segi ekonomi dalam memanipulasi penggunaan sisa buangan. Selain itu, kewujudan sisa buangan menimbulkan masalah dari segi pelupusan dan sering dilupuskan melalui pembakaran secara terbuka dan mengakibatkan kepada pencemaran alam sekitar. Oleh itu, hampas pelelah kelapa sawit (OPF) diperkenalkan sebagai bahan mentah dalam kajian ini untuk memaksimumkan penggunaan sisa kelapa sawit. Objektif kajian ini adalah untuk mengenalpasti komponen yang terdapat di dalam hampas OPF sebelum dan selepas pra-rawatan alkali dan untuk menyaring dan mengoptimumkan faktor-faktor yang memberi kesan terhadap penguraian enzim menggunakan Sacchariseb C6 bagi penghasilan gula. Hampas OPF ditelah melalui pra-rawatan alkali dan natrium hidroksida digunakan sebagai pelarut sebelum meneruskan proses rawatan penguraian enzim dengan menggunakan Sacchariseb C6. Semasa proses pra-rawatan, struktur biojisim selulosa akan diubah dan delignifikasi telah berlaku yang membuatkan struktur selulosa lebih terbuka untuk proses seterusnya iaitu rawatan penguraian enzim dengan menukar selulosa kepada gula. Kaedah gerak balas permukaan (RSM) telah digunakan untuk menyaring dan mengoptimumkan keadaan penguraian enzim. Pencirian hampas OPF mentah yang dihasilkan adalah sebanyak 40.7% glukosa, 26.1% xylan, 4.5% ekstraktif, 26.2% lignin dan 1.8% abu. Sementara itu, bagi hampas OPF yang telah melalui pra-rawatan mengandungi 61.4% glukosa, 20.4% xylan, 0.3% ekstraktif, 13.3% lignin dan 1.3% abu. Daripada kajian analisis faktorial, penguraian enzim yang terbaik telah menghasilkan 33.01 ± 0.73 g / L glukosa pada kelajuan kisaran 200 rpm, 60 FPU/g pengisian enzim, 4% (w/v) pengisian glucan, suhu pada 55 °C dan 72 jam masa tindak balas. Dalam reka bentuk komposit pusat (CCD), keadaan optimum bagi rawatan penguraian enzim diperolehi pada 50 °C untuk 87.93 jam masa penguraian yang menghasilkan 41.11 ± 0.11 g/L glukosa. Secara keseluruhannya, rawatan penguraian enzim Sacchariseb C6 keatas hampas OPF berpotensi menghasilkan glukosa di mana glukosa ini boleh digunakan dalam pelbagai industri bagi menghasilkan produk yang bernilai.

ABSTRACT

Lignocellulosic biomass (LCB) is the most abundant renewable biomass that gives high potential source in production of various beneficial products. Recently, oil palm crops are known as the most potential LCB which can be employed for sugar production. Normally, wastes from oil palm crops are used as animal feed but this is not the ideal economically valuable method of manipulating the wastes. Besides, the existence of wastes created disposal problems and often been disposed off by open burning that may lead to the environmental pollution. Therefore, oil palm frond (OPF) bagasse was introduced as a raw material in this study to maximize the utilization of oil palm waste. This study aims to characterize the composition of OPF bagasse before and after alkaline pretreatment and to screen and optimize the factors affecting enzymatic hydrolysis by using Sacchariseb C6 for glucose production. OPF bagasse was treated using alkaline pretreatment and sodium hydroxide used as a solvent before proceeding with enzymatic hydrolysis using Sacchariseb C6. During pretreatment process, cellulosic biomass structure will be altered and delignification occurred which make cellulose more accessible to the subsequent enzymatic hydrolysis process by converting it into simple sugars. Response Surface Methodology (RSM) was employed to screen and optimize the enzymatic hydrolysis condition. Characterization of raw OPF bagasse produced 40.7 % glucan, 26.1 % xylan, 4.5 % extractives, 26.2 % lignin and 1.8 % ash. Meanwhile, pre-treated OPF bagasse composed of 61.4 % glucan, 20.4 % xylan, 0.3 % extractives, 13.3 % lignin and 1.3 % ash. In factorial analysis study, the best enzymatic hydrolysis condition yielded 33.01 ± 0.73 g/L of glucose when performed at 200 rpm of agitation speed, 60 FPU/g of enzyme loading, 4% (w/v) of glucan loading, temperature at 55 °C and 72 hours of reaction time. In central composite design (CCD), the optimum condition for enzymatic hydrolysis was obtained at 50 °C for 87.93 hours of hydrolysis time which produced 41.11 ± 0.11 g/L of glucose. Overall, enzymatic hydrolysis of OPF bagasse by using Sacchariseb C6 has high potential for production of glucose which later can be utilized for various industrial application to produce valuable value-added products.

REFERENCES

- Abdul, P. M., Jahim, J. M., Harun, S., Markom, M., Lutpi, N. A., Hassan, O., Mohd Nor, M. T. (2016). Effects of changes in chemical and structural characteristic of ammonia fibre expansion (AFEX) pretreated oil palm empty fruit bunch fibre on enzymatic saccharification and fermentability for biohydrogen. *Bioresource Technology*, 211, 200–208. <http://doi.org/10.1016/j.biortech.2016.02.135>
- Abdul Khalil, H. P. S., Firdaus, M. Y. N., Jawaid, M., Anis, M., Ridzuan, R., & Mohamed, A. R. (2010). Development and material properties of new hybrid medium density fibreboard from empty fruit bunch and rubberwood. *Materials and Design*, 31(9), 4229–4236. <http://doi.org/10.1016/j.matdes.2010.04.014>
- Abdul Khalil, H. P. S., Nur Firdaus, M. Y., Anis, M., & Ridzuan, R. (2008). The Effect of Storage Time and Humidity on Mechanical and Physical Properties of Medium Density Fiberboard (MDF) from Oil Palm Empty Fruit Bunch and Rubberwood. *Polymer-Plastics Technology and Engineering*, 47(10), 1046–1053. <http://doi.org/10.1080/03602550802355644>
- Abdul Khalil, H. P. S., Nurul Fazita, M. R., Bhat, A. H., Jawaid, M., & Nik Fuad, N. A. (2010). Development and material properties of new hybrid plywood from oil palm biomass. *Materials and Design*, 31(1), 417–424. <http://doi.org/10.1016/j.matdes.2009.05.040>
- Abdullah, N., & Sulaiman, F. (2013). The oil palm wastes in Malaysia. *Biomass Now – Sustainable Growth and Use*, 75–100. <http://doi.org/10.5772/55302>
- Akpınar, O., Erdogan, K., Bakır, U., & Yilmaz, L. (2010). Comparison of acid and enzymatic hydrolysis of tobacco stalk xylan for preparation of xylooligosaccharides. *LWT - Food Science and Technology*, 43(1), 119–125. <http://doi.org/10.1016/j.lwt.2009.06.025>
- Alvira, P., Tomas-Pejo, E., Ballesteros, M., & Negro, M. J. (2010). Pretreatment technologies for an efficient bioethanol production process based on enzymatic hydrolysis: A review. *Bioresource Technology*, 101(13), 4851–4861. <http://doi.org/10.1016/j.biortech.2009.11.093>
- Anwar, Z., Gulfraz, M., & Irshad, M. (2014). Agro-industrial lignocellulosic biomass a key to unlock the future bio-energy: A brief review. *Journal of Radiation Research and Applied Sciences*, 7(2), 163–173. <http://doi.org/10.1016/j.jrras.2014.02.003>
- Bajpai, P. (2016). Pretreatment of Lignocellulosic Biomass for Biofuel Production. *SpringerBriefs in Green Chemistry for Sustainability*, 34, 86. <http://doi.org/10.1007/978-981-10-0687-6>
- Barkoula, N. M., Alcock, B., Cabrera, N. O., & Peijs, T. (2008). Fatigue properties of highly oriented polypropylene tapes and all-polypropylene composites. *Polymers and Polymer*

Composites, 16(2), 101–113. <http://doi.org/10.1002/pc>

Beilen, J. B. van, & Li, Z. (2002). Enzyme technology: An overview. *Current Opinion in Biotechnology*, 13(4), 338–344. [http://doi.org/10.1016/S0958-1669\(02\)00334-8](http://doi.org/10.1016/S0958-1669(02)00334-8)

Bezerra, M. A., Santelli, R. E., Oliveira, E. P., Villar, L. S., & Escaleira, L. A. (2008). Response surface methodology (RSM) as a tool for optimization in analytical chemistry. *Talanta*, 76(5), 965–977. <http://doi.org/10.1016/j.talanta.2008.05.019>

Bhaumik, P., & Dhepe, P. L. (2015). Conversion of Biomass into Sugars. In *Biomass Sugars for Non-fuel Application* (pp. 27–28).

Bidlack, J., Malone, M., & Benson, R. (1992). Molecular Structure and Component Integration of Secondary Cell Walls in Plants. *Proceedings of the Oklahoma Academy of Science*, 72, 51–56.

Binod, P., Janu, K. U., Sindhu, R., & Pandey, A. (2011). *Hydrolysis of Lignocellulosic Biomass for Bioethanol Production*. *Biofuels* (1st ed.). Elsevier Inc. <http://doi.org/10.1016/B978-0-12-385099-7.00010-3>

Binod, P., Satyanagalakshmi, K., Sindhu, R., Janu, K. U., Sukumaran, R. K., & Pandey, A. (2012). Short duration microwave assisted pretreatment enhances the enzymatic saccharification and fermentable sugar yield from sugarcane bagasse. *Renewable Energy*, 37(1), 109–116. <http://doi.org/10.1016/j.renene.2011.06.007>

Bouza, R. J., Gu, Z., & Evans, J. H. (2016). Screening conditions for acid pretreatment and enzymatic hydrolysis of empty fruit bunches. *Industrial Crops and Products*, 84, 67–71. <http://doi.org/10.1016/j.indcrop.2016.01.041>

Bugg, T. D., & Rahmanpour, R. (2015). Enzymatic conversion of lignin into renewable chemicals. *Current Opinion in Chemical Biology*, 29, 10–17. <http://doi.org/10.1016/j.cbpa.2015.06.009>

Cara, C., Moya, M., Ballesteros, I., Negro, M. J., González, A., & Ruiz, E. (2007). Influence of solid loading on enzymatic hydrolysis of steam exploded or liquid hot water pretreated olive tree biomass. *Process Biochemistry*, 42(6), 1003–1009. <http://doi.org/10.1016/j.procbio.2007.03.012>

Çetin, N. S., & Özmen, N. (2002). Use of organosolv lignin in phenol-formaldehyde resins for particleboard production: I. Organosolv lignin modified resins. *International Journal of Adhesion and Adhesives*, 22(6), 477–480. [http://doi.org/10.1016/S0143-7496\(02\)00058-1](http://doi.org/10.1016/S0143-7496(02)00058-1)

Chakar, F. S., & Ragauskas, A. J. (2004). Review of current and future softwood kraft lignin process chemistry. In *Industrial Crops and Products* (Vol. 20, pp. 131–141). <http://doi.org/10.1016/j.indcrop.2004.04.016>

- Cheng, J., Su, H., Zhou, J., Song, W., & Cen, K. (2011). Microwave-assisted alkali pretreatment of rice straw to promote enzymatic hydrolysis and hydrogen production in dark- and photo-fermentation. *International Journal of Hydrogen Energy*, 36(3), 2093–2101. <http://doi.org/10.1016/j.ijhydene.2010.11.021>
- Cherry, J. R., & Fidantsef, A. L. (2003). Directed evolution of industrial enzymes: An update. *Current Opinion in Biotechnology*, 14(4), 438–443. [http://doi.org/10.1016/S0958-1669\(03\)00099-5](http://doi.org/10.1016/S0958-1669(03)00099-5)
- Chu, Q., Li, X., Ma, B., Xu, Y., Ouyang, J., Zhu, J., Yong, Q. (2012). Bioethanol production: An integrated process of low substrate loading hydrolysis-high sugars liquid fermentation and solid state fermentation of enzymatic hydrolysis residue. *Bioresource Technology* (Vol. 123).
- Chundawat, S. P. S., Donohoe, B. S., da Costa Sousa, L., Elder, T., Agarwal, U. P., Lu, F., Dale, B. E. (2011). Multi-scale visualization and characterization of lignocellulosic plant cell wall deconstruction during thermochemical pretreatment. *Energy & Environmental Science*, 4(3), 973. <http://doi.org/10.1039/c0ee00574f>
- da Silva, A. S., Teixeira, R. S. S., Moutta, R. D. O., Ferreira-Leitão, V. S., de Barros, R. D. R. O., Ferrara, M. A., & Bon, E. P. D. S. (2013). Sugarcane and woody biomass pretreatments for ethanol production. In *Sustainable Degradation of Lignocellulosic Biomass - Techniques, Applications and Commercialization* (pp. 47–88). Intech. <http://doi.org/10.5772/53378>
- Dahlan, I. (2000). Oil Palm Frond , a Feed for Herbivores. *Asian Australasian Journal of Animal Sciences*, 13(July 2000), 300–303.
- Diniz, F. M., & Martin, A. M. (1996). Use of response surface methodology to describe the combined effects of pH, temperature and E/S ratio on the hydrolysis of dogfish (*Squalus acanthias*) muscle. *International Journal of Food Science and Technology*, 31(5), 419–426. <http://doi.org/10.1046/j.1365-2621.1996.00351.x>
- Du, J., Cao, Y., Liu, G., Zhao, J., Li, X., & Qu, Y. (2017). Identifying and overcoming the effect of mass transfer limitation on decreased yield in enzymatic hydrolysis of lignocellulose at high solid concentrations. *Bioresource Technology*, 229, 88–95. <http://doi.org/10.1016/j.biortech.2017.01.011>
- Ebrahimi, M., Rajion, M. a., Goh, Y. M., Sazili, a. Q., Soleimani, a. F., & Schonewille, J. T. (2013). Oil Palm (*Elaeis guineensis Jacq.*) Frond Feeding of Goats in the Humid Tropics. *Journal of Animal and Veterinary Advances*.
- Eklund, R., Galbe, M., & Zacchi, G. (1990). Optimization of temperature and enzyme concentration in the enzymatic saccharification of steam-pretreated willow. *Enzyme and Microbial Technology*, 12(3), 225–228. [http://doi.org/10.1016/0141-0229\(90\)90043-P](http://doi.org/10.1016/0141-0229(90)90043-P)

- Emtiazi, G., Naghavi, N., & Bordbar, A. (2001). Biodegradation of lignocellulosic waste by *Aspergillus terreus*. *Biodegradation*, 12(4), 259–263.
<http://doi.org/10.1023/a:1013155621336>
- Eriksson, T., Börjesson, J., & Tjerneld, F. (2002). Mechanism of surfactant effect in enzymatic hydrolysis of lignocellulose. *Enzyme and Microbial Technology*, 31(3), 353–364.
[http://doi.org/10.1016/S0141-0229\(02\)00134-5](http://doi.org/10.1016/S0141-0229(02)00134-5)
- Fang, H., Zhao, C., & Song, X. Y. (2010). Optimization of enzymatic hydrolysis of steam-exploded corn stover by two approaches: Response surface methodology or using cellulase from mixed cultures of *Trichoderma reesei* RUT-C30 and *Aspergillus niger* NL02. *Bioresource Technology*, 101(11), 4111–4119.
<http://doi.org/10.1016/j.biortech.2010.01.078>
- Ferreira, S., Duarte, A. P., Ribeiro, M. H. L., Queiroz, J. A., & Domingues, F. C. (2009). Response surface optimization of enzymatic hydrolysis of *Cistus ladanifer* and *Cytisus striatus* for bioethanol production. *Biochemical Engineering Journal*, 45(3), 192–200.
<http://doi.org/10.1016/j.bej.2009.03.012>
- Gabhan, J., William, S., Vaidya, A., Anand, D., & Wate, S. (2014). Pretreatment of garden biomass by alkali-assisted ultrasonication: effects on enzymatic hydrolysis and ultrastructural changes. *Journal of Environmental Health Science and Engineering*, 12(1), 76. <http://doi.org/10.1186/2052-336X-12-76>
- Gama, R., Van Dyk, J. S., & Pletschke, B. I. (2015). Optimisation of enzymatic hydrolysis of apple pomace for production of biofuel and biorefinery chemicals using commercial enzymes. *3 Biotech*, 5(6), 1075–1087. <http://doi.org/10.1007/s13205-015-0312-7>
- Ghaffar, S. H., & Fan, M. (2013). Structural analysis for lignin characteristics in biomass straw. *Biomass and Bioenergy*, 57, 264–279. <http://doi.org/10.1016/j.biombioe.2013.07.015>
- Ghaffar, S. H., & Fan, M. (2014). Lignin in straw and its applications as an adhesive. *International Journal of Adhesion and Adhesives*, 48, 92–101.
<http://doi.org/10.1016/j.ijadhadh.2013.09.001>
- Ghose, T. K. (1987). International Union of Pure Commission on Biotechnology * Measurement of. *Pure and Applied Chemistry*, 59(2), 257–268.
<http://doi.org/10.1351/pac198759020257>
- Goh, C. S., Lee, K. T., & Bhatia, S. (2010). Hot compressed water pretreatment of oil palm fronds to enhance glucose recovery for production of second generation bio-ethanol. *Bioresource Technology*, 101(19), 7362–7367. Retrieved from <http://www.ncbi.nlm.nih.gov/pubmed/20471249>
- Goh, C. S., Tan, H. T., & Lee, K. T. (2012). Pretreatment of oil palm frond using hot compressed water: an evaluation of compositional changes and pulp digestibility using

severity factors. *Bioresource Technology*, 110, 662–9. Retrieved from <http://www.sciencedirect.com/science/article/pii/S0960852412001071>

Goh, C. S., Tan, K. T., Lee, K. T., & Bhatia, S. (2010). Bio-ethanol from lignocellulose: Status, perspectives and challenges in Malaysia. *Bioresource Technology*, 101(13), 4834–4841. <http://doi.org/10.1016/j.biortech.2009.08.080>

Gregg, D. J., & Saddler, J. N. (1996). Factors Affecting Cellulose Hydrolysis and the Potential of Enzyme Recycle to Enhance the Efficiency of an Integrated Wood to Ethanol Process. *Biotechnology and Bioengineering*, 51, 375–383. [http://doi.org/10.1002/\(SICI\)1097-0290\(19960820\)51](http://doi.org/10.1002/(SICI)1097-0290(19960820)51)

Gunjikar, T. P., Sawant, S. B., & Joshi, J. B. (2001). Glucosidase in a Mechanically Agitated Reactor. *Biotechnol. Prog.*, 17, 1166–1168.

Guo, H., Su, R., Huang, R., Qi, W., & He, Z. (2015). Co-optimization of sugar yield and input energy by the stepwise reduction of agitation rate during lignocellulose hydrolysis. *Food and Bioproducts Processing*, 95, 1–6. <http://doi.org/10.1016/j.fbp.2015.03.005>

Gupta, A., & Verma, J. P. (2015). Sustainable bio-ethanol production from agro-residues : A review. *Renewable and Sustainable Energy*, 41, 550–567.

Haghghi Mood, S., Hossein Golfeshan, A., Tabatabaei, M., Salehi Jouzani, G., Najafi, G. H., Gholami, M., & Ardjmand, M. (2013b). Lignocellulosic biomass to bioethanol, a comprehensive review with a focus on pretreatment. *Renewable and Sustainable Energy Reviews*, 27, 77–93. <http://doi.org/10.1016/j.rser.2013.06.033>

Hames, B., Ruiz, R., Scarlata, C., Sluiter, A., Sluiter, J., & Templeton, D. (2008). Preparation of Samples for Compositional Analysis. *National Renewable Energy Laboratory*, (August), 1–9.

Han, J. S., & Rowell, J. S. (1997). Chemical Composition of Fibers. In R. M. Rowell, R. A. Young, & J. K. Rowell (Eds.), *Paper and composites from agro-based resources* (pp. 83–134). New York: CRC Lewis Publishers.

Harmsen, P., Huijgen, W., López, L., & Bakker, R. (2010). Literature Review of Physical and Chemical Pretreatment Processes for Lignocellulosic Biomass. *BioSynergy*. Retrieved from <http://www.ecn.nl/docs/library/report/2010/e10013.pdf>

Harun, S., Balan, V., Takriff, M. S., Hassan, O., Jahim, J., & Dale, B. E. (2013). Performance of AFEX™ pretreated rice straw as source of fermentable sugars: the influence of particle size. *Biotechnology for Biofuels*, 6(1), 40. <http://doi.org/10.1186/1754-6834-6-40>

Harun, S., Jahim, J. M., Takriff, M. S., & Hassan, O. (2015). Chemical Composition of Native and Ammonia Fiber Expansion. *Chemical Composition of Native and Ammonia Fiber*

- Hashim, R., Saari, N., Sulaiman, O., Sugimoto, T., Hiziroglu, S., Sato, M., & Tanaka, R. (2010). Effect of particle geometry on the properties of binderless particleboard manufactured from oil palm trunk. *Materials and Design*, 31(9), 4251–4257. <http://doi.org/10.1016/j.matdes.2010.04.012>
- Hassan, O., Ling, T. P., Maskat, M. Y., Illias, R. M., Badri, K., Jahim, J., & Mahadi, N. M. (2013). Optimization of pretreatments for the hydrolysis of oil palm empty fruit bunch fiber (EFBF) using enzyme mixtures. *Biomass and Bioenergy*, 56, 137–146. Retrieved from <http://www.sciencedirect.com/science/article/pii/S0961953413002134>
- Hayes, D. J., Fitzpatrick, S., Hayes, M. H. B., & Ross, J. R. H. (2008). The Biofine Process - Production of Levulinic Acid, Furfural, and Formic Acid from Lignocellulosic Feedstocks. In *Biorefineries-Industrial Processes and Products: Status Quo and Future Directions* (Vol. 1, pp. 139–164). <http://doi.org/10.1002/9783527619849.ch7>
- Hendriks, A. T. W. M., & Zeeman, G. (2009). Pretreatments to enhance the digestibility of lignocellulosic biomass. *Bioresource Technology*, 100(1), 10–8. <http://doi.org/10.1016/j.biortech.2008.05.027>
- Hodge, D. B., Karim, M. N., Schell, D. J., & McMillan, J. D. (2009). Model-based fed-batch for high-solids enzymatic cellulose hydrolysis. *Applied Biochemistry and Biotechnology*, 152(1), 88–107. <http://doi.org/10.1007/s12010-008-8217-0>
- Howard, R. L., Abotsi, E., Jansen, van R. E. L., & Howard, S. (2003). Lignocellulose biotechnology: issues of bioconversion and enzyme production. *African Journal of Biotechnology*, 2(12), 602–619. <http://doi.org/10.5897/AJB2003.000-1115>
- Hu, J., Chandra, R., Arantes, V., Gourlay, K., van Dyk, J. S., & Saddler, J. N. (2015). The addition of accessory enzymes enhances the hydrolytic performance of cellulase enzymes at high solid loadings. *Bioresource Technology*, 186, 149–53. <http://doi.org/10.1016/j.biortech.2015.03.055>
- Hussin, M. H., Rahim, A. A., Mohamad Ibrahim, M. N., & Brosse, N. (2013). Physicochemical characterization of alkaline and ethanol organosolv lignins from oil palm (*Elaeis guineensis*) fronds as phenol substitutes for green material applications. *Industrial Crops and Products*, 49, 23–32. <http://doi.org/10.1016/j.indcrop.2013.04.030>
- Iberahim, N. I., Jahim, J. M., Harun, S., Nor, M. T. M., & Hassan, O. (2013). Sodium Hydroxide Pretreatment and Enzymatic Hydrolysis of Oil Palm Mesocarp Fiber. *International Journal of Chemical Engineering and Applications*, 4(3), 101–105. <http://doi.org/10.7763/IJCEA.2013.V4.272>
- Indera Luthfi, A. A., Jahim, J. M., Harun, S., Tan, J. P., & Mohammad, A. W. (2016). Biorefinery approach towards greener succinic acid production from oil palm frond

- bagasse. *Process Biochemistry*. <http://doi.org/10.1016/j.procbio.2016.08.011>
- Ingesson, H., Zacchi, G., Yang, B., Esteghlalian, A. R., & Saddler, J. N. (2001). The effect of shaking regime on the rate and extent of enzymatic hydrolysis of cellulose. *Journal of Biotechnology*, 88(2), 177–182. [http://doi.org/10.1016/S0168-1656\(01\)00273-5](http://doi.org/10.1016/S0168-1656(01)00273-5)
- Isikgor, F. H., & C. Remzi Becer. (2015). Lignocellulosic Biomass: a sustainable platform for production of bio-based chemicals and polymers. *Polymer Chemistry*, 6, 4497–4559. <http://doi.org/10.1039/c3py00085k>
- Ismail, N. (2017, March 10). Global palm oil output forecast to increase by 6 million tonnes. *The Star*, p. 1.
- Jorgensen, H., Kristensen, J. B., & Felby, C. (2007). Perspective: Jatropha cultivation in southern India: Assessing farmers' experiences. *Biofuels, Bioproducts and Biorefining*, 6(3), 246–256. <http://doi.org/10.1002/bbb>
- Juturu, V., & Wu, J. C. (2012). Microbial xylanases: Engineering, production and industrial applications. *Biotechnology Advances*, 30(6), 1219–1227. <http://doi.org/10.1016/j.biotechadv.2011.11.006>
- Juturu, V., & Wu, J. C. (2014). Microbial cellulases: Engineering, production and applications. *Renewable and Sustainable Energy Reviews*, 33, 188–203. <http://doi.org/10.1016/j.rser.2014.01.077>
- Karazhiyan, H., Razavi, S. M. A., & Phillips, G. O. (2011). Extraction optimization of a hydrocolloid extract from cress seed (*Lepidium sativum*) using response surface methodology. *Food Hydrocolloids*, 25(5), 915–920. <http://doi.org/10.1016/j.foodhyd.2010.08.022>
- Karthika, K., Arun, A. B., & Rekha, P. D. (2012). Enzymatic hydrolysis and characterization of lignocellulosic biomass exposed to electron beam irradiation. *Carbohydrate Polymers*, 90(2), 1038–1045. <http://doi.org/10.1016/j.carbpol.2012.06.040>
- Kaur, P. P., Arneja, J. S., & Singh, J. (1998). Enzymic hydrolysis of rice straw by crude cellulase from *Trichoderma reesei*. *Bioresource Technology*, 66(3), 267–269. [http://doi.org/10.1016/S0960-8524\(97\)00138-7](http://doi.org/10.1016/S0960-8524(97)00138-7)
- Khalil, H. P. S. A., Jawaid, M., Hassan, A., Paridah, M. T., & Zaidon, A. (2012). Oil Palm Biomass Fibres and Recent Advancement in Oil Palm Biomass Fibres Based Hybrid Biocomposites. *Composites and Their Applications*, 187–220. <http://doi.org/10.5772/48235>
- Kim, I., & Han, J.-I. (2012). Optimization of alkaline pretreatment conditions for enhancing glucose yield of rice straw by response surface methodology. *Biomass and Bioenergy*, 46,

210–217. <http://doi.org/10.1016/j.biombioe.2012.08.024>

Kim, S. M., Dien, B. S., Tumbleson, M. E., Rausch, K. D., & Singh, V. (2016). Improvement of sugar yields from corn stover using sequential hot water pretreatment and disk milling. *Bioresource Technology*, 216, 706–713. <http://doi.org/10.1016/j.biortech.2016.06.003>

Kim, Y., Hendrickson, R., Mosier, N. S., Ladisch, M. R., Bals, B., Balan, V., & Dale, B. E. (2008). Enzyme hydrolysis and ethanol fermentation of liquid hot water and AFEX pretreated distillers' grains at high-solids loadings. *Bioresource Technology*, 99(12), 5206–5215. <http://doi.org/10.1016/j.biortech.2007.09.031>

Kootstra, A. M. J., Beeftink, H. H., Scott, E. L., & Sanders, J. P. M. (2009). Comparison of dilute mineral and organic acid pretreatment for enzymatic hydrolysis of wheat straw. *Biochemical Engineering Journal*, 46(2), 126–131.
<http://doi.org/10.1016/j.bej.2009.04.020>

Kristensen, J. B., Felby, C., & Jørgensen, H. (2009). Yield-determining factors in high-solids enzymatic hydrolysis of lignocellulose. *Biotechnology for Biofuels*, 2(1), 11.
<http://doi.org/10.1186/1754-6834-2-11>

Kuhad, R. C., Gupta, R., & Singh, A. (2011). Microbial Cellulases and their industrial Applications. *Enzyme Research*, 1–10.

Kumar, P., Barrett, D. M., Delwiche, M. J., & Stroeve, P. (2009). Methods for Pretreatment of Lignocellulosic Biomass for Efficient Hydrolysis and Biofuel Production. *Industrial and Engineering Chemistry (Analytical Edition)*, 48(8), 3713–3729.
<http://doi.org/10.1021/ie801542g>

Laureano-Perez, L., Teymour, F., Alizadeh, H., & Dale, B. E. (2005). Understanding factors that limit enzymatic hydrolysis of biomass. *Applied Biochemistry and Biotechnology*, 124, 1081–1099. http://doi.org/10.1007/978-1-59259-991-2_91

Lee, J. M., Jameel, H., & Venditti, R. A. (2010). A comparison of the autohydrolysis and ammonia fiber explosion (AFEX) pretreatments on the subsequent enzymatic hydrolysis of coastal Bermuda grass. *Bioresource Technology*, 101(14), 5449–5458.
<http://doi.org/10.1016/j.biortech.2010.02.055>

Lee, H. V., Hamid, S. B. A., & Zain, S. K. (2014). Conversion of lignocellulosic biomass to nanocellulose: Structure and chemical process. *Scientific World Journal*, 2014.
<http://doi.org/10.1155/2014/631013>

Lenihan, P., Orozco, A., Neill, E. O., Ahmad, M. N. M., Rooney, D. W., & Walker, G. M. (2010). Dilute acid hydrolysis of lignocellulosic biomass. *Chemical Engineering Journal*, 156, 395–403. <http://doi.org/10.1016/j.cej.2009.10.061>

- Li, C., & Champagne, P. (2009). Fixed-bed column study for the removal of cadmium (II) and nickel (II) ions from aqueous solutions using peat and mollusk shells. *Journal of Hazardous Materials*, 171(1), 872–878. <http://doi.org/10.1016/j.jhazmat.2009.06.084>
- Li, H., Wang, X., Liu, C., Ren, J., Zhao, X., Sun, R., & Wu, A. (2016). An efficient pretreatment for the selectively hydrothermal conversion of corncob into furfural: The combined mixed ball milling and ultrasonic pretreatments. *Industrial Crops and Products*, 94, 721–728. <http://doi.org/10.1016/j.indcrop.2016.09.052>
- Li, X., Ouyang, J., Xu, Y., Chen, M., Song, X., Yong, Q., & Yu, S. (2009). Optimization of culture conditions for production of yeast biomass using bamboo wastewater by response surface methodology. *Bioresource Technology*, 100(14), 3613–3617. <http://doi.org/10.1016/j.biortech.2009.03.001>
- Lim, J. S., Abdul Manan, Z., Wan Alwi, S. R., & Hashim, H. (2012). A review on utilisation of biomass from rice industry as a source of renewable energy. *Renewable and Sustainable Energy Reviews*. <http://doi.org/10.1016/j.rser.2012.02.051>
- Lim, K. O., Zainal, Z. A., Quadir, G. A., & Abdullah, M. Z. (2000). Plant based energy potential and biomass utilisation in malaysian LIM.pdf. *International Energy Journal*, 1(September 2016), 77–88.
- Lin, L., Yan, R., Liu, Y., & Jiang, W. (2010). In-depth investigation of enzymatic hydrolysis of biomass wastes based on three major components: Cellulose, hemicellulose and lignin. *Bioresource Technology*, 101(21), 8217–8223. <http://doi.org/10.1016/j.biortech.2010.05.084>
- Lin, Q., Li, H., Ren, J., Deng, A., Li, W., Liu, C., & Sun, R. (2017). Production of xylooligosaccharides by microwave-induced, organic acid-catalyzed hydrolysis of different xylan-type hemicelluloses: Optimization by response surface methodology. *Carbohydrate Polymers*, 157, 214–225. <http://doi.org/10.1016/j.carbpol.2016.09.091>
- Ma, H., & Wu, Y.-J. (2009). Enhanced enzymatic saccharification of rice straw by microwave pretreatment. *Bioresource Technology*, 100, 1279–1284.
- Mahat, N. (2010). Comparison Study on Oil Palm Trunk and Oil Palm Fruit Bunch Fibre Reinforced Laterite Bricks. *Journal of Modern Applied Science*, 4(7), 119–129.
- Malherbe, S., & Cloete, T. E. (2002). Lignocellulose biodegradation: Fundamentals and applications. *Reviews in Environmental Science and Biotechnology*, 1(2), 105–114. <http://doi.org/10.1023/A:1020858910646>
- McIntosh, S., & Vancov, T. (2010). Enhanced enzyme saccharification of Sorghum bicolor straw using dilute alkali pretreatment. *Bioresource Technology*, 101(17), 6718–6727. <http://doi.org/10.1016/j.biortech.2010.03.116>

- McKendry, P. (2002). Energy production from biomass (part 1): Overview of biomass. *Bioresource Technology*. [http://doi.org/10.1016/S0960-8524\(01\)00118-3](http://doi.org/10.1016/S0960-8524(01)00118-3)
- Meng, X., & Ragauskas, A. J. (2014). Recent advances in understanding the role of cellulose accessibility in enzymatic hydrolysis of lignocellulosic substrates. *Current Opinion in Biotechnology*, 27, 150–8. <http://doi.org/10.1016/j.copbio.2014.01.014>
- Michelin, M., & Teixeira, J. A. (2016). Liquid hot water pretreatment of multi feedstocks and enzymatic hydrolysis of solids obtained thereof. *Bioresource Technology*, 216, 862–869. <http://doi.org/10.1016/j.biortech.2016.06.018>
- Miettinen-Oinonen, A., Londesborough, J., Joutsjoki, V., Lantto, R., & Vehmaanpera, J. (2004). Three cellulases from Melanocarpus albomyces for textile treatment at neutral pH. *Enzyme and Microbial Technology*, 34(3–4), 332–341. <http://doi.org/10.1016/j.enzmictec.2003.11.011>
- Modenbach, A. A., & Nokes, S. E. (2013). Enzymatic hydrolysis of biomass at high-solids loadings – A review. *Biomass and Bioenergy*, 56, 526–544. <http://doi.org/10.1016/j.biombioe.2013.05.031>
- Mohagheghi, A., Evans, K., Chou, Y.-C., & Zhang, M. (2002). Cofermentation of glucose, xylose, and arabinose by genomic DNA-integrated xylose/arabinose fermenting strain of Zymomonas mobilis AX101. *Applied Biochemistry and Biotechnology*, 98–100, 885–898. <http://doi.org/10.1385/ABAB:98-100:1-9:885>
- Mohamad Ibrahim, M. N., Zakaria, N., Sipaut, C. S., Sulaiman, O., & Hashim, R. (2011). Chemical and thermal properties of lignins from oil palm biomass as a substitute for phenol in a phenol formaldehyde resin production. *Carbohydrate Polymers*, 86(1), 112–119. <http://doi.org/10.1016/j.carbpol.2011.04.018>
- Mosier, N., Wyman, C., Dale, B., Elander, R., Lee, Y. Y., Holtapple, M., & Ladisch, M. (2005a). Features of promising technologies for pretreatment of lignocellulosic biomass. *Bioresource Technology*, 96(6), 673–686. <http://doi.org/10.1016/j.biortech.2004.06.025>
- Motta, F., Andrade, C., & Santana, M. (2013). A Review of Xylanase Production by the Fermentation of Xylan: Classification, Characterization and Applications. In *Sustainable Degradation of Lignocellulosic Biomass - Techniques, Applications and Commercialization* (Vol. Chapter 10, pp. 251–275). InTech. <http://doi.org/10.5772/53544>
- Muktham, R., Ball, A. S., Bhargava, S. K., & Bankupalli, S. (2016). Bioethanol production from non-edible de-oiled Pongamia pinnata seed residue-optimization of acid hydrolysis followed by fermentation. *Industrial Crops and Products*, 94, 490–497. <http://doi.org/10.1016/j.indcrop.2016.09.019>
- Mussatto, S. I., Dragone, G., Fernandes, M., Milagres, A. M. F., & Roberto, I. C. (2008). The

- effect of agitation speed, enzyme loading and substrate concentration on enzymatic hydrolysis of cellulose from brewer's spent grain. *Cellulose*, 15(5), 711–721.
<http://doi.org/10.1007/s10570-008-9215-7>
- Nath, A., & Chattopadhyay, P. K. (2007). Optimization of oven toasting for improving crispness and other quality attributes of ready to eat potato-soy snack using response surface methodology. *Journal of Food Engineering*, 80(4), 1282–1292.
<http://doi.org/10.1016/j.jfoodeng.2006.09.023>
- Ngadi, N., & Rusli, N. S. (2014). Ultrasound-Assisted Extraction of Lignin from Oil Palm Frond. *Jurnal Teknologi*, 67(4), 67–70.
- Nieves, D. C., Ruiz, H. A., de Cárdenas, L. Z., Alvarez, G. M., Aguilar, C. N., Ilyina, A., & Martínez Hernández, J. L. (2016a). Enzymatic hydrolysis of chemically pretreated mango stem bark residues at high solid loading. *Industrial Crops and Products*, 83, 500–508.
<http://doi.org/10.1016/j.indcrop.2015.12.079>
- Nlewem, K. C., & Thrash, M. E. (2010). Comparison of different pretreatment methods based on residual lignin effect on the enzymatic hydrolysis of switchgrass. *Bioresource Technology*, 101(14), 5426–30. <http://doi.org/10.1016/j.biortech.2010.02.031>
- Noparat, P., Prasertsan, P., O-Thong, S., & Pan, X. (2015). Dilute Acid Pretreatment of Oil Palm Trunk Biomass at High Temperature for Enzymatic Hydrolysis. *Energy Procedia*, 79, 924–929. <http://doi.org/10.1016/j.egypro.2015.11.588>
- Norgren, M., & Edlund, H. (2014). Current Opinion in Colloid & Interface Science Lignin : Recent advances and emerging applications. *Current Opinion in Colloid & Interface Science*, 19(5), 409–416. <http://doi.org/10.1016/j.cocis.2014.08.004>
- Nurfahmi, Ong, H. C., Jan, B. M., Tong, C. W., Fauzi, H., & Chen, W. H. (2016). Effects of organosolv pretreatment and acid hydrolysis on palm empty fruit bunch (PEFB) as bioethanol feedstock. *Biomass and Bioenergy*, 95, 78–83.
<http://doi.org/10.1016/j.biombioe.2016.09.008>
- Ofori-Boateng, C., & Lee, K. T. (2013). Comparative thermodynamic sustainability assessment of lignocellulosic pretreatment methods for bioethanol production via exergy analysis. *Chemical Engineering Journal*, 228, 162–171. Retrieved from <http://www.sciencedirect.com/science/article/pii/S1385894713005780>
- Ofori-Boateng, C., & Lee, K. T. (2014). An oil palm-based biorefinery concept for cellulosic ethanol and phytochemicals production: Sustainability evaluation using exergetic life cycle assessment. *Applied Thermal Engineering*, 62(1), 90–104. Retrieved from <http://www.sciencedirect.com/science/article/pii/S135943111300656X>
- Ofori-Boateng, C., Lee, K. T., & Saad, B. (2014). A biorefinery concept for simultaneous recovery of cellulosic ethanol and phenolic compounds from oil palm fronds: Process

- optimization. *Energy Conversion and Management*, 81, 192–200. Retrieved from <http://www.sciencedirect.com/science/article/pii/S0196890414001496>
- Omar, R., Idris, a., Yunus, R., Khalid, K., & Aida Isma, M. I. (2011). Characterization of empty fruit bunch for microwave-assisted pyrolysis. *Fuel*, 90(4), 1536–1544. Retrieved from <http://www.sciencedirect.com/science/article/pii/S0016236111000251>
- Orozco, A., Ahmad, M., Rooney, D., & Walker, G. Á. (2000). Dilute acid hydrolysis of cellulose and cellulosic bio-waste using a microwave reactor system, 85, 446–449. <http://doi.org/10.1205/psep07003>
- Pallapolu, V. R., Lee, Y. Y., Garlock, R. J., Balan, V., Dale, B. E., Kim, Y., Warner, R. E. (2011). Effects of enzyme loading and β -glucosidase supplementation on enzymatic hydrolysis of switchgrass processed by leading pretreatment technologies. *Bioresource Technology*, 102(24), 11115–20. <http://doi.org/10.1016/j.biortech.2011.03.085>
- Patel, H., Chapla, D., & Shah, A. (2017). Bioconversion of pretreated sugarcane bagasse using enzymatic and acid followed by enzymatic hydrolysis approaches for bioethanol production. *Renewable Energy*, 109, 323–331. <http://doi.org/10.1016/j.renene.2017.03.057>
- Pengilly, C., García-Aparicio, M. P., Diedericks, D., Brienz, M., & Görgens, J. F. (2015). Enzymatic hydrolysis of steam-pretreated sweet sorghum bagasse by combinations of cellulase and endo-xylanase. *Fuel*, 154, 352–360. <http://doi.org/10.1016/j.fuel.2015.03.072>
- Percival Zhang, Y. H., Himmel, M. E., & Mielenz, J. R. (2006). Outlook for cellulase improvement: Screening and selection strategies. *Biotechnology Advances*, 24(5), 452–481. <http://doi.org/10.1016/j.biotechadv.2006.03.003>
- Pereira, S. C., Maehara, L., Machado, C. M. M., & Farinas, C. S. (2016). Physical–chemical–morphological characterization of the whole sugarcane lignocellulosic biomass used for 2G ethanol production by spectroscopy and microscopy techniques. *Renewable Energy*, 87, 607–617. <http://doi.org/10.1016/j.renene.2015.10.054>
- Pérez, J., Muñoz-Dorado, J., De La Rubia, T., & Martínez, J. (2002). Biodegradation and biological treatments of cellulose, hemicellulose and lignin: An overview. *International Microbiology*, 5(2), 53–63. <http://doi.org/10.1007/s10123-002-0062-3>
- Piarpuzán, D., Quintero, J. A., & Cardona, C. A. (2011). Empty fruit bunches from oil palm as a potential raw material for fuel ethanol production. *Biomass and Bioenergy*, 35(3), 1130–1137. <http://doi.org/10.1016/j.biombioe.2010.11.038>
- Pothiraj, C., Kanmani, P., & Balaji, P. (2006). Potential bioproducts and their applications : Biomass Production of extracellular enzymes by fungi : Extensive. *Mycobiology*, 34(4), 159–165.

- Prasad, K. S. H., Rao, C. S., & Rao, D. N. (2012). Application of Design of Experiments to Plasma Arc Welding Process : A Review. *Journal of the Brazilian Society of Mechanical Sciences and Engineering*, 34(1), 75–81. <http://doi.org/10.1590/S1678-58782012000100010>
- Punsuvon, V. (2013). Optimization of Delignification and Enzyme Hydrolysis of Steam Exploded Oil Palm Trunk for Ethanol Production by Response Surface Methodology. *InTech*, 157–178.
- Qiao, D., Hu, B., Gan, D., Sun, Y., Ye, H., & Zeng, X. (2009). Extraction optimized by using response surface methodology, purification and preliminary characterization of polysaccharides from *Hyriopsis cumingii*. *Carbohydrate Polymers*, 76(3), 422–429. <http://doi.org/10.1016/j.carbpol.2008.11.004>
- Ramachandriya, K. D., Wilkins, M., Atiyeh, H. K., Dunford, N. T., & Hiziroglu, S. (2013). Effect of high dry solids loading on enzymatic hydrolysis of acid bisulfite pretreated Eastern redcedar. *Bioresource Technology*, 147, 168–176. <http://doi.org/10.1016/j.biortech.2013.08.048>
- Ramos, L. P., da Silva, L., Ballem, A. C., Pitarelo, A. P., Chiarello, L. M., & Silveira, M. H. L. (2015). Enzymatic hydrolysis of steam-exploded sugarcane bagasse using high total solids and low enzyme loadings. *Bioresource Technology*, 175, 195–202. <http://doi.org/10.1016/j.biortech.2014.10.087>
- Rana, D., Rana, V., & Ahring, B. K. (2012). Producing high sugar concentrations from loblolly pine using wet explosion pretreatment. *Bioresource Technology*, 121, 61–7. Retrieved from <http://www.ncbi.nlm.nih.gov/pubmed/22854131>
- Rasat M. S. M., Razak R., Sulaiman O., Moktar J., Mohamed A., Tabet T. A., and K. I. (2011). Properties of Composite Boards From Oil Palm. *Bioresources*, 6(November), 4389–4403.
- Rasheed, T., Mathew, A., Mathew, M., Sukumaran, R. K., & Elyas, K. . (2015). Optimization of process parameters for b-glucosidase production by *Byssochlamys fulva* in slurry state fermentation. *Biotechnological Research Journal*, 1(1), 101–108.
- Ratnasingam, J., Tek, T. C., Farrokhpayam, S. R., & Ioras, F. (2009). Tool wear characteristics of oil palm empty fruit bunch (OPEFB) particleboard. *Journal of the Institute of Wood Science*, 19(2), 91–94. <http://doi.org/10.1179/002032009X12536100262277>
- Rodrigues, I. M., Carvalho, M. G. V. S., & Rocha, J. M. S. (2014). Increasing the protein content of rapeseed meal by enzymatic hydrolysis of carbohydrates. *BioResources*, 9(2), 2010–2025.
- Rodríguez-Chong, A., Ramírez, J. A., Garrote, G., & Vázquez, M. (2004). Hydrolysis of sugar cane bagasse using nitric acid: A kinetic assessment. *Journal of Food Engineering*, 61(2), 143–152. [http://doi.org/10.1016/S0260-8774\(03\)00080-3](http://doi.org/10.1016/S0260-8774(03)00080-3)

Romero, I., Ruiz, E., Castro, E., & Moya, M. (2009). Chemical Engineering Research and Design Acid hydrolysis of olive tree biomass, 8(September), 633–640.
<http://doi.org/10.1016/j.cherd.2009.10.007>

Rovio, S., Simolin, H., Koljonen, K., & Sirén, H. (2008). Determination of monosaccharide composition in plant fiber materials by capillary zone electrophoresis. *Journal of Chromatography A*, 1185(1), 139–144. <http://doi.org/10.1016/j.chroma.2008.01.031>

Rozman, H. D., Lai, C. Y., Ismail, H., & Ishak, Z. A. M. (2000). The effect of coupling agents on the mechanical and physical properties of oil palm empty fruit bunch-polypropylene composites. *Polymer International*, 49(11), 1273–1278. [http://doi.org/10.1002/1097-0126\(200011\)49:11<1273::AID-PI469>3.0.CO;2-U](http://doi.org/10.1002/1097-0126(200011)49:11<1273::AID-PI469>3.0.CO;2-U)

Rozman, H. D., Tay, G. S., Kumar, R. N., Abusamah, a, Ismail, H., & Ishak, Z. a M. (2001). Polypropylene oil palm empty fruit bunch glass bre hybrid composites : a preliminary study on the flexural and tensile properties. *European Polymer Journal*, 37, 1283–1291. [http://doi.org/10.1016/S0014-3057\(00\)00243-3](http://doi.org/10.1016/S0014-3057(00)00243-3)

Sabiha-Hanim, S., Noor, M. A. M., & Rosma, A. (2011). Effect of autohydrolysis and enzymatic treatment on oil palm (*Elaeis guineensis Jacq.*) frond fibres for xylose and xylooligosaccharides production. *Bioresource Technology*, 102(2), 1234–9. Retrieved from <http://www.ncbi.nlm.nih.gov/pubmed/20797853>

Saha, B. C. (2003). Hemicellulose bioconversion. *Journal of Industrial Microbiology and Biotechnology*, 30(5), 279–291. <http://doi.org/10.1007/s10295-003-0049-x>

Saini, R., Saini, J. K., Adsul, M., Patel, A. K., Mathur, A., Tuli, D., & Singhania, R. R. (2015). Enhanced cellulase production by *Penicillium oxalicum* for bio-ethanol application. *Bioresource Technology*, 188, 240–246. <http://doi.org/10.1016/j.biortech.2015.01.048>

Sakdaronnarong, C., Srimarut, N., Lucknakhul, N., Na-songkla, N., & Jonglertjunya, W. (2014). Two-step acid and alkaline ethanolysis/alkaline peroxide fractionation of sugarcane bagasse and rice straw for production of polylactic acid precursor. *Biochemical Engineering Journal*, 85, 49–62. Retrieved from <http://www.sciencedirect.com/science/article/pii/S1369703X14000254>

Sant'Ana da Silva, A., Fernandes de Souza, M., Ballesteros, I., Manzanares, P., Ballesteros, M., & P. S. Bon, E. (2016). High-solids content enzymatic hydrolysis of hydrothermally pretreated sugarcane bagasse using a laboratory-made enzyme blend and commercial preparations. *Process Biochemistry* (Vol. 51).

Segal, L., Creely, J. J., Martin, a. E., & Conrad, C. M. (1959). An Empirical Method for Estimating the Degree of Crystallinity of Native Cellulose Using the X-Ray Diffractometer. *Textile Research Journal*, 29(10), 786–794. <http://doi.org/10.1177/004051755902901003>

Selig, M., Weiss, N., & Ji, Y. (2008). Enzymatic Saccharification of Lignocellulosic Biomass. *National Renewable Energy Laboratory*.

Series, W. (2013). Aqueous Pretreatment of Plant Biomass for Biological and Chemical Conversion to Fuels and Chemicals. (C. E. Wyman, Ed.) (1st ed.). John Wiley & Sons, Ltd.

Sindhu, R., & Pandey, A. (2016). Biological pretreatment of lignocellulosic biomass – An overview. *Bioresource Technology*, 199, 76–82.
<http://doi.org/10.1016/j.biortech.2015.08.030>

Siti Aisyah, M. S., Uemura, Y., & Yusup, S. (2014). The Effect of Alkaline Addition in Hydrothermal Pretreatment of Empty Fruit Bunches on Enzymatic Hydrolysis Efficiencies. *Procedia Chemistry*, 9, 151–157. <http://doi.org/10.1016/j.proche.2014.05.018>

Sluiter, A., Ruiz, R., Scarlata, C., Sluiter, J., And Templeton, D. (2008). Determination of Extractives in Biomass. *Technical Report NREL/TP-510-42619*, (July 2005), 1–9. Retrieved from <http://www.nrel.gov/biomass/pdfs/42619.pdf>

Sluiter, A., Hames, B., Ruiz, R., Scarlata, C., Sluiter, J., & Templeton, D. (2008). Determination of ash in biomass. *Nrel/Tp-510-42622*, (April 2005), 18. <http://doi.org/NREL/TP-510-42619>

Sluiter, A., Hames, B., Ruiz, R., Scarlata, C., Sluiter, J., Templeton, D., & Crocker, D. (2011). Determination of Structural Carbohydrates and Lignin in Biomass. *National Renewable Energy Laboratory*.

Subhedar, P. B., & Gogate, P. R. (2014). Alkaline and ultrasound assisted alkaline pretreatment for intensification of delignification process from sustainable raw-material. *Ultrasonics Sonochemistry*, 21(1), 216–225. <http://doi.org/10.1016/j.ultsonch.2013.08.001>

Sudiyani, Y., Styarini, D., Tri wahyuni, E., Sudiyarmanto, Sembiring, K. C., Aristiawan, Y., Han, M. H. (2013). Utilization of biomass waste empty fruit bunch fiber of palm oil for bioethanol production using pilot - Scale unit. *Energy Procedia*, 32, 31–38. <http://doi.org/10.1016/j.egypro.2013.05.005>

Sukri, S., & Rahman, R. (2014). Optimization of Alkaline Pretreatment Conditions of Oil Palm Fronds in Improving the Lignocelluloses Contents for Reducing Sugar Production. *Romanian Biotechnological Letters*, 19(1), 9006–9018. Retrieved from http://www.rombio.eu/vol19nr1/11_lucr_16_Sabrina_rec_15_.082013_ac_04.12.pdf

Sun, F., & Chen, H. (2008). Organosolv pretreatment by crude glycerol from oleochemicals industry for enzymatic hydrolysis of wheat straw. *Bioresource Technology*, 99(13), 5474–5479. <http://doi.org/10.1016/j.biortech.2007.11.001>

- Sun, F. F., Hong, J., Hu, J., Saddler, J. N., Fang, X., Zhang, Z., & Shen, S. (2015). Accessory enzymes influence cellulase hydrolysis of the model substrate and the realistic lignocellulosic biomass. *Enzyme and Microbial Technology*, 79–80, 42–48. <http://doi.org/10.1016/j.enzmictec.2015.06.020>
- Sun, S., Cao, X., Sun, S., Xu, F., Song, X., Sun, R.-C., & Jones, G. L. (2014). Improving the enzymatic hydrolysis of thermo-mechanical fiber from *Eucalyptus urophylla* by a combination of hydrothermal pretreatment and alkali fractionation. *Biotechnology for Biofuels*, 7(1), 116. <http://doi.org/10.1186/s13068-014-0116-8>
- Sun, S., Sun, S., Cao, X., & Sun, R. (2016). The role of pretreatment in improving the enzymatic hydrolysis of lignocellulosic materials. *Bioresource Technology*, 199, 49–58. <http://doi.org/10.1016/j.biortech.2015.08.061>
- Sun, Y., & Cheng, J. (2002). Hydrolysis of lignocellulosic materials for ethanol production: a review. *Bioresource Technology*, 83(1), 1–11. [http://doi.org/10.1016/S0960-8524\(01\)00212-7](http://doi.org/10.1016/S0960-8524(01)00212-7)
- Talukder, M. M. R., Goh, H. Y., & Puah, S. M. (2017a). Interaction of silica with cellulase and minimization of its inhibitory effect on cellulose hydrolysis. *Biochemical Engineering Journal*, 118, 91–96. <http://doi.org/10.1016/j.bej.2016.11.016>
- Tan, H. T., & Lee, K. T. (2012). Understanding the impact of ionic liquid pretreatment on biomass and enzymatic hydrolysis. *Chemical Engineering Journal*, 183, 448–458. Retrieved from <http://www.sciencedirect.com/science/article/pii/S1385894711016378>
- Tan, H. T., Lee, K. T., & Mohamed, A. R. (2011). Pretreatment of lignocellulosic palm biomass using a solvent-ionic liquid [BMIM]Cl for glucose recovery: An optimisation study using response surface methodology. *Carbohydrate Polymers*, 83(4), 1862–1868. Retrieved from <http://www.sciencedirect.com/science/article/pii/S0144861710008659>
- Tan, I. S., & Lee, K. T. (2014). Enzymatic hydrolysis and fermentation of seaweed solid wastes for bioethanol production: An optimization study. *Energy*, 78, 53–62. <http://doi.org/10.1016/j.energy.2014.04.080>
- Tan, J. P., Jahim, J. M., Harun, S., Wu, T. Y., & Mumtaz, T. (2016). Utilization of oil palm fronds as a sustainable carbon source in biorefineries. *International Journal of Hydrogen Energy*, 41(8), 4896–4906. <http://doi.org/10.1016/j.ijhydene.2015.08.034>
- Taneda, D., Ueno, Y., Ikeo, M., & Okino, S. (2012). Characteristics of enzyme hydrolysis of cellulose under static condition. *Bioresource Technology*, 121, 154–160. <http://doi.org/10.1016/j.biortech.2012.06.104>
- Tippkötter, N., Duwe, A.-M., Wiesen, S., Sieker, T., & Ulber, R. (2014). Enzymatic hydrolysis of beech wood lignocellulose at high solid contents and its utilization as substrate for the production of biobutanol and dicarboxylic acids. *Bioresource Technology*, 167, 447–55.

<http://doi.org/10.1016/j.biortech.2014.06.052>

Tong, Z., Cheng, N., & Pratap, P. (2013). Pretreatment of Lignocellulosic Biomass for Biofuels and bioproducts. *Agricultural and Biological Engineering , UF/IFAS Extension*, 1–4. Retrieved from <http://edis.ifas.ufl.edu/about.html>

Umagiliyage, A. L., Choudhary, R., Liang, Y., Haddock, J., & Watson, D. G. (2015). Laboratory scale optimization of alkali pretreatment for improving enzymatic hydrolysis of sweet sorghum bagasse. *Industrial Crops and Products*, 74, 977–986.
<http://doi.org/10.1016/j.indcrop.2015.05.044>

Velmurugan, R., & Muthukumar, K. (2012). Ultrasound-assisted alkaline pretreatment of sugarcane bagasse for fermentable sugar production: Optimization through response surface methodology. *Bioresource Technology*, 112, 293–299.
<http://doi.org/10.1016/j.biortech.2012.01.168>

Verardi, A., Bari, I. De, Ricca, E., & Calabro, V. (2012). Hydrolysis of Lignocellulosic Biomass : Current Status of Processes and Technologies and Future Perspectives. In M. A. P. Lima (Ed.), *Bioethanol* (p. 290). <http://doi.org/10.5772/23987>

Vinardell, M. P., Ugartondo, V., & Mitjans, M. (2008). Potential applications of antioxidant lignins from different sources. *Industrial Crops and Products*, 27(2), 220–223.
<http://doi.org/10.1016/j.indcrop.2007.07.011>

Wahid, M. (2017). Overview of the Malaysian Oil Palm Industry. *Malaysian Palm Oil Board*.

Wan Rosli, W. D., Law, K. N., Zainuddin, Z., & Asro, R. (2004). Effect of pulping variables on the characteristics of oil-palm frond-fiber. *Bioresource Technology*, 93(3), 233–240.
<http://doi.org/10.1016/j.biortech.2003.11.016>

Wan Zahari, M., Abu Hassan, O., Wong, H. K., & Liang, J. B. (2003). Utilization of oil palm frond - Based diets for beef and dairy production in Malaysia. *Asian-Australasian Journal of Animal Sciences*, 16(4), 625–634. <http://doi.org/10.5713/ajas.2003.625>

Wang, W., Kang, L., Wei, H., Arora, R., & Lee, Y. Y. (2011). Study on the decreased sugar yield in enzymatic hydrolysis of cellulosic substrate at high solid loading. *Applied Biochemistry and Biotechnology*, 164(7), 1139–1149. <http://doi.org/10.1007/s12010-011-9200-8>

Wang, Z., Bay, H., Chew, K., & Geng, A. (2014). High-loading oil palm empty fruit bunch saccharification using cellulases from *Trichoderma koningii* MF6. *Process Biochemistry*, 49(4), 673–680. <http://doi.org/10.1016/j.procbio.2014.01.024>

Wang, Z., Keshwani, D. R., Redding, A. P., & Cheng, J. J. (2010). Sodium hydroxide pretreatment and enzymatic hydrolysis of coastal Bermuda grass. *Bioresource*

Technology, 101(10), 3583–3585. <http://doi.org/10.1016/j.biortech.2009.12.097>

Wojtusik, M., Zurita, M., Villar, J. C., Ladero, M., & Garcia-Ochoa, F. (2016). Influence of fluid dynamic conditions on enzymatic hydrolysis of lignocellulosic biomass: Effect of mass transfer rate. *Bioresource Technology*, 216, 28–35.
<http://doi.org/10.1016/j.biortech.2016.05.042>

Wyman, C., Decker, S., Himmel, M., Brady, J., Skopec, C., & Viikari, L. (2004). Hydrolysis of Cellulose and Hemicellulose. In M. Dekker (Ed.), *Applied biochemistry and biotechnology* (pp. 994–1033). <http://doi.org/10.1201/9781420030822.ch43>

Xing, Y., Bu, L., Zheng, T., Liu, S., & Jiang, J. (2016). Enhancement of high-solids enzymatic hydrolysis of corncob residues by bisulfite pretreatment for biorefinery. *Bioresource Technology*, 221, 461–468. <http://doi.org/10.1016/j.biortech.2016.09.086>

Yemiş, O., & Mazza, G. (2011). Acid-catalyzed conversion of xylose, xylan and straw into furfural by microwave-assisted reaction. *Bioresource Technology*, 102(15), 7371–7378.
<http://doi.org/10.1016/j.biortech.2011.04.050>

Yuan, L., Wang, W., Pei, Y., & Lu, F. (2012). Screening and Identification of Cellulase-Producing Strain of *Fusarium Oxysporum*. *Procedia Environmental Sciences*, 12(Icese 2011), 1213–1219. <http://doi.org/10.1016/j.proenv.2012.01.410>

Yuan, Z., Long, J., Wang, T., Shu, R., Zhang, Q., & Ma, L. (2015). Process intensification effect of ball milling on the hydrothermal pretreatment for corn straw enzymolysis. *Energy Conversion and Management*, 101, 481–488.
<http://doi.org/10.1016/j.enconman.2015.05.057>

Zabed, H., Sahu, J. N., Boyce, A. N., & Faruq, G. (2016). Fuel ethanol production from lignocellulosic biomass : An overview on feedstocks and technological approaches. *Renewable and Sustainable Energy Reviews*, 66, 751–774.

Zahari, M. A. K. M., Zakaria, M. R., Ariffin, H., Mokhtar, M. N., Salihon, J., Shirai, Y., & Hassan, M. A. (2012). Renewable sugars from oil palm frond juice as an alternative novel fermentation feedstock for value-added products. *Bioresource Technology*, 110, 566–571.
<http://doi.org/10.1016/j.biortech.2012.01.119>

Zakaria, M. R., Hirata, S., Fujimoto, S., & Hassan, M. A. (2015). Combined pretreatment with hot compressed water and wet disk milling opened up oil palm biomass structure resulting in enhanced enzymatic digestibility. *Bioresource Technology*, 193, 128–134.
<http://doi.org/10.1016/j.biortech.2015.06.074>

Zhao, X., Cheng, K., & Liu, D. (2009). Organosolv pretreatment of lignocellulosic biomass for enzymatic hydrolysis. *Applied Microbiology and Biotechnology*, 82(5), 815–827.
<http://doi.org/10.1007/s00253-009-1883-1>

- Zheng, J., Choo, K., Bradt, C., Lehoux, R., & Rehmann, L. (2014). Enzymatic hydrolysis of steam exploded corncob residues after pretreatment in a twin-screw extruder. *Biotechnology Reports*, 3, 99–107. <http://doi.org/10.1016/j.btre.2014.06.008>
- Zhong, C., Jia, H., & Wei, P. (2017). Enhanced saccharification of wheat straw with the application of ultrasonic-assisted quaternary ammonium hydroxide pretreatment. *Process Biochemistry*, 53, 180–187.
- Zhu, L., O'Dwyer, J. P., Chang, V. S., Granda, C. B., & Holtzapple, M. T. (2008). Structural features affecting biomass enzymatic digestibility. *Bioresource Technology*, 99(9), 3817–28. <http://doi.org/10.1016/j.biortech.2007.07.033>
- Zhu, M. Q., Wen, J. L., Su, Y. Q., Wei, Q., & Sun, R. C. (2015). Effect of structural changes of lignin during the autohydrolysis and organosolv pretreatment on Eucommia ulmoides Oliver for an effective enzymatic hydrolysis. *Bioresource Technology*, 185, 378–385. <http://doi.org/10.1016/j.biortech.2015.02.061>
- Zulkefli, S., Abdulmalek, E., & Abdul Rahman, M. B. (2017). Pretreatment of oil palm trunk in deep eutectic solvent and optimization of enzymatic hydrolysis of pretreated oil palm trunk. *Renewable Energy*, 107, 36–41. <http://doi.org/10.1016/j.renene.2017.01.037>