



A Comprehensive Review on *Ocimum basilicum*

Balakrishnan Purushothaman^{1,2}, Ramalingam PrasannaSrinivasan², Purushothaman Suganthi², Balu Ranganathan^{3,4}, Jolius Gimbut⁵ and Kumaran Shanmugam^{1*}

¹Department of Biotechnology, Periyar Maniammai Institute of Science & Technology, Thanjavur – 613403, Tamilnadu, India; kumarans@pmu.edu

²TANBIO R&D Solution, Periyar Technology Business Incubator, Thanjavur – 613403, Tamilnadu, India

³Palms Connect Sdn Bhd, Shah Alam 40460, Selangor Darul Ehsan, Malaysia

⁴Palms Connect LLC, Showcase Lane, Sandy, UT 84094, USA

⁵Centre of Excellence for Advanced Research in Fluid Flow (CARIFF), Universiti Malaysia Pahang, Gambang 26300, Pahang Malaysia

Abstract

The genus *Ocimum* has a number of species that are used to treat different types of disorders and diseases from ancient times. In this genus, *O. basilicum* plays a vital role due to its various medicinal properties. It is universally cultivated as herbaceous, perennial plant but originated from Asian continent. *O. basilicum* is used as a culinary herb and also has a number of pharmacological activities to prevent or treat cardiovascular disorders, diabetes, menstrual cramps, digestive disorders, neuro-degenerated disorders and cancer. In addition to that, it has been reported for antioxidant, antimicrobial, and larvicidal activities. Chemical constituents such as linalool; eugenol; 1, 8-cineone; methyl eugenol and anthocyanins are mostly responsible for the above mentioned activities. Some traditional uses of this plant coincidences with experimental results. However, the studies conducted based on its traditional use are negligible. This review is an attempt to provide a pharmaceutical perspective of *Ocimum basilicum*.

Keywords: Cancer, Chemical Constituents, Immunomodulatory, Pharmacological Activity, GC-MS

1. Introduction

The genus *Ocimum* belongs to the family Lamiaceae. There are about 150 species of *Ocimum*¹. The genus *Ocimum* has a number of species that are used to treat different types of ailments from ancient time, especially the species *Ocimum basilicum*². It is otherwise known as sweet basil. It is a universally cultivated herbaceous, perennial plant³. It is a popular herb used in Italian and Southeast cuisines of Thailand and Vietnam⁴. It has numerous potent activities due to the metabolites present in it. As a consequence of its virulent metabolites, it is used in traditional medicine⁵⁻⁶ and also as an ornamental plant⁷. The extracts of essential oils of *Ocimum basilicum*

are used as the flavors for the food products. It is used as a kitchen herb, culinary herb and ornamental herb⁸. It has also been used as commercial fragrances, flavors and to improve the food products shelf life⁹⁻¹¹.

O. basilicum has been reported numerously in areas related to agriculture, food, and pharmacology. Hence, this review would shed more lights on different dimension of *O. basilicum* to the researchers. At the same time, the disappearance of *O. basilicum* in certain regions is increasing day by day so, it is vital to create awareness on the medicinal importance of this plant to prevent its extinction. All these things made us to pen down this review which is mainly focused on the pharmaceutical prospective of *Ocimum basilicum*.

*Author for correspondence

2. Historical Perspectives

Ayurveda and Unani physicians used this plant in various forms to cure ringworm, rashes, and other skin troubles. Since 1930, *Ocimum* is targeted to study the chemical combination of basil oil and also as a kitchen herb and decorative plant¹². The genus *Ocimum* covers over 150 species of herbs in addition shrubs¹³⁻¹⁴. It is broadly used in food, pharmaceutical, cosmetic, aromatherapy and perfumery industries¹⁵⁻¹⁶ and used all over the world because of its sense of taste especially, in Mediterranean food court¹⁷. The infusions of *Ocimum basilicum* are extensively used as old style medicine to shrink plasma lipid content in Mediterranean areas¹⁸. Above 100 herbs and shrubs of genus *Ocimum* originated from tropical and sub-tropical region and cultivated around the world¹⁹. The taxonomical hierarchy of *O. basilicum* is shown in Table 1.

Table 1. Taxonomical Hierarchy of *O. basilicum*

Kingdom	Plantae
Phylum	Magnoliophyta
Class	Magnoliopsida
Order	Lamiales
Family	Lamiaceae
Genus	<i>Ocimum</i>
Species	<i>basilicum</i>

3. Traditional Claims

In Turkey, *Ocimum basilicum* is used as a folk medicine and traditional Uyghur medicine to prevent and treat diabetics²⁰ and cardiovascular disorders^{21, 22}. In Indian Siddha medicine, it is used for treating pimples on face²³. Traditionally basil has been used to treat headaches, coughs, diarrhea and kidney malfunctions¹. It also used in the treatment of insect stings, snake bites and skin infections externally. In Bulgaria, it is used as a folk medicine for the treatment of aches and pains²⁴. In Spain, it is used as a sedative²⁵.

4. Perfumery

The ethanolic steam distillation extract of flowers is used as pleasant smelling compound in perfumes²⁶. The presence of essential oils determines the aroma of the *O. basilicum*.

5. Biogeography and Ecology

Basil is a vital oil bearing herb which can be grown in various environmental conditions on a wide range of terrain. Among varied aromatic and medicinal crops, basil is one of the crops for utilization of sodic wasteland owing towards its tolerance to higher salt, pH and exchangeable sodium percentage²⁸. Hence, *O. basilicum* habitation varies from tropical areas of Asia, Africa, Central and south America¹ but it is highly cultivated in Iran, Japan, China and Turkey²⁹.

6. Phytochemical Studies

O. basilicum majorly contains about 20 compounds such as linalool, estragole, methyl eugenol, 1, 8-cineole, etc., which has been identified by GC-MS³⁰. Camphor, limonene, thymol, citral, α -linalool, β -linalool, estragole, are the monoterpenes of *O. basilicum*. Methyl eugenol is the active compound of *Ocimum basilicum*. Chichoric acid was found in the fresh basil leaves³¹. Crude extract of various morphological parts of *Ocimum* are rich in phenolics. The intensely purple pigment of flower is due to the presence of anthocyanins³². Linalool (52.42%), methyl eugenol (18.74%), 1,8-cineol (5.61%) are the major compounds in *O. basilicum* which are isolated by the HPLC method³⁰. Myrcene, borneol and neral are the minor compounds present at 5%, 9%, 8% w/w respectively. Fourteen different anthocyanins have been isolated by HPLC within that 11 has cyanidin based pigments and 3 has peonidin based pigments³². It is also used in the preparation of cosmetics and perfumes. Extracts of *Ocimum* show strong inhibitory effects on HIV-1 reverse transcriptase and platelet aggregation³³. Chemical constituents of *O. basilicum* and their biological activity are shown in Table 2. The chemical structures of *O. basilicum* constituents are shown in Figure 1.

7. Effluent Treatment

Over 50,000 tons of diverse artificial colorants are yearly produced as of many industrialized processes and around 1-10 % of them are settled into the surroundings⁸⁴. Due to synthetic source and compound molecular structure, dyes are challenging to biodegrade when at large to the environment and their half-finished degradation frequently produces toxic compounds⁸⁵. There are

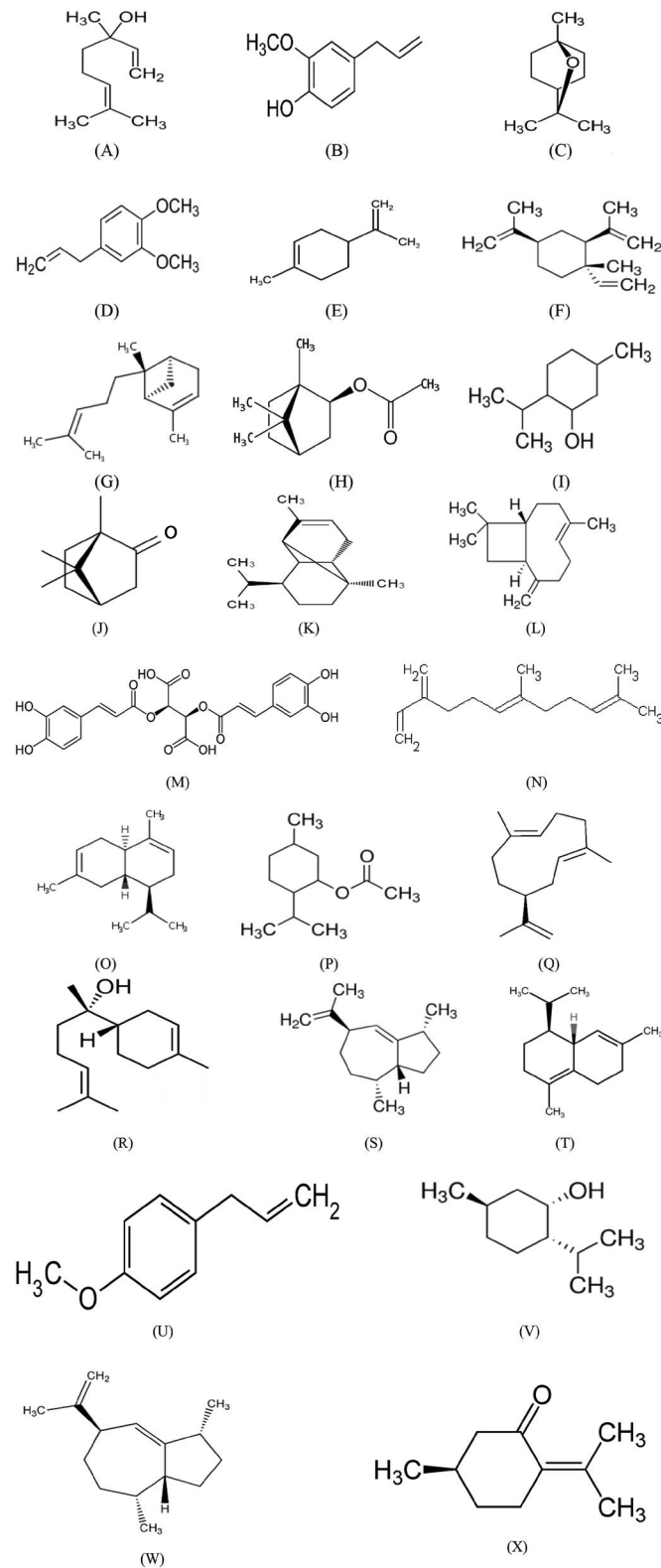


Figure 1. Chemical structures of (A) Linalool, (B) Eugenol, (C) 1,8-cineole, (D) Methyl eugenol, (E) Limonene, (F) β -elemene, (G) α -bergamotene, (H) Bornyl acetate, (I) Menthol, (J) Camphor, (K) α -copaene, (L) β -caryophyllene, (M) Chicoric acid, (N) Farnesene, (O) β -cadinene, (P) Menthyl acetate, (Q) Germacrene, (R) α -bisabolol, (S) δ -gurjunene, (T) δ -cadinene, (U) Estragole, (V) Neo isomenthol, (W) Guaiene, (X) Pulegone.

Table 2. Extraction methods, chemical constituents and biological activities of *O. basilicum*

S.No	Chemical constituents	Chemical class	Extraction method	Biological activity
1	Linalool	Monoterpene alcohol	Solvent free microwave extraction and conventional hydrodistillation ³⁴	Antihyperalgesic, Antinociceptive ³⁵
2	Eugenol	Phenylpropanoid	Steam distillation ³⁶	Neuro-protective, Anti-cancer, local anesthetic ³⁷
3	1,8-cineole	Monoterpenoid	Hydrodistillation ³⁸	Antiulcer, Wound healing activity ³⁹
4	Methyl eugenol	Phenylpropene	Hydrodistillation ⁴⁰	Anticonvulsant and Anesthetic ⁴¹
5	Limonene	Monoterpene	Solvent at high pressure high temperature extraction ⁴²	Motor relaxant, Anti-inflammatory ⁴³
6	β -elemene	Sesquiterpenoid	Hydro and glycol distillation ⁴⁴	Antineoplastic, Anticancer ⁴⁵
7	α -bergamotene	Sesquiterpene	Cold maceration ⁴⁶	Abiotic stresses release ⁴⁷
8	Bornyl acetate	Acetate ester	Microwave assisted extraction ⁴⁸	Analgesic, Anti-inflammatory ⁴⁹
9	Menthol	Alcohol	Soxhlets solvent extraction ⁵⁰	Local anesthetic ⁵¹
10	Camphor	Terphenoid	Liquid liquid extraction ⁵²	Decongesting, Antipruritic, counterirritant ⁵³
11	α -copaene	Sesquiterpene	Supercritical fluid extraction ⁵⁴	Cytotoxic, Anti-genotoxic, Antioxidant ⁵⁵
12	β -caryophyllene	Sesquiterpene	Supercritical fluid extraction ⁵⁶	Antibiotic, Antioxidant, anti-Carcinogenic ⁵⁷
13	Chicoric acid	Phenylpropanoid	Solvent microwave extraction ⁵⁸	Antioxidant, Immunostimulatory ⁵⁹
14	Farnesene	Sesquiterpene	Steam distillation ⁶⁰	Anti-oxidant, Anti-Insecticidal ⁶¹
15	β -cadinene	Bicyclic sesquiterpene	Supercritical fractioned extraction ⁶²	Antinociceptive, Antiproliferative ⁶³
16	Menthyl acetate	Monoterpene	Solid-phase microextraction ⁶⁴	Antioxidant, Antibacterial ⁶⁵
17	Germacrene	Sesquiterpene	Solvent extraction ⁶⁶	Analgesic ⁶⁷ , Anti-inflammatory ⁶⁸
18	α -bisabolol	Sesquiterpene alcohol	Supercritical carbon dioxide extraction and in situ extraction ⁶⁹	Analgesic ⁷⁰ , Antibiotic ⁷⁰ , Anticancer ⁷¹
19	δ -gurjunene	Cyclo aromadendrane sesquiterpenoid	Supercritical fluid extraction method ⁷²	Antitumor, Anti-inflammatory, Anti-oedematous ⁷³
20	δ -cadinene	Bicyclic sesquiterpene	Headspace solid-phase Microextraction ⁷⁴	Antioxidant, Antimicrobial ⁷⁵
21	Estragole	Phenylpropene	Hydrodistillation ⁷⁶	Neuronal excitability ⁷⁷
22	Neo isomenthol	Menthane monoterpenoid	Steam distillation ⁷⁸	Nasal sensation ⁷⁹
23	Guaiene	Sesquiterpene	Steam distillation ⁸⁰	Antiplatelet, Antithrombotic, Aphrodisiac, Antidepressant ⁸¹
24	Pulegone	Monoterpenoid	Salting-out assisted liquid-liquid extraction ⁸²	Antinociceptive ⁸³

several methods which are active to reduce the dyes⁸⁶. Coagulation-flocculation is one of the furthestmost effective and inexpensive processes for treatment of wastewater containing dye⁸⁷. The plant-based coagulants are harmless to human health, cost effective and biodegradable⁸⁸. They produce a lesser amount of sludge and do not modify pH of the treated water⁸⁹. The *Ocimum basilicum* was used as a natural coagulant for the treatment of textile waste water⁸⁴. The seeds obtained from this plant are also used for biosorption of copper⁹⁰. The plant is also used as a potential bio-sorbent for chromium uptake with its high biosorption capacity of the seeds⁹¹. The plant is adaptable for all soil conditions, this plant also adaptable to grow in sodic soil which was proved by the bacterial isolates from the soil where it is isolated⁹¹.

8. Analytical Analysis

8.1 GC-MS

In GC-MS, helium gas Agilent-Gas chromatography was equipped with HP-innowax fused silica capillary column (30 mm × 0.25 mm, film thickness 0.25 μm)²⁹. The GC-MS analysis were performed with a carloErba HRGC 5160 mega gas chromatography equipped with FID and a Hitachi 2000 integrator¹⁵. For the different types of basil Automated HS-SPME of basil, volatiles was performed using a CombiPal multipurpose sampler connected to a GC-ITMS system⁹²⁻⁹⁴.

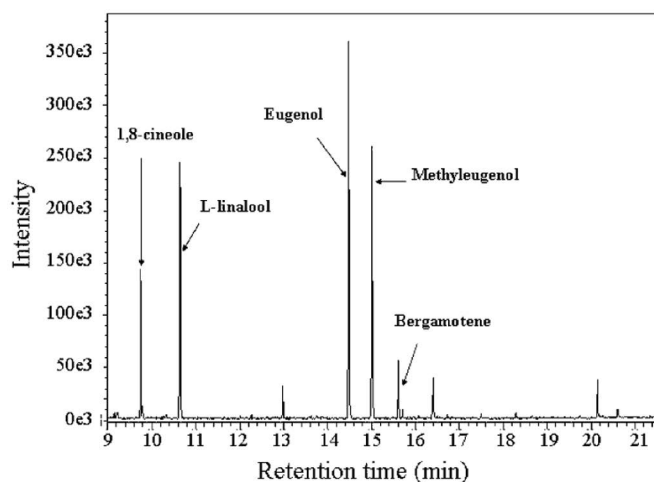


Figure 2. GC-MS image of Methyl *tert*-butyl ether extract of *O. basilicum* (50).

“Reprinted (adapted) with permission from Effect of Chitosan on the Biological Properties of Sweet Basil (*Ocimum basilicum* L.)⁹⁵”.

The GC-MS study (Figure 2) confirms the presence of terpenoids such as eugenol, methyl eugenol, 1, 8-cineole, L-linalool, bergamotene in the methyl *tert*-butyl ether extracts of *O. basilicum* at different retention time. It also reveals that the eugenol has the higher percentage (33%) in the composition when compared with others. Identification of these compounds was done by GC-MS equipment from Shimadzu, model no: GC-17A with a DB-5 capillary column (60mm-0.25 mm, thickness 0.25 μm) and the detector used was GCMS-QP 5050 mass spectrometer⁵⁰.

8.2 SEM

A SEM study has examined the leaves at various stages of development which is dehydrated in the ethanolic series and the solvent-substituted liquid carbon-dioxide and sputter coated with gold. The material was examined using a Jeol JSM 35 SEM⁹⁶.

8.3 HPLC

The aqueous extract of basil has been separated to specific compounds with the help of HPLC²¹. The injector and detector parts of the HPLC are maintained at 523 K and 573 K respectively⁹⁷. Rosmarinic acid and caffeic acid were quantified in all basil samples using a dual pump waters HPLC system⁹⁸. The Identification of the essential oil compounds was based on the retention time⁹⁹. It has been proved by the presence of peaks in various retention time at 280 nm. For the above analysis, Shimadzu LC-10 AS with diode array detector (Shimadzu, SPD-10A) and the reverse phase Spherisorb ODS II (RP18), analytical column (250 mm × 4.6 mm, particle size 5 μm) was used.

9. Pharmacological Activities

It is mostly used to treat anxiety, cold, fevers, migraines, diabetes, menstrual cramps, sinusitis, cardiovascular diseases, nerve pain, insect bites, and headache⁹⁹⁻¹⁰¹. Moreover, it also acts as an anticonvulsant^{96,25}, anti-hyperlipidemic¹⁰², anti-inflammatory¹⁰³, anti-oxidant¹⁰⁴⁻¹⁰⁵, antiplatelet property¹⁰², anti-thrombotic¹⁷,

anti-microbial⁹⁻¹¹, insecticidal²⁵, immunomodulatory¹⁰⁶ and cytotoxicity effect¹⁰⁷. It also acts against neuro-degenerated disorders and digestive disorders. It is also used as cardiogenic and abdominal pain reliever¹¹¹. Moreover, it also has an effect on the coccidial activity in the broiler chicks¹¹². It also has spasmolytic, carminative, hepatoprotective, diuretic and stimulating properties that lead to the production of various drugs from *Ocimum* in pharma industries.

9.1 Antimicrobial

Numerous therapeutic plants have their own antimicrobial agents have gained popularity in recent years²⁶. Due to the accidental and extensive practice of antibiotics for the action of infectious and communicable diseases, the pathogenic microorganisms remain developed into multiple disease resistance at recent years¹⁰⁸. One of the finest methods toward to resolve this problem is to look for new therapeutic agents from plants which contain antimicrobial activities against the pathogenic microorganisms¹⁰⁹. Extracts obtained from *Ocimum basilicum* has a rich source of flavor compounds and volatile oils which contain variety of compounds and these compounds possess antimicrobial activity^{106,33}. The essential oil from *O. basilicum* is shown to have an inhibitory effect on *Aspergillus ochraceus*¹¹⁰. The hairy root cultures of *Ocimum basilicum* is used to achieve antimicrobial activity against bacteria and fungi by inhibiting z^{++} ion assays in contradiction of the organisms such as *P. aeruginosa* strains (PAO1) (PA14), *A. rhizogenes*, *P. fluorescens*, *X. campestris*, *E. carotovora*, *P. drechsleri*, *Phytophthora megasperma*, *Phytophthora parasitica*, *A. niger*, *Rhizoctonia solani*, *F. oxysporum*, *P. aphanidermatum*, *P. ultimum*, *Versicillium dahailia*, *Alternaria solani* and *Alternaria brassicae*¹¹¹. The leaf extract is prepared into a powder and soaked with 95% ethanol and kept free of oil obtained from this plant also shows antimicrobial activity by ethanolic extracts from its leaves which were observed against *E. coli* and *Staphylococcus aureus* with its inhibition level²⁶. Anti-fungal activity of the *O. basilicum* is performed in 2045 fungal species which showed more effect on the *Aspergillus flavus* which is toxigenic in nature¹¹². Basil oil also has been reported for its significant effect in *in vitro* studies¹¹³.

9.2 Antioxidant

Ocimum basilicum contains several active antioxidant compounds¹¹⁴. The oil obtained from this plant also shows antioxidant properties to assist sperm parameters and enhance sperm quality in rats to increase spermatogenesis²⁶. The antioxidant property is due to the polyphenoid rosmarinic acid which is a derivative of cinnamic acid³².

9.3 Anti-Hyperlipidemic

The aqueous *O. basilicum* extract significantly lowered both plasma triglycerides (TG) and cholesterol in acute hyperlipidemia induced by Triton WR-1339 in rats¹⁰².

9.4 Anticonvulsant

Hydro distillation of aqueous leaf extract of *Ocimum basilicum* has strong anticonvulsant activity majorly due to the eugenol present in it²⁵.

9.5 Anti-Inflammatory

Solvent extraction of methanolic extract of *Ocimum basilicum* aerial parts have potent anti-inflammatory activity against macrophage (RAW264.7), human chondrosarcoma (SW1353) cell lines and human primary chondrocytes¹⁰³.

9.6 Antiplatelet

The aqueous extract of *Ocimum basilicum* aerial parts was concentrated in a rotatory evaporator under vacuum at 65°C and studied against its antiplatelet activity using thrombin (0.5 U/ml) and ADP (5µM) as agonists. It inhibits ADP-induced platelet aggregation by 13%, 28.2%, 30.5%, 44.7% and 53% at a dose of 1, 2, 3, 4 and 5 g/l, respectively¹⁰².

9.7 Anti-Thrombotic

Platelet aggregation induced by ADP (5 AM) and thrombin (4 UI), and thrombus weight in an arteriovenous thrombosis (AVT) model were tested after 2 weeks treatment with 15, 75 and 375 mg/kg OBL orally in rats, compared to 8.8 mg/ kg/day aspirin Thrombin-induced aggregation reached 33%, 22%, 21% for the aqueous extract of *O. basilicum*¹⁷.

9.8 Insecticidal Activity

Usage of synthetic insecticides causes some significant consequences such as environmental pollution, pests or vector resistance and toxicity to other non-target creatures including human beings¹⁰⁷. The vector mosquitoes are accomplished of communicating potential pathogens to human beings and them in charge for several infectious diseases like malaria, filariasis, Japanese encephalitis, yellow fever, dengue and chikungunya¹¹⁵. Moreover, the essential oil extract from the leaf *O. basilicum* was evaluated against *Culex tritaeniorhynchus*, *Aedes albopictus* and *Anopheles subpictus* to predict the larvicidal activity¹¹⁶. It has its own significant effect against third stage larvae of *Culex tritaeniorhynchus*, *Aedes albopictus* and *Anopheles subpictus* with an LC₅₀ and 9.75 ppm and LC₉₀ values of 23.44, 21.17 and 18.56 ppm respectively⁹⁷.

9.9 Immunomodulatory

The extracts from the plant *Ocimum basilicum* indicates its immunomodulatory action taking place in the cellular level, including platelet anti-aggregant property and inhibitory activity to counter HIV-1 reverse transcriptase^{106,33}. The mechanisms underlying the hypolipidemic properties of *Ocimum basilicum* and anti-atherogenic potential of this plant at the cellular level is mysterious¹⁸.

9.10 Cytotoxicity Effect

The cytotoxicity effect of the active compounds (methyl cinnamate, linalool) of leaves extract of *O. basilicum* was performed by methyl thiazol tetrazolium (MTT) assay. In Table 3, the different concentration of extracts were affected the viability of human cancer cell line such as HeLa, HEp-2 and NIH 3T3 has been tabulated along with their IC₅₀ value.

The HeLa and HEp-2 cells were grown in Eagles Minimum Essential Medium containing 10% foetal bovine serum (FBS) and NIH 3T3 fibroblasts were grown in Dulbecco's Modified Eagles Medium containing 10% FBS. For the screening experiment, the cells were seeded into 96-well plates in 100 mL of the respective medium containing 10% FBS, at plating density of 10,000 cells per well, and incubated at 37°C, 5% CO₂, 95% air and 100% relative humidity for 24 h. The essential oil was solubilized in dimethylsulphoxide (DMSO) and diluted in respective medium containing 1% FBS. After 24 h, the

Table 3. IC₅₀ value of different cancer cell lines

S. No	Cancer cell line type	Parts of the plant	IC ₅₀ (µg/mL)
1	HeLa ¹⁰⁷	Leaves	90.0
2	MCF-7 ^{107,115}	Leaves	260.3–270.3
3	NIH 3T3 mouse embryonic fibroblasts ¹⁰⁷	Leaves	120.7
4	Ln-CaP ^{107,115}	Leaves	70.1–172
5	P388 KB ^{107,115}	Leaves	36.2
6	HEp-2 ¹⁰⁷	Leaves	96.5
7	EAC ^{109,115}	Leaves	30.33

medium was replaced with respective medium with 1% FBS containing the oil at various concentrations (12.5, 25, 50, 100, 200, and 300 mg/mL) and incubated at the previously set parameters⁶³⁻⁶⁴. Triplicate was maintained and the medium without the oil served as control. After 48 h, 10 mL of MTT (5 mg/ 1mL) in phosphate-buffered saline was added to each well and incubated at 37C for 4 h. The medium with MTT was then flicked off and the formed formazan crystals were solubilized in 100 mL of DMSO and absorbance was measured at 570 nm using microplate reader¹⁰⁷.

10. Diseases Affecting the *O. basilicum* sp

Although various medicinal properties has been exhibited by *O. basilicum*, it also been affected by some diseases like wilt, mold, spot and rot. Most diseases occurs in leaves and stem of basil plants due to the infection of fungi. However it can be eradicated by using proper natural fungicide to maintain its original medical nature. Some of the diseases affecting basil are shown in Table 4.

11. Companies Manufacturing *O. basilicum* Products

Basil products were commercially used all over the world for its nutritive purposes and medicinal values. Because of its multi dimensional use, many manufacturers produce basil products. Most of the basil products producing companies are located in Asian countries as shown in Table 5.

12. Adverse Effects and Caution

Quercetin (a flavonoid) may be a co-carcinogen in bracken fern (*Pteridium aquilinum*). It has been suggested that it may interact with bovine papilloma

Table 4. Diseases affecting *O. basilicum*

Sl. No	Disease Name	Parts of the Plant	Symptoms	Causing agents
1.	Fusarium wilt ¹¹⁸	Leaves and stem	Yellow and wilting leaves, death of the plant	<i>Fusarium oxysporum f.</i>
2.	Gray mold ¹¹⁹	Leaves and stem	leaves dying and dropping from plant	<i>Botrytis cinerea</i>
3.	black spot ¹²⁰	Leaves and stem	Black spots on leaves	<i>Colletotrichum gloeosporioides</i>
4.	Basal rot ¹²¹	Leaves	Wilting of leaves	<i>Rhizoctonia solani</i>
5.	Cercospora leaf spot ¹¹⁸	Leaves	Circular to irregular dark spots on leaves	<i>Cercospora ocimicola</i>
6.	Downy mildew ¹¹⁹	Leaves	Yellowing leaves	<i>Peronospora belbahrii</i>
7.	Leaf spot ¹¹⁹	Leaves	streaks on stems	<i>Pseudomonas spp</i>
8.	Root rot ¹¹⁹	Stem	Failure of seeds to germinate	<i>Rhizoctonia solani</i>

Table 5. Companies manufacturing basil products

S.No	Company Name	Country	Products	Source
1.	Global Merchants	India	Basil oil.	www.global-merchants.in/
2.	Right Future International	India	Leaves for flavoring purpose.	www.rightfutureinternational.co.in/
3.	Frontier Natural Products Co-op.	US	Basil leaf for flavors.	www.frontiercoop.com/
4.	Haridass Aggarwal & sons	India	Leaves for flavoring.	www.indiamart.com/haridas-agarwal-sons/
5.	Natural Healthy concepts	India	Basil oils.	www.naturalhealthyconcepts.com/
6.	Zhongbei Northland Bio-chem industry Co.Ltd.	China	Basil oils for tonic purpose.	http://daxinganlinglily.en.china.cn/
7.	Jkh exports	India	Leaves for aroma products.	www.jkhexports.com/
8.	Dujardin Foods Nv	Belgium	Basil oils.	www.dujardin-foods.com/
9.	Jdg seeds company	India	Basil seeds.	www.jdgseeds.com/
10.	Silver line Chemicals	India	Basil essential oils.	www.silverlinechemicals.com/

virus type 4, leading to malignant epithelial papillomas¹²². Caffeic acid and P-coumaric acid (phenolic acids) may inhibit digestion of plant cell walls in ruminants, because of their antimicrobial activity. When these phenolic acids are metabolized by rumen microbes, benzoic acid, 3-phenylpropionic acid and cinnamic acid may be formed. On further detoxification, hippuric acid is formed. 3-phenylpropionic acid and hippuric acid can decrease metabolic efficiency and also decreases productivity. Safrole, which was used to flavor sodas, was banned as a food additive in the US because it has the potential to cause cancer in rats¹²². Moreover, it should not be used during pregnancies.

13. Conclusion and Future Perspectives

This review majorly focused on the distinctive pharmacological activities of *O. basilicum* and the specific extraction procedures for the chemical constituents present in it. Basil possesses anticonvulsant, anti-hyperlipidemic, anti-inflammatory, anti-oxidant, antiplatelet property, anti-thrombotic, anti-microbial, insecticidal, immunomodulatory and cytotoxicity activities. Moreover, it also delivers those chemical constituents which is very helpful for the sustainable development of drugs for human consume. Furthermore, it also speaks about some of the manufacturers of basil products. On the converse, basil also has some adverse

effects, so it should be consumed with proper prescribed medications. In the future, research may make a transgenic basil plant to eradicate the environmental stress of the plant and to address a specific chemical constituent to act against various human diseases. Here we imaged the rough and trough of basil, so it will be very helpful for the future researcher to work in different dimensions of basil to make new products. *O. basilicum* is the important product of nature so “we would use and should protect it”.

14. Acknowledgement

We thank American Cancer Society (ACS) for the reuse of Figure 2 in this review article. We also acknowledge Jaina Afrin Mohammed Farook of Department of Biotechnology, Periyar Maniammai Institute of Science & Technology for her kind assistantship.

15. Conflict of Interest

There is no any conflict of interest among the authors.

16. References

- Simon JE, Morales MR, Phippen WB, Vieira RF, Hao Z. Basil: A source of aroma compounds and a popular culinary and ornamental herb. Perspectives on New crops and new uses. Alexandria: ASHS Press. 1999.
- Siddiqui BS, Bhatti HA, Begum S, Perwaiz S. Evaluation of the Anti-mycobacterium activity of the constituents from *Ocimum basilicum* against *Mycobacterium tuberculosis*. *Journal of Ethnopharmacology*. 2012; 144: 220–2. <https://doi.org/10.1016/j.jep.2012.08.003> PMID:22982011
- Bantis F, Ouzounis T, Radoglou K. Artificial LED lighting enhances growth characteristics and total phenolic content of *Ocimum basilicum* but variably affects transplant success. *Scientia Horticulturae*. 2016; 198: 277–83. <https://doi.org/10.1016/j.scienta.2015.11.014>
- Snoussi M, Dehmani A, Noumi E, Flamini G, Papetti A. Chemical composition and antibiofilm activity of *Petroselinum crispum* and *Ocimum basilicum* essential oils against *Vibrio* spp. Strains. *Microbial Pathogenesis*. 2016; 90: 13–21.
- Bora KS, Arora S, Shri R. Role of *Ocimum basilicum* L. in prevention of reperfusion induced cerebral damage, and motor dysfunctions in mice brain. *Journal of Ethnopharmacology*. 2011; 137: 1360–5. <https://doi.org/10.1016/j.jep.2011.07.066> PMID:21843615
- Loughrin JH, Kasperbauer MJ. Light reflected from colored mulches affects aroma and phenol content of sweet basil (*Ocimum basilicum* L.) Leaves. *Journal of Agricultural and Food Chemistry*. 2001; 49: 1331–5. <https://doi.org/10.1021/jf0012648>
- Javanmardi J, Stushnoff C, Locke E, Vivanco JM. Antioxidant activity and total phenolic content of Iranian *Ocimum* accessions. *Food Chemistry*. 2003; 83: 547–50. [https://doi.org/10.1016/S0308-8146\(03\)00151-1](https://doi.org/10.1016/S0308-8146(03)00151-1)
- Gulcin I, Elmastas M, Aboul-Enein HY. Determination of antioxidant and scavenging activity of Basil (*Ocimum basilicum* L. family lamiaceae) assayed by different methodologies. *Phytotherapy Research*. 2007; 21: 354–61. <https://doi.org/10.1002/ptr.2069> PMID:17221941
- Makinen S, Paakkonen K, Hiltunen R, Holm Y. Processing and use of basil in foodstuffs, beverages and in food preparation. *Basil: the genus Ocimum*. Netherlands: Harwood Academic Publishers. 1999.
- Suppakul P, Miltz J, Sonneveld K, Bigger SW. Antimicrobial properties of basil and its possible application in food packaging. *Journal of Agricultural and Food Chemistry*. 2003; 51: 3197–207. <https://doi.org/10.1021/jf021038t> PMID:12744643
- Nguyen PM, Niemeyer ED. Effects of nitrogen fertilization on the phenolic composition and antioxidant properties of basil (*Ocimum basilicum* L.). *Journal of Agricultural and Food Chemistry*. 2008; 56: 8685–91. <https://doi.org/10.1021/jf801485u> PMID:18712879
- Chang SS, Ostric-Matijasevic B, Hsieh OAL, Huang CL. Natural antioxidant from rosemary and sage. *Journal of Food Science*. 1977; 42: 1102–7. <https://doi.org/10.1111/j.1365-2621.1977.tb12676.x>
- Lachowicz KJ, Jones JP, Briggs DR, Bienvenu FE, Palmer MV, Mishra V, Hunter MM. Characteristics of Plants and Plant Extracts from Five Varieties of Basil (*Ocimum basilicum* L.) Grown in Australia. *Journal of Agricultural and Food Chemistry*. 1997; 45(7): 2660–5. <https://doi.org/10.1021/jf960791h>
- Khazaeia N, Esmailia M, Djomehb ZE, Ghasemlouc M, Joukid M. Characterization of new biodegradable edible film made from basil seed (*Ocimum basilicum* L.) gum. *Carbohydrate Polymers*. 1997; 102: 199–206. <https://doi.org/10.1016/j.carbpol.2013.10.062> PMID:24507273
- Marotti M, Piccaglia R, Giovannelli E. Differences in essential oil composition of basil (*Ocimum basilicum* L.) Italian cultivars related to morphological characteristics. *Journal of Agricultural and Food Chemistry*. 1996; 44: 3926–9. <https://doi.org/10.1021/jf9601067>

16. Zheljzkov VD, Cantrell CL, Tekwani B, Khan SI. Content, composition, and bioactivity of the essential oils of three basil genotypes as a function of harvesting. *Journal of Agricultural and Food Chemistry*. 2008; 56: 380–5. <https://doi.org/10.1021/jf0725629> PMID:18095647
17. Tohti I, Tursun M, Umar A, Turddi S, Imin H, Nicholas M. Aqueous extracts of *Ocimum basilicum* L. (Sweet basil) decrease platelet aggregation induced by ADP and thrombin in vivo arteriovenous shunt thrombosis in vivo. *Thrombosis Research*. 2006; 118: 733–9. <https://doi.org/10.1016/j.thromres.2005.12.011> PMID:16469363
18. Bravo E, Amrani S, ArizM, Harnafi H, Napolitano M. *Ocimum basilicum* ethanolic extract decreases cholesterol synthesis and lipid accumulation in human macrophages. *Fitoterapia*. 2008; 79: 515–23. <https://doi.org/10.1016/j.fitote.2008.05.002> PMID:18620033
19. Libera Z, Carovic-Stanko K, Politeoc O, Strikic F, Kolak I, Milos M, Satovic Z: Chemical Characterization and Genetic Relationships among *Ocimum basilicum* L. Cultivars. *Chemistry & Biodiversity*. 2011; 8(11): 1978–89. <https://doi.org/10.1002/cbdv.201100039> PMID:22083911
20. Rai V, Mani UV, Iyer UM. Effect of *Ocimum sanctum* leaf powder on blood lipoproteins, glycated proteins and total Amino acids in patients with Non-insulin-dependent Diabetes Mellitus. *Journal of Nutritional & Environmental Medicine*. 2009; 7: 113–8.
21. Harnafia H, Azizb M, Amrania S. Sweet basil (*Ocimum basilicum* L.) improves lipid metabolism in hypercholesterolemic rats. *The European e-Journal of Clinical Nutrition and Metabolism*. 2009; 4: 181–6. <https://doi.org/10.1016/j.eclnm.2009.05.011>
22. Umar A, Zhou W, Abdusalam E, Tursun A, Reyim N, Tohti I, Moore N. Effect of *Ocimum basilicum* L. on cyclo-oxygenase isoforms and prostaglandins involved in thrombosis. *Journal of Ethnopharmacology*. 2014; 152: 151–5. <https://doi.org/10.1016/j.jep.2013.12.051> PMID:24412551
23. Tsai KD, Lin BR, Perng DS, Wei JC, Yu YW, Cherng JM: Immunomodulatory effects of aqueous extract of *Ocimum basilicum* (Linn.) and some of its constituents on human immune cells. *Journal of Medicinal Plants Research*. 2011; 5: 1873–83.
24. Opalchenova G, Obreshkova D. Comparative studies on the activity of basil-an essential oil from *Ocimum basilicum* L. against multidrug resistant clinical isolates of the genera *Staphylococcus* and *Pseudomonas* by using different test methods. *Journal of Microbiological Methods*. 2003; 54: 105–10. [https://doi.org/10.1016/S0167-7012\(03\)00012-5](https://doi.org/10.1016/S0167-7012(03)00012-5)
25. Freire MM, Marques OM, Costa M. Effects of seasonal variation on the central nervous system activity of *Ocimum gratissimum* L. essential oil. *Journal of Ethnopharmacology*. 2006; 105: 161–6. <https://doi.org/10.1016/j.jep.2005.10.013> PMID:16303272
26. Khalil A. Antimicrobial activity of ethanolic extracts of *Ocimum basilicum* leaf from Saudi Arabia. *Biotechnology*. 2013; 12: 61–4. <https://doi.org/10.3923/biotech.2013.61.64>
27. Hassanpouraghdam MB, Hassani A, Vojodi L, Farsad-akhtar N. Drying Method Affects Essential Oil Content and Composition of Basil (*Ocimum basilicum* L.). *Journal of Essential Oil Bearing Plants*. 2010; 13: 759–66.
28. Aishwath OP, Nibauria SV. Utilization of problematic soil and water resources through aromatic plants. *Ecology, Environment and Conservation*. 2009; 15(4): 715–24.
29. Alves-Silva JM, Dias dos Santos SM, Pintadob ME, Perez-Alvarezc JA, Fernandez-Lopezc J, Viuda-Martosc M. Chemical composition and in vitro antimicrobial, antifungal and antioxidant properties of essential oils obtained from some herbs widely used in Portugal. *Food Control*. 2013; 32: 371–8. <https://doi.org/10.1016/j.foodcont.2012.12.022>
30. Radulovic NS, Blagojevic PD, Miltojevic AB. α -Linalool marker compound of forged/synthetic sweet basil (*Ocimum basilicum* L.) essential oils. *Journal of the Science of Food and Agriculture*. 2013; 93: 3292–303. <https://doi.org/10.1002/jsfa.6175> PMID:23584979
31. Lee J, Scagel CF. Chicoric acid found in basil (*Ocimum basilicum* L.) leaves. *Food Chemistry*. 2009; 115: 650–6. <https://doi.org/10.1016/j.foodchem.2008.12.075>
32. Phippen WB, Simon JE. Anthocyanins in basil (*Ocimum basilicum* L.). *Journal of Agricultural and Food Chemistry*. 1998; 46: 1734–8. <https://doi.org/10.1021/jf970887r>
33. Yamasaki K, Nakano M, Kawahata T, Mori H, Otake T, Ueba N, Oishi I, Inami R, Yamane M, Nakamura M, et al., Anti-HIV-1 activity of herbs in Labiatae. *Biological and Pharmaceutical Bulletin*. 1998; 21: 829–33. <https://doi.org/10.1248/bpb.21.829> PMID:9743251
34. Galhiane MS, Rissato SR, Chierice GO, Almeida MV, Silva LC. Influence of different extraction methods on the yield and linalool content of the extracts of *Eugenia uniflora* L. *Talanta*. 2006; 70(2): 286–92. <https://doi.org/10.1016/j.talanta.2006.02.040> PMID:18970765

35. Kim YK, Lee HS, Min SS, Seol GH. Neuroprotective Effect of (-)-Linalool against sodium nitroprusside induced cytotoxicity. *Medicinal Chemistry*. 2015; 5(4): 178–82.
36. Khalil AA, Rahman U, Khan MR, Sahar A, Mehmoodac T, Muneeb Khana. Essential oil eugenol: sources, extraction techniques and nutraceutical perspectives. *RSC Advances*. 2017; 7(52): 32669–81.
37. Pramod K, Ansari SH, Ali J. Eugenol: a natural compound with versatile pharmacological actions. *Natural Product Communications*. 2010; 5(12): 1999–2006. PMID:21299140
38. Wu H, Hendrawinata W, Yu Y, Gao X, Li Y, Bartle J, Grayling P. Effect of Hydro distillation on 1,8-Cineole Extraction from Mallee leaf and the fuel properties of spent biomass. *Industrial & Engineering Chemistry Research*. 2011; 50(19): 11280–7. <https://doi.org/10.1021/ie201092h>
39. Rocha Caldas GF, Silva Oliveira AR, Araujo AV, Lopes Lafayette SS, Albuquerque GS, Silva-Neto JC, et al. Gastroprotective Mechanisms of the Monoterpene 1,8-Cineole (Eucalyptol). *PLoS ONE*. 2015; 10(8): 134558. <https://doi.org/10.1371/journal.pone.0134558> PMID:26244547 PMCID:PMC4526535
40. Benitez PN, Leon MEM, Stashenko EE. Eugenol and methyl eugenol chemotypes of essential oil of species *Ocimum gratissimum* L. and *Ocimum campechianum* from Colombia. *Journal of Chromatographic Science*. 2009; 47(9): 800–3. <https://doi.org/10.1093/chromsci/47.9.800>
41. Ding J, Huang C, Peng Z, Xie Y, Deng S, Nie YZ, Xu TL, Ge WH, Li WG, Li F. Electrophysiological characterization of methyleugenol: a novel agonist of GABA (A) receptors. *ACS Chemical Neuroscience*. 2014; 5(9): 803–11. <https://doi.org/10.1021/cn500022e> PMID:24980777
42. Lopresto CG, Petrillo F, Casazza AA, Aliakbarian B, Perego P, Calabro V. A non-conventional method to extract D-limonene from waste lemon peels and comparison with traditional Soxhlet extraction. *Separation and Purification Technology*. 2014; 137: 13–20.
43. Vale TG, Furtado EC, Santos JG, Viana GS. Central effects of citral, myrcene and limonene, constituents of essential oil chemotypes from *Lippia alba* (Mill). *Phytomedicine*. 2002; 9(8): 709–14. <https://doi.org/10.1078/094471102321621304>
44. Hui-Ping W, Mu Z, Yong L, Weng-Quan L. Extraction and isolation of β -elemene from *Eupatorium adenophorum*. *Journal of Chemical and Pharmaceutical Research*. 2014; 6(5): 161–5.
45. Quentin Li Q, Wang G, Huang F, Banda M, Reed E. Antineoplastic effect of β -elemene on prostate cancer cells and other types of solid tumour cells. *Journal of Pharmacy and Pharmacology*. 2010; 62(8): 1018–27. <https://doi.org/10.1111/j.2042-7158.2010.01135.x> PMID:20663036
46. Jadhav NM, Thergaonkar RS, Deodhar MA. Extraction of essential oil from flowers of *Mesua ferrea* linn. GC-MS analysis and incorporation in cosmetic product. *International Journal of Pharmaceutical Sciences and Research*. 2016; 7(12): 5106–10.
47. Palmer-Young EC, Veit D, Gershenzon J, Schuman MC. The Sesquiterpenes (E)- β -Farnesene and (E)- α -Bergamotene quench ozone but fail to protect the wild Tobacco *nicotiana attenuata* from ozone, UVB, and Drought Stresses. *PLoS ONE*. 2015; 10(6): e0127296. <https://doi.org/10.1371/journal.pone.0127296> PMID:26030663 PMCID:PMC4452144
48. Fan S, Chang J, Zong Y, Hu G, Jia J. GC-MS analysis of the composition of the essential oil from *Dendranthema indicum* Var. *Aromaticum* using three extraction methods and two columns. *Molecules*. 2018; 23(3): 576. <https://doi.org/10.3390/molecules23030576> PMID:29510531 PMCID:PMC6017652
49. Wu X, Xiao F, Zhang Z, Li X, Xu Z. Research on the analgesic effect and mechanism of bornyl acetate in volatile oil from *Amomum villosum*. *Zhong Yao Cai*. 2005; 28(6): 505–7. PMID:16209271
50. Alvi MN, Ahmad S, Rehman K. Short communication preparation of menthol crystals from mint (*Mentha arvensis*). *International Journal of Agriculture and Biology*. 2001; 3(4): 527–8.
51. Galeotti N, Ghelardini C, Cesare Mannelli LD, Mazzanti G, Baghiroli L, Bartolini A. Local anaesthetic activity of (+)- and (\pm)-Menthol. *Planta Medica*. 2001; 67: 174–6. <https://doi.org/10.1055/s-2001-11515> PMID:11301871
52. Nozala MJ, Bernal JL, Jimenez JJ, Gonzalez MJ, Higes M. Extraction of thymol, eucalyptol, menthol, and camphor residues from honey and beeswax: Determination by gas chromatography with flame ionization detection. *Journal of Chromatography A*. 2002; 954(1-2): 207–15. [https://doi.org/10.1016/S0021-9673\(02\)00153-X](https://doi.org/10.1016/S0021-9673(02)00153-X)
53. Zuccarini P. Camphor: risks and benefits of a widely used natural product. *Journal of Applied Sciences and Environmental Management*. 2009; 13(2): 69–74.
54. Tam CU, Yang FQ, Zhang QW, Guan J, Li SP. Optimization and comparison of three methods for extraction of volatile compounds from *Cyperus*

- rotundus evaluated by gas chromatography-mass spectrometry. *Journal of Pharmaceutical and Biomedical Analysis*. 2007; 44(2): 444–9. [https://doi.org/10.1016/S0021-9673\(02\)00153-X](https://doi.org/10.1016/S0021-9673(02)00153-X) PMID:17127024
55. Turkez H, Celik K, Togar B. Effects of copaene, a tricyclic sesquiterpene, on human lymphocytes cells in vitro. *Cytotechnology*. 2013; 66(2014): 597–603. <https://doi.org/10.1007/s10616-013-9611-1> PMID:24287609 PMID:PMC4082788
56. Quispe-Condori S, Foglio MA, Rosa PTV, Meirelesa MAM. Obtaining β -caryophyllene from *Cordia verbenacea* de Candolle by supercritical fluid extraction. *The Journal of Supercritical Fluids*. 2008; 46(1): 27–32.
57. Legault J, Pichette A. Potentiating effect of beta-caryophyllene on anticancer activity of alpha-humulene, isocaryophyllene and paclitaxel. *Journal of Pharmacy and Pharmacology*. 2007; 59(12): 1643–7. <https://doi.org/10.1211/jpp.59.12.0005> PMID:18053325
58. Lekar AV, Borisenko SN, Filonova OV, Vetrova EV, Maksimenko EV, Borisenko NI, Minkin VI. Extraction of caftaric and cichoric acids from *Echinacea purpurea* L. in subcritical water. *Russian Journal of Physical Chemistry B*. 2013; 7(8): 968–75. <https://doi.org/10.1134/S199079311308006X>
59. Kuban-Jankowska A, Sahu KK, Gorska M, Tuszyński JA, Wozniak M. Chicoric acid binds to two sites and decreases the activity of the YopH bacterial virulence factor. *Oncotarget*. 2016; 7(3): 2229–38. <https://doi.org/10.18632/oncotarget.6812> PMID:26735581 PMID:PMC4823031
60. Green CP, Osborne P. Rapid methods for obtaining essential oil from hops. *Journal of the Institute of Brewing*. 1993; 99: 335–9. <https://doi.org/10.1002/j.2050-0416.1993.tb01172.x>
61. Sun Y, Qiao H, Ling Y, Yang S, Rui C, Pelosi P, Yang X. New Analogues of (E)- β -Farnesene with insecticidal activity and binding affinity to aphid odorant binding proteins. *Journal of Agricultural and Food Chemistry*. 2011; 59(6):2456–61. <https://doi.org/10.1021/jf104712c> PMID:21341697
62. Marzouki H, Piras A, Marongiu B, Rosa A, Dessi MA. Extraction and separation of volatile and fixed oils from berries of *Laurus nobilis* L. by supercritical CO₂. *Molecules* 2008; 13: 1702–11. <https://doi.org/10.3390/molecules13081702> PMID:18794780 PMID:PMC6245310
63. Cascaes MM, Skelding Pinheiro Guilhon GM, Aguiar rade EH, Bichara Zoghbi MG, Silva Santos L. Constituents and pharmacological activities of *Myrcia* (Myrtaceae): A review of an aromatic and medicinal group of plants. *International Journal of Molecular Sciences*. 2015; 16: 23881–904. <https://doi.org/10.3390/ijms161023881> PMID:PMC4632730
64. Rohloff J. Monoterpene composition of essential oil from peppermint (*Mentha piperita* L.) with regard to leaf position using solid phase microextraction and Gas Chromatography/Mass Spectrometry Analysis. *Journal of Agricultural and Food Chemistry*. 1999; 47 (9): 3782–6. <https://doi.org/10.1021/jf981310s> PMID:10552722
65. Muftah RS, Shushni Asma Belkheir AM. Antibacterial and antioxidant activities of *Mentha piperita* L. *Arabian Journal of Chemistry*. 2015; 8(3): 322–8. <https://doi.org/10.1016/j.arabjc.2011.01.019>
66. Noge K, Becerra JX. Germacrene D, A common sesquiterpene in the Genus *Bursera* (Burseraceae). *Molecules*. 2009; 14:5289–97. <https://doi.org/10.3390/molecules14125289> PMID:20032892 PMID:PMC6255432
67. Del-Vechio-Vieira G, Vieira de Sousa O, Miranda MA, Senna-Valle L, Coelho Kaplan MA. Analgesic and anti-inflammatory properties of essential oil from *Ageratum fastigiatum*. *Brazilian Archives of Biology and Technology*. 2009; 52(5): 1115–21. <https://doi.org/10.1590/S1516-89132009000500008>
68. SilvErio MS, Del-Vechio-Vieira G, Pinto MAO, Alves MS, Sousa OV. Chemical composition and biological activities of essential oils of *Eremanthus erythropappus* McLeisch (Asteraceae). *Molecules*. 2013; 18: 9785–96. <https://doi.org/10.3390/molecules18089785> PMID:23959191
69. Han GH, Kim SK, Kyung Seok Yoon P, Kang Y, Kim BS, Fu Y. Fermentative production and direct extraction of (-)- α -bisabolol in metabolically engineered *Escherichia coli*. *Microbial Cell Factories*. 2016; 15: 185. <https://doi.org/10.1186/s12934-016-0588-2> PMID:27825357 PMID:PMC5101696
70. Queiroz A, Cajaiba J. A sustainable process for (-)- α -bisabolol extraction from *Eremanthus erythropappus* using supercritical CO₂ and ethanol as co-solvent. *The Journal of Supercritical Fluids*. 2015; 110: 39–46. <https://doi.org/10.1016/j.supflu.2015.12.011>
71. Kamatou GPP, Viljoen AM. A review of the application and pharmacological properties of α -Bisabolol and α -Bisabolol rich oils. *Journal of the*

- American Oil Chemists' Society. 2010; 87(1): 1–7. <https://doi.org/10.1007/s11746-009-1483-3>
72. Naseem MK, Younis A, Khan MA, Ahmad R. Gas chromatography-mass spectrometry of *Murraya exotica* essential oil extracted through different extraction techniques. *The Journal of Animal & Plant Sciences*. 2015; 25(6): 1730–6.
 73. Muley BP, Khadabadi SS, Banarase NB. Phytochemical constituents and pharmacological activities of *Calendula officinalis* L. (Asteraceae): A review. *Tropical Journal of Pharmaceutical Research*. 2009; 8(5): 455–65. <https://doi.org/10.4314/tjpr.v8i5.48090>
 74. Nekoei M, Mohammad hosseini M. Chemical composition of the essential oils and volatiles of *Salvia leriifolia* by three different extraction methods prior to Gas Chromatographic-Mass Spectrometric determination: Comparison of HD with SFME and HS-SPME. *Journal of Essential Oil Bearing Plants*. 2017; 20(2): 410–25. <https://doi.org/10.1080/0972060X.2017.1305918>
 75. Scura MC, Pintoa FGS, Pandini JA, Costa WF, Leite CW, Temponi LG. Antimicrobial and antioxidant activity of essential oil and different plant extracts of *Psidium cattleianum* Sabine. *Brazilian Journal of Biology*. 2016; 76(1): 101–8. <https://doi.org/10.1590/1519-6984.13714PMid:26871744>
 76. Andrade TCB, Lima SG, Freitas RM, Rocha MS, Islam T, Silva TG, et al. Isolation, characterization and evaluation of antimicrobial and cytotoxic activity of estragole, obtained from the essential oil of *Croton zehntneri* (euphorbiaceae). *Anais da Academia Brasileira de Ciencias*. 2015; 87(1): 173–82. <https://doi.org/10.1590/0001-3765201520140111PMid:25789792>
 77. Silva-Alves KS, Ferreira-da-Silva FW, Peixoto-Neves D, Viana-Cardoso KV, Moreira-Junior L, Oquendo MB. Estragole blocks neuronal excitability by direct inhibition of Na⁺ channels. *Brazilian Journal of Medical and Biological Research*. 2013; 46(12): 1056–63. <https://doi.org/10.1590/1414-431X20133191PMid:24345915PMCID:PMC3935278>
 78. Manuale DL, Betti C, Juan C, Yori JM. Synthesis of liquid menthol by hydrogenation of dementholized peppermint oil over Ni catalysts. *Quimica Nova*. 2010; 33(6): 1231–4. <https://doi.org/10.1590/S0100-40422010000600002>
 79. Eccles R, Griffiths DH, Newton CG, Tolley NS. The effects of menthol isomers on nasal sensation of airflow. *Clinical Otolaryngology*. 1988; 13(1): 25–9. <https://doi.org/10.1111/j.1365-2273.1988.tb00277.xPMid:3370851>
 80. Gakuubi MM. Steam distillation extraction and chemical composition of essential oils of *Toddalia asiatica* L. and *Eucalyptus camaldulensis* Dehnh. *Journal of Pharmacognosy and Phytochemistry*. 2016; 5(2): 99–104.
 81. Swamy MK, Sinniah UR. A comprehensive review on the phytochemical constituents and pharmacological activities of *Pogostemon cablin* Benth.: An aromatic medicinal plant of industrial importance. *Molecules*. 2015; 20: 8521–47. <https://doi.org/10.3390/molecules20058521PMid:25985355>
 82. Alshishani AA, Saad B, Semail NF, Salhimi SM, Mohd Talib MK. Salting-out assisted liquid-liquid extraction method coupled to gas chromatography for the simultaneous determination of thujones and pulegone in beverages. *International Journal of Food Properties*. 2017; 20(3): 2776–85. <https://doi.org/10.3390/molecules20058521>
 83. Sousaa DP, Nobrega FFF, Lima MRV, Almeida RN. Pharmacological activity of (R)-(+)-Pulegone, A chemical constituent of essential oils. *Zeitschrift fur Naturforschung C*. 2011; 66: 353–9. <https://doi.org/10.1515/znc-2011-7-806>
 84. Sarwan B, Pare B, Acharya AD, Jonnalagadda SB. Mineralization and toxicity reduction of textile dye neutral red in aqueous phase using Biocphotocatalysis. *Journal of Photochemistry and Photobiology B: Biology*. 2012; 166: 48–55. <https://doi.org/10.1016/j.jphotobiol.2012.07.006PMid:22964463>
 85. Gosavi VD, Sharma S. A general review on various treatment methods for textile wastewater. *JECET: Journal of Environmental Science, Computer Science and Engineering and Technology*. 2013; 3: 29–39.
 86. Solanki M, Suresh S, Das SN, Shukla K. Treatment of real textile wastewater using coagulation technology. *International Journal of ChemTech Research*. 2013; 5: 610–5.
 87. Yin CY. Emerging usage of plant-based coagulants for water and wastewater treatment. *Process Biochemistry*. 2010; 45: 1437–44. <https://doi.org/10.1016/j.procbio.2010.05.030>
 88. Sciban M, Klasnja M, Antov M, Skrbic B. Removal of water turbidity by natural coagulants obtained from chestnut and acorn. *Bioresource Technology*. 2010; 100: 6639–43. <https://doi.org/10.1016/j.biortech.2009.06.047PMid:19604691>
 89. Gupta A, Karjekar M, Nair J. Biosorption of copper using mucilaginous seeds of *Ocimum basilicum*. *Acta Biologica Indica*. 2010; 1: 113–9.

90. Melo JS, souza SF. Removal of chromium by mucilaginous seeds off *Ocimum basilicum*. *Bioresource Technology*. 2004; 92: 151–5. <https://doi.org/10.1016/j.biortech.2003.08.015> PMID:14693447
91. Sahay R, patra DD. Identification and performance of sodicity tolerant phosphate solubilizing bacteria isolates on *Ocimum basilicum* in sodic soil. *Ecological Engineering*. 2014; 71: 639–43. <https://doi.org/10.1016/j.ecoleng.2014.08.007>
92. Klimankova E, Holadova K, Hajslova J, Cajka K, Poustka J, Koudela M. Aroma profiles of five basil (*Ocimum basilicum* L.) cultivars grown under conventional and organic conditions. *Food Chemistry*. 2001; 107: 464–72. <https://doi.org/10.1016/j.foodchem.2007.07.062>
93. Werker E, Putievsky E, Ravid U, Dudai N, Katzir I. Glandular hairs and essential oil in developing leaves of *Ocimum basilicum* L. (Lamiaceae). *Annals of Botany*. 2001; 71: 43–50. <https://doi.org/10.1006/anbo.1993.1005>
94. Govindarajan M, Sivakumar R, Rajeswary M, Yogalakshmi K. Chemical composition and larvicidal activity of essential oil from *Ocimum basilicum* (L.) against *Culex tritaeniorhynchus*, *Aedes albopictus* and *Anopheles subpictus* (Diptera: Culicidae). *Experimental Parasitology*. 2013; 134: 7–11. <https://doi.org/10.1016/j.exppara.2013.01.018> PMID:23391742
95. Zeggwagh N, Sulpice A, Eddouks TM. Anti-hyperglycaemic and hypolipidemic effects of *Ocimum basilicum* aqueous extract in diabetic rats. *American Journal of Pharmacology and Toxicology*. 2007; 2: 123–9. <https://doi.org/10.3844/ajtpsp.2007.123.129>
96. Nyugen PM, Kwee EM, Niemeyer ED. Potassium rate alters the antioxidant capacity and phenolic concentration of basil (*Ocimum basilicum* L.) leaves. *Food Chemistry*. 2010; 123: 1235–41. <https://doi.org/10.1016/j.foodchem.2010.05.092>
97. Hossain MA, Kabir MJ, Salehuddin SM, Das AK, Singha SK, Alam MK, Rahman A. Antibacterial properties of essential oils and methanol extracts of sweet basil *Ocimum basilicum* occurring in Bangladesh. *Pharmaceutical Biology*. 2010; 5: 504–11. <https://doi.org/10.3109/13880200903190977> PMID:20645791
98. Kim HJ, Chen F, Wang X, Rajapakse NC. Effect of chitosan on the biological properties of sweet basil (*Ocimum basilicum* L.). *Journal of Agricultural and Food Chemistry*. 2005; 53: 3696–701. <https://doi.org/10.1021/jf0480804> PMID:15853422
99. Ozcan M, Chalchat JC. Essential oil of *Ocimum basilicum* L. and *Ocimum minimum* L, in turkey. *Journal of Food Science*. 2005; 20: 223–8.
100. Sethuraman J, Nehru H, Shanmugam K, Balakrishnanan P. Evaluation of potent phytochemicals and antidiabetic activity of *Ficus racemosa* L. *World Journal of Pharmaceutical Research*. 2017; 6(15): 909–20.
101. Ramalingam PS, Sagayaraj M, Ravichandiran P, Balakrishnanan P, Nagarasan S, Shanmugam K. Lipid peroxidation and anti-obesity activity of *Nigella sativa* seeds. *World Journal of Pharmaceutical Research*. 2017; 6(10): 882–92.
102. Amrani S, Harnafi H, Gadi D, Mechfi H, Legssyer A, Ariz M, Martin-Nizard F, Bosca L.: Vasorelaxant antiplatelet aggregation effects of aqueous *Ocimum basilicum* extract. *Journal of Ethnopharmacology*. 2009; 125: 157–62. <https://doi.org/10.1016/j.jep.2009.05.043> PMID:19505553
103. Raina P, Deepak M, Chandrasekaran CV, Agarwal A, Wagh N, Kaul-Ghanekar R. Comparative analysis of anti-inflammatory activity of aqueous and methanolic extracts of *Ocimum basilicum* RAW 264.7, SW1353 and human primary chondrocytes. *Journal of Herbal Medicine*. 2016; 6(1): 28–36. <https://doi.org/10.1016/j.hermed.2016.01.002>
104. Pandey V, Patel A, Patra DD. Integrated nutrient regimes ameliorate crop productivity, nutritive value, antioxidant activity and volatiles in basil (*Ocimum basilicum* L.). *Industrial Crops and Products*. 2016; 87: 124–31. <https://doi.org/10.1016/j.indcrop.2016.04.035>
105. Flanigan PM, Niemeyer ED. Effect of cultivar on phenolic levels, anthocyanin composition, and antioxidant properties in purple basil (*Ocimum basilicum* L.). *Food Chemistry*. 2014; 164: 518–26. <https://doi.org/10.1016/j.foodchem.2014.05.061> PMID:24996365
106. Okazaki K, Nakayama S, Kawazoe K, Takaishi Y. Anti-aggregant effects on human platelets of culinary herbs. *Phytotherapy Research*. 2011; 12: 603–5. [https://doi.org/10.1002/\(SICI\)1099-1573\(199812\)12:8<603::AID-PTR372>3.0.CO;2-G](https://doi.org/10.1002/(SICI)1099-1573(199812)12:8<603::AID-PTR372>3.0.CO;2-G)
107. Aarthi N, Murugan K. Larvicidal and repellent activity of *Vetiveria zizanioides* L, *Ocimum basilicum* L and the microbial pesticide spinosad against malarial vector, *Anopheles stephensi* Liston (Insecta: Diptera: Culicidae). *Journal of Biopesticides*. 2010; 3: 199–204.
108. Davies J. Inactivation of antibiotics and the dissemination of resistance genes. *Science*. 1994; 64: 375–82. <https://doi.org/10.1126/science.8153624>

109. Faleiro JR, El-Saad MA, Al-Abbad AH. Pheromone trap density to mass trap *Rhynchophorus ferrugineus* (Coleoptera: Curculionidae/Rhynchophoridae/Dryophthoridae) in date plantations of Saudi Arabia. *International Journal of Tropical Insect Science*. 2011; 31: 75–7. <https://doi.org/10.1017/S1742758411000099>
110. Basilico MZ, Basilico JC. Inhibitory effects of some spice essential oils on *Aspergillus ochraceus* NRRL 3174 growth and ochratoxin. A production. *Letters in Applied Microbiology*. 1999; 29: 238–41. <https://doi.org/10.1046/j.1365-2672.1999.00621.x> PMID:10583751
111. Bais HP, Walker TS, Schweizer HP, Vivanco JM. Root specific elicitation and antimicrobial activity of rosmarinic acid in hairy root cultures of sweet basil (*Ocimum basilicum* L.). *Plant Physiology and Biochemistry*. 2002; 40: 983–95. [https://doi.org/10.1016/S0981-9428\(02\)01460-2](https://doi.org/10.1016/S0981-9428(02)01460-2)
112. Kumar A, Shukla R, Singh P, Prakash B, Dubey NK. Chemical composition of *Ocimum basilicum* L. essential oil and its efficacy as a preservative against fungal and aflatoxin contamination of dry fruits. *International Journal of Food Science and Technology*. 2011; 46: 1840–6.
113. Oxenham SK, Svoboda KP, Walters DR. Antifungal activity of the essential oil of basil (*Ocimum basilicum*). *Journal of Phytopathology*. 2005; 153: 174–80. <https://doi.org/10.1111/j.1439-0434.2005.00952.x>
114. Marinava EM, Ynishlieva NV. Antioxidative activity of extracts from selected species of the family of Lamiaceae in sunflower oil. *Food Chemistry*. 1997; 58: 245. [https://doi.org/10.1016/S0308-8146\(96\)00223-3](https://doi.org/10.1016/S0308-8146(96)00223-3)
115. Nauen R. Insecticide resistance in disease vectors of public health importance. *Pest Management Science*. 2007; 63: 628–33. <https://doi.org/10.1002/ps.1406> PMID:17533649
116. Kathirvel P, Ravi S. Chemical composition of the essential oil from basil (*Ocimum basilicum* Linn.) and its in vitro cytotoxicity against HeLa and HEP-2 human cancer cell lines and NIH 3T3 mouse embryonic fibroblasts. *Natural Product Research*. 2012; 26: 1112–8. <https://doi.org/10.1080/14786419.2010.545357> PMID:21939371
117. Hussain AI, Anwar F, Sherazi STH, Przybylski R. Chemical composition, antioxidant and antimicrobial activities of basil (*Ocimum basilicum*) essential oils depend on seasonal variations. *Food Chemistry*. 2008; 108: 986–95. <https://doi.org/10.1016/j.foodchem.2007.12.010> PMID:26065762
118. Taie, NA, Salama, ZA, Radwan S. Potential activity of basil plants as a source of antioxidants and anticancer agents as affected by organic and Bio-organic fertilization. *Notulae Botanicae Horti Agrobotanici Cluj-Napoca*. 2010; 38: 119–27.
119. Tran T. Basil Diseases. Cornell University Department of Plant Pathology and Plant-Microbe Biology. 2011. Available from: <http://plantclinic.cornell.edu/factsheets/basil diseases.pdf>.
120. CABI Crop Protection Compendium. *Ocimum basilicum* datasheet. 2008. Available from: <http://www.cabi.org/cpc/datasheet/36858>.
121. Gamliel A, Yarden O. Diversification of diseases affecting herb crops in Israel accompanies the increase in herb crop production. *Phytoparasitica*. 1998; 26: 53. <https://doi.org/10.1007/BF02981266>
122. Araldi RP, Assaf SMR, Carvalho RF, Caldas MA, Carvalho R, Souza JM, Magnelli RF, Modolo GD, Roperto FP, Stocco C and Becak W. Papillomaviruses: a systematic review. *Genetics and Molecular Biology*. 2017; 40(1): 1–21. <https://doi.org/10.1590/1678-4685-gmb-2016-0128> PMID:28212457 PMCID:PMC5409773
121. Gamliel A, Yarden O. Diversification of diseases affecting herb crops in Israel accompanies the increase in herb crop production. *Phytoparasitica*. 1998; 26: 53. <https://doi.org/10.1007/BF02981266>
122. Araldi RP, Assaf SMR, Carvalho RF, Caldas MA, Carvalho R, Souza JM, Magnelli RF, Modolo GD, Roperto FP, Stocco C and Becak W. Papillomaviruses: a systematic review. *Genetics and Molecular Biology*. 2017; 40(1): 1–21. <https://doi.org/10.1590/1678-4685-gmb-2016-0128>. PMID:28212457 PMCID:PMC5409773