Antioxidant Activities And Total Phenolic Content Of Malaysian Herbs As Active Packaging Film In Meat Patties

Wan Amnin WanYahaya^{1,b)}, Alyaa Syahierra Ghazali^{1,c)}, Khadijah Husna Abd. Hamid^{1,d)}, Neenasha Bebe Mohd Nor^{1,e)}, Nurul Aini Mohd Azman^{1,a)}

¹Faculty of Chemical & Natural Resources Engineering, Universiti Malaysia Pahang, 26300 Gambang, Pahang, Malaysia

> ^{a)}Corresponding author: ainiazman@ump.edu.my ^{b)}amninyahaya@gmail.com ^{c)} alyaasyahierra@gmail.com ^{d)}khadijah.husnaa@gmail.com ^{e)} neenashabebe@gmail.com

Abstract. Active packaging incorporated with natural extracts is a promising technology to extend shelf life of perishable food. Therefore, this study aims to produce the bioactive edible film from semi refined carrageenan (SRC) plasticized with glycerol (G) and incorporated with natural extract to improve lipid oxidative stability and storage quality of meat patties. Evaluation of five Malaysia herbs (*Cosmos caudate;* CC, *Piper sarmentosum;* PS, *Persicaria minor;* PM, *Centella asiatica;* CA and *Syzygium polyanthum;* SP) were analysed for their phenolic content and antioxidant activities and total phenolic content using Folin-Ciocaltea method, and the antioxidant activities of plants were determined using 2,2-diphenyl-1-picrylhydrazyl (DPPH) method, trolox equivalent antioxidant capacity (TEAC) and oxygen radical absorbance capacity (ORAC). PM extract demonstrated highest total phenolic content (1.629 mgGAE/Lsample) and antioxidant activities value in TEAC (27.166 mg TE/ L sample), DPPH (719.89 mg eq. trolox/L sample) and ORAC (5.81 mg TE/ L sample). Thus, PM extract was selected to be incorporated into active packaging film with concentration of 0.4,1.5 and 2.0% in 2% SRC and 0.9% G (w/w) film formulation. The developed films were wrapped with meat patties under refrigerated condition (4±2°C) and monitored for 14 days storage period. Film wrapped with 2% (w/w) PM exhibited significantly lower lipid deterioration rate analyzed by TBARS method (p<0.05) and low changes of % metmyoglobin value that indicates the minimum development of brown color (p<0.05). Hence, the active packaging edible films incorporating PM demonstrated potential as a packaging material to improve food safety and quality of the meat.

INTRODUCTION

Food spoilage is usually occur by lipid oxidation. Generally, deterioration of foodstuff easily occur with high lipid content of food especially those with unsaturation high grade. The oxidation of lipids in foodstuffs results in severe problem that cause development of off-flavors, rancidity, modify texture and colour, and leads to the growth of microorganisms and vitamin losses [1]. According to previous literature, the formation of toxic aldehydes and the loss of nutritional quality because of polyunsaturated fatty acid (PUFA) degradation cause by lipid oxidation. To minimize lipid oxidation, many strategies have been enforced [2]. For example, the direct addition of antioxidants to foods or the design of a suitable packaging technology such as vacuum or modified-atmosphere packaging combined with high-barrier packaging materials [3]. Commonly, the packaging system are introduced by conventional packaging which it consist of synthetic antioxidants such as Butylated Hydroxyanisole (BHA) and Butylated Hydroxytoluene (BHT) that can cause potential health risks cause by such compound. The combination of natural preservatives and biodegradable plastic into one food packaging formulation was a promising approach to extending product shelf life [6].

- 6. Suppakul P., Miltz J., Sonneveld K., Bigger S. W. (2003). Active packaging technologies with an emphasis on antimicrobial packaging and its applications. *J. Food. Sci.* 68 408–420.
- Mayachiew, P., Devahastin, S., Mackey, B. M., & Niranjan, K. (2010). Effects of drying methods and conditions on antimicrobial activity of edible chitosan films enriched with galangal extract. *Food Research International*, 43(1), 125–132.
- dos Santos Caetano, K., Almeida Lopes, N., Haas Costa, T. M., Brandelli, A., Rodrigues, E., Hickmann Flôres, S., & Cladera-Olivera, F. (2018). Characterization of active biodegradable films based on cassava starch and natural compounds. *Food Packaging and Shelf Life*, *16* (2017), 138–147.
- 9. Kam, W. J., Mirhosseini, H., Abas, F., Hussain, N., Hedayatnia, S., & Chong, H. F. (2018). Antioxidant activity enhancement of biodegradable film as active packaging utilizing crude extract from durian leaf waste. *Food Control*, *90*, 66–72.
- 10. Breda, C. A., Morgado, D. L., Benedito, O., Assis, G., Cristina, M., & Duarte, T. (2017). Processing and Characterization of Chitosan Films with Incorporation of Ethanolic Extract from "Pequi" Peels, 1049–1056.
- 11. Messina, C. M., Bono, G., Renda, G., La Barbera, L., & Santulli, A. (2015). Effect of natural antioxidants and modified atmosphere packaging in preventing lipid oxidation and increasing the shelf-life of common dolphinfish (Coryphaena hippurus) fillets. *LWT Food Science and Technology*, 62(1), 271–277.
- Ganiari, S., Choulitoudi, E., & Oreopoulou, V. (2017). Edible and active films and coatings as carriers of Toohey, E. S., Kerr, M. J., Ven, R. Van De, & Hopkins, D. L. (2011). The effect of a kiwi fruit based solution on meat traits in beef m. semimembranosus (topside). *MESC*, 88(3), 468–471.
- 13. Abreu, D. A. P. De, Losada, P. P., Maroto, J., & Cruz, J. M. (2011). Natural antioxidant active packaging fi lm and its effect on lipid damage in frozen blue shark (Prionace glauca). *Innovative Food Science and Emerging Technologies*, *12*(1), 50–55.
- 14. Seydim, A. C., & Sarikus, G. (2006). Antimicrobial activity of whey protein based edible films incorporated with oregano, rosemary and garlic essential oils, *39*, 639–644.
- 15. Vimala S, Ilham MA, Rashih AA, Rohana S, Juliza M., (2006). Antioxidant and skin whitening standardized extracts: Cosmeceutical and neutraceutical products development and commercialization in FRIM. In: Highlights of FRIM's IRPA Projects 2005: Identifying Potential Commercial Collaborations, edited by Zanariah, N. Forest Research Institute Malaysia.
- 16. Shi-Hui Cheng, Mohd Yusof Barakatun-Nisak, Joseph Anthony, and Amin Ismail, (2015). Potential medicinal benefits of Cosmos caudatus (Ulam Raja): A scoping review
- 17. Seyedreihani S. F., Thuan-Chew Tan, Abbas F. M. Alkarkhi & Azhar Mat Easa, (2016). Total phenolic content and antioxidant activity of *Ulam raja* (*Cosmos caudatus*) and quantification of its selected marker compounds: Effect of extraction.
- 18. Zaidan M. R., Noor R. A., Badrul A. R., Adlin A., Norazah A., Zakiah I., (2005). In vitro screening of five local medicinal plants for antibacterial activity using disc diffusion method. *Tropical Biomedicine* 22, 165–170.
- 19. Ridtitid, W., Ruangsang, P., Reanomongkol, W., Malinee, W., 2007. Studies of the anti-inflammatory and antipyretic activities of the methanolic extract of *Piper sarmentosum* Roxb. leaves in rats. Songklanakarin *Journal of Science Technology* 29, 1520–1527.
- Sireeratawong, S., Vannasiri, S., Sritiwong, S., Itharat, A., Jaijoy, K., (2010). Anti-inflammatory, antinociceptive and antipyretic effects of the ethanol extract from root of *Piper sarmentosum* Roxb. *Journal of Medical Association of Thailand* 93, S1–6.
- 21. Zakaria Z. A., Patahuddin H., Mohamad A. S., Israf D. A., Sulaiman, M. R., 2010. In vivo anti-nociceptive and anti-inflammatory activities of the aqueous extract of the leaves of *Piper sarmentosum*. Journal of Ethnopharmacology 128, 42–48
- 22. Chanwitheesuk, A., Teerawutgulrag, A., Rakariyatham, N., 2005. Screening of antioxidant activity and antioxidant compounds of some edible plants of Thailand. Food Chemistry 92, 491–497.
- 23. Ugusman, A., Zakaria, Z., Hui, C. K., Nordin, N. A., 2011. *Piper sarmentosum* inhibits ICAM-1 and Nox4 gene expression in oxidative stress-induced human umbilical vein endothelial cells. BMC Complementary and Alternative Medicine 11, 31–36.
- 24. Sharifah Farhana Syed Ab Rahman, Kamaruzaman Sijam and Dzolkhifli Omar, 2014. Chemical composition of *Piper sarmentosum* extracts and antibacterial activity against the plant pathogenic bacteria *Pseudomonas fuscovaginae* and *Xanthomonas oryzae pv*. Oryzae.
- 25. Nanasombat, S. and Teckchuen, N., 2009. Antimicrobial, antioxidant and anticancer activities of Thai local vegetables. Journal of Medicinal Plants Research 3(5): 443-449

- 26. Siró, É. Fenyvesi, L. Szente, B. De Meulenaer, F. Devlieghere, J. Orgoványi, *et al.* Release of alpha-tocopherol from antioxidative low-density polyethylene film into fatty food simulant: Influence of complexation in beta-cyclodextrin *Food Additives & Contaminants*, 23 (8) (2006), pp. 845-853.
- 27. Vimala, S. & Mohd Ilham, A. (2011). Antioxidant Evaluation in Malaysian Medicinal Plant: *Persicaria* minor (Huds.) Leaf, Science Journal of Medicine & Clinical Trials 5(1): 32–38.
- 28. Murray, Joseph E. Pizzorno, Jr., Michael T., 2012. Textbook of natural medicine (4th edition). Edinburgh: Churchill Livingstone. p. 650.
- 29. Hidayati, M. D., Ersam, T., Shimizu, K., & Fatmawati, S. (2017). Antioxidant Activity of Syzygium polynthum Extracts, *17*(1), 49–53.
- 30. Azman A. Nurul, Husni Shafik, Almajano P. Maria, G. G. M. (2013). Solvent Effect on Antioxidant Activity and Total Phenolic Content of Betula alba and Convolvulus arvensis, 7(5), 351–356.
- 31. Farhan, A., and Hani, N. M., 2017, Characterization of edible packaging films based on semi-refined kappacarrageenan plasticized with glycerol and sorbitol, *Food Hydrocolloid*, 64, 48–58.
- 32. Grau, A., Guardiola, F., Boatella, J., Barroeta, A. and Codony, R. (2000). Measurement of 2-thiobarbituric acid values in dark chicken meat through derivative spectrophotometry: Influence of various parameters, *Journal of Agricultural and Food Chemistry*, 48(4): 1155 1159
- Xu, Z., Tang, M., Li, Y., Liu, F., Li, X. and Dai, R. (2010). Antioxidant properties of Du-zhong (*Eucommia ulmoides Oliv.*) extracts and their effects on color stability and lipid oxidation of raw pork patties, *Journal of Agricultural and Food Chemistry*, 58(12): 7289 7296.
- 34. Puziah Hashim, Hamidah Sidek, Mohd Helme M. Helan, Aidawati Sabery, Uma Devi Palanisamy and Mohd Ilham, 2011. Triterpene Composition and Bioactivities of *Centella asiatica*.
- 35. Reihani, S. F. S. and Azhar, M. E., 2012. Antioxidant activity and total phenolic content in aqueous extracts of selected traditional Malay salads (Ulam), *International Food Research Journal* 19(4): 1439-1444.
- 36. Beatriz C. B. S. Mello & Miriam D. Hubinger, 2012. Antioxidant activity and polyphenol contents in Brazilian green propolis extracts prepared with the use of ethanol and water as solvents in different pH values. International *Journal of Food Science and Technology* 47, 2510–2518
- Miguel, M.C., Nunes, S., Dandlen, S.A., Cavaco, A.M. & Antunes, M.D., 2010. Phenols and antioxidant activity of hydro-alcoholic extracts of propolis from Algarve, South of Portugal. Food and Chemical Toxicology, 48, 3418–3423
- Thaipong, K., Boonprakob, U., Crosby, K., Cisneros-Zevallos, L. & Byrne, D.H., 2006. Comparison of ABTS, DPPH, FRAP, and ORAC assays for estimating antioxidant activity from guava fruit extracts. Journal of Food Composition and Analysis, 19, 669–675.
- 39. Gardner, P.T., White, T.A.C., Mcphail, D.B. & Duthie, G.G., 2000. The relative contributions of vitamin C, carotenoids and phenolics to the antioxidant potential of fruit juices. Food Chemistry, 65, 471–474.
- 40. Gil, M.I., Thomas-Barberan, F.A., Hess-Pierce, B. & Kader, A.A., 2002. Antioxidant capacities, phenolic compounds, carotenoids, and vitamin C contents of nectarine, peach and plum cultivars from California. Journal of Agricultural and Food Chemistry, 50, 4976–4982.
- 41. Distantina, S., Rochmadi, Fahrurrozi, M., and Wiratni, 2013, Preparation and characterization of glutaraldehydecrosslinked kappa carrageenan hydrogel, *Eng. J.*, 17 (3), 57–66.
- 42. Rhim, J. W., and Wang, L. F., 2014, Preparation and characterization of carrageenan-based nanocomposite films reinforced with clay mineral and silver nanoparticles, *Appl. Clay Sci.*, 97–98, 174–181.
- Kaya, M., Khadem, S., Cakmak, Y. S., Mujtaba, M., Ilk, S., Akyuz, L., ... Deligöz, E. (2018). Antioxidative and antimicrobial edible chitosan films blended with stem, leaf and seed extracts of Pistacia terebinthus for active food packaging. *RSC Advances*, 8(8), 3941–3950.
- 44. Kongjao, S., Damronglerd, S., & Hunsom, M. (2010). Purification of crude glycerol derived 596 from waste used-oil methyl ester plant. Korean Journal of Chemical Engineering, 27, 944–949. A
- 45. Pereira, L., Amado, A. M., Critchley, A. T., van de Velde, F., & Ribeiro-Claro, P. J. A. (2009). 623 Identification of selected seaweed polysaccharides (phycocolloids) by vibrational 624 spectroscopy (FTIR-ATR and FT-Raman). Food Hydrocolloids, 23, 1903–1909
- 46. Lee, H. E., Kim, D. H., Park, S. J., Kim, J. M., Lee, Y. W., Jung, J. M., Lee, C. H., Hong, J. G., Liu, X., Cai, M. and Park, K. J. (2012). Neuroprotective effect of sinapic acid in a mouse model of amyloid β1-42 protein-induced alzheimer's disease, *Pharmacology Biochemistry and Behaviour*, 103: 260–266.
- 47. Azman, N. A. M., Gallego, M. G., Segovia, F., Abdullah, S., Sharani, S. M., Pablos, M. P. A. (2016). Study of the Properties of Bearberry Leaf Extract as a Natural Antioxidant in Model Foods, *Antioxidants*, 5, 11.

- 48. Lin, Y., Huang, M., Zhou, G., Zou, Y. and Xu, X. (2011). Prooxidant effects of the combination of green tea extract and sodium nitrite for accelerating lipolysis and lipid oxidation in pepperoni during storage, *Journal of Food Science*, 76(5): 694–700.
- 49. Hamid, K. H. A., Azman, N., Sharaani, S., Zain, N., Ahmad, N., Sulaiman, A. Z., Chik, S. S. T. Ishak, W. F. W., Pablos, M. P. A. (2017). *Alchemilla vulgaris* and *Filipendula ulmaria* extracts as potential natural preservatives in beef patties, *Malaysian Journal of Analytical Sciences*, 21, 986–995.
- 50. Gómez, F., Sánchez, S., Iradi, M., Azman, N., & Almajano, M. (2014). Avocado Seeds: Extraction Optimization and Possible Use as Antioxidant in Food. *Antioxidants*, *3*(2), 439–454.
- 51. Azman, A., Gallego, M., Julià, L., Fajari, L., & Almajano, M. P. (2015). The effect of convolvulus arvensis dried extract as a potential antioxidant in food models. *Antioxidants*, 4(1), 170–184.
- 52. Azman, A., Gallego, M., Julià, L., Fajari, L., & Almajano, M. P. (2015). The effect of convolvulus arvensis dried extract as a potential antioxidant in food models. *Antioxidants*, 4(1), 170–184.
- 53. Skowyra, M.; Falguera, V.; Gallego, G.; Peiró, S.; Almajano, M.P. (2014). Antioxidant properties of aqueous and ethanolic extracts of Tara (*Caesalpinia spinosa*) pods in vitro and in model food emulsions, *J. Sci. Food Agric.*, 94, 911–918.
- 54. Camo, J., Beltrán, J. A., & Roncalés, P. (2008). Extension of the display life of lamb with an antioxidant active packaging, *Meat Science*, 80(4), 1086-1091.
- Contini, C., Álvarez, R., O'Sullivan, M., Dowling, D. P., Gargan, S. Ó., & Monahan, F. J. (2014). Effect of an active packaging with citrus extract on lipid oxidation and sensory quality of cooked turkey meat, *Meat Science*, 96(3), 1171-1176.