

GREEN SYNTHESIS OF SILVER AND COPPER
NANOPARTICLES USING HYDROXYETHYL
CELLULOSE AND ITS ANTIBACTERIAL ACTIVITY

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We hereby declare that we have checked this thesis and in our opinion, this thesis is adequate in terms of scope and quality for the award of the degree of Master of Science in Advanced Materials

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STUDENT'S DECLARATION

I hereby declare that the work in this thesis is based on my original work except for quotations and citations which have been duly acknowledged. I also declare that this thesis has not been previously or concurrently submitted for any other degree at Universiti Malaysia Pahang or any other institution.

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ABSTRAK

Disertasi ini memfokuskan pada pengenalan satu kaedah baharu mensintesis zarah-zarah nano argentum dan kuprum (AgNPs dan CuNPs) menggunakan hidroksietil selulosa (HEC). Kajian ini dilakukan melalui kaedah lestari untuk mengelakkan penggunaan bahan kimia yang berbahaya dan juga untuk menjimatkan kos pengeluaran. Disertasi ini juga menerangkan keadaan-keadaan optimum untuk sintesis zarah-zarah nano dengan menganalisis pelbagai parameter, seperti isipadu pelopor-pelopor argentum nitrat (AgNO_3) dan kuprum nitrat ($\text{Cu(NO}_3)_2$), kepekatan HEC, masa tindak balas serta suhu. Pada $100\text{ }^\circ\text{C}$, tindak balas AgNPs selesai dalam 30 minit manakala CuNPs pula dalam 3 minit. Kehadiran kedua-dua AgNPs dan CuNPs dibuktikan oleh spektroskopi ultraungu-tampak (UV-Vis), yang menunjukkan puncak resonans plasmon permukaan (SPR) masing-masing pada 410 - 430 nm dan 550 - 600 nm. Kehadiran zarah-zarah nano dan struktur kristal disahkan oleh difraktometri sinar-X (XRD) dan sinar-X sebaran tenaga (EDX). Pencirian struktur dan morfologi AgNPs dan CuNPs dilakukan menggunakan mikroskop elektron penghantaran (TEM) dan mikroskop elektron pengimbasan pancaran medan (FESEM). Aktiviti antibakteria zarah-zarah nano juga diuji melalui kaedah peresapan agar-agar, kepekatan perencatan minimum (MIC) dan kepekatan bakterisidal minimum (MBC). Kajian ini juga menguji kesan AgNPs, CuNPs dan campuran Ag-CuNPs terhadap bakteria Gram-positif dan Gram-negatif. Zarah-zarah nano menunjukkan aktiviti antibakteria yang baik terhadap *Bacillus subtilis* (*B. subtilis*), *Escherichia coli* (*E. coli*), *Pseudomonas aeruginosa* (*P. aeruginosa*), *Enterococcus faecalis* (*E. faecalis*) dan *Staphylococcus aureus* (*S. aureus*). Zon perencatan bagi zarah-zarah nano ini berubah mengikut jenis bakteria. Zon perencatan yang lebih besar diperhatikan pada bakteria Gram-negatif (*E. coli*). Saiz zon perencatan tersebut ialah 19 mm untuk AgNPs, manakala 16 mm untuk CuNPs dan Ag-CuNPs. Oleh itu, disertasi ini menyimpulkan bahawa penemuan kaedah lestari yang sesuai untuk sintesis AgNPs dan CuNPs mempunyai kesan perencatan yang baik terhadap bakteria, justeru terdapat pelbagai aplikasi yang berpotensi untuk zarah-zarah nano ini.

ABSTRACT

This dissertation is mainly focused about the introduction of a new method of synthesizing silver and copper nanoparticles (AgNPs and CuNPs) using hydroxyethyl cellulose (HEC). The study was done via green chemistry method to avoid the usage of some hazardous chemicals and also to save the cost of production. This thesis describes the optimal conditions for the synthesis of nanoparticles by analysing various parameters, such as the volume of the precursors silver nitrate (AgNO_3) and copper nitrate ($\text{Cu}(\text{NO}_3)_2$), the concentration of HEC, reaction times and temperature. At 100 °C, the AgNPs reaction went to completion in 30 min while the CuNPs reaction about 3 min. The presence of both AgNPs and CuNPs were assured by ultraviolet visible spectroscopy (UV-Vis), which showed surface plasmon resonance (SPR) peaks at 410 - 430 nm and 550 - 600 nm respectively. The presence of the nanoparticles and the crystal structure were confirmed by X-ray diffractometry (XRD) and energy-dispersive X-ray (EDX). The structural and morphological characterisations of the AgNPs and CuNPs were performed using a transmission electron microscope (TEM) and field emission scanning electron microscope (FESEM). The antibacterial activities of the nanoparticles were also studied via the agar-well diffusion method, minimum inhibition concentration (MIC) and minimum bactericidal concentration (MBC). The research also tested the effects of AgNPs, CuNPs and Ag-CuNP mixtures on Gram-positive and Gram-negative bacteria. The nanoparticles showed good antibacterial activity against *Bacillus subtilis* (*B. subtilis*), *Escherichia coli* (*E. coli*), *Pseudomonas aeruginosa* (*P. aeruginosa*), *Enterococcus faecalis* (*E. faecalis*) and *Staphylococcus aureus* (*S. aureus*). The inhibition zones for these nanoparticles varied based on the type of bacteria. The larger inhibition zone was observed on Gram-negative bacteria (*E. coli*). The sizes of the said inhibition zones were 19 mm for AgNPs, 16 mm for CuNPs and Ag-CuNPs. Thus, it can be concluded that the invention of a feasible green method for the synthesis of AgNPs and CuNPs had a good inhibitory effect on the bacteria and hence, there are various potential applications for these nanoparticles.

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LIST OF SYMBOLS

+	Add
α	Alpha
Z	Atomic number
β	Beta
Cu^{2+}	Copper (II) ions
Cu^0	Copper particles
$^{\circ}$	Degree
$^{\circ}\text{C}$	Degree Celcius
\$	Dollar
g	Gram
g/mol	Gram per mol
h	Hour
keV	Kilo electronvolt
Kg	Kilogram
kV	Kilovolts
L	Litre
μL	Microliter
mA	Milliampere
mL	Millilitre
mm	Millimetre
min	Minutes
M	Molarity
M_w	Molecular weight
nm	Nanometer
n	Number of mol
%	Percentage
pH	Potential of hydrogen
M_r	Relative molar mass
Ag^+	Silver ion
Ag^0	Silver particles
θ	Theta
V	Volume
λ	Wavelength
Wt%	Weight percentage

LIST OF ABBREVIATIONS

Al	Aluminium
ATCC	American Type Culture Collection
C ₆ H ₈ O ₆	Ascorbic acid
(C ₁₆ H ₁₈ N ₂ O ₄ SNa)	Benzylpenicillin sodium
Co	Cobalt
CFU	Colony forming unit
Cu	Copper
Cu(NO ₃) ₂	Copper (II) nitrate
CuNPs	Copper nanoparticles
EDX	Energy dispersive X-ray
E-Test	Epsilometer test
Fcc	Face-centered cubic
Au	Gold
AuNPs	Gold nanoparticles
HTAB	Hexadecyltrimethylammonium bromide
HRTEM	High-resonance transmission electron microscopy
H ₂ O ₂	Hydrogen peroxide
HEC	Hydroxyethyl cellulose
OH	Hydroxy groups
HP	Hyperbranched polyurethane
FESEM	Field emission scanning electron microscopy
Fe	Iron
FeNPs	Iron nanoparticles
Fe ₂ O ₃	Iron oxide
Mn	Manganese
MBC	Minimum bactericidal concentration
MIC	Minimum inhibition concentration
MRI	Magnetic resonance imaging
MHA	Mueller Hinton agar
MHB	Mueller Hinton broth
NPs	Nanoparticles
Ni	Nickel
PAN	Polyacrylonitrile
PTFE	Polytetrafluoroethylene
PVA	Polyvinyl alcohol
PVP	Polyvinylpyrrolidone
rpm	Revolutions per minute
Si	Silicon
Ag	Silver
Ag-CuNPs	Silver-copper nanoparticles
AgNPs	Silver nanoparticles
AgNO ₃	Silver nitrate
H ₂ SO ₄	Sulphuric acid
ROS	Reactive oxygen species
R&D	Research & Development
SPR	Surface plasmon resonance
Ti	Titanium
TEM	Transmission electron microscopy
UV-Vis	Ultraviolet visible spectroscopy
USD	United State Dollar
H ₂ O	Water

XRD
Zn

X-ray diffraction
Zinc

REFERENCES

- Abboud, Y., Saffaj, T., Chagraoui, A., Bouari, A. El, Brouzi, K., Tanane, O., & Ihssane, B. (2014). Biosynthesis , characterization and antimicrobial activity of copper oxide nanoparticles (CONPs) produced using brown alga extract (*Bifurcaria bifurcata*). *Applied Nanoscience*, (4), 571–576. <https://doi.org/10.1007/s13204-013-0233-x>
- Ahmed, S., Ahmad, M., Swami, B. L., & Ikram, S. (2016). A review on plants extract mediated synthesis of silver nanoparticles for antimicrobial applications: A green expertise. *Journal of Advanced Research*, 7(1), 17–28. <https://doi.org/10.1016/j.jare.2015.02.007>
- Argueta-Figueroa, L., Morales-Luckie, R. a., Scougall-Vilchis, R. J., & Olea-Mejía, O. F. (2014). Synthesis, characterization and antibacterial activity of copper, nickel and bimetallic Cu–Ni nanoparticles for potential use in dental materials. *Progress in Natural Science: Materials International*, 24(4), 321–328. <https://doi.org/10.1016/j.pnsc.2014.07.002>
- Asmussen, S. V., Arenas, G. F., & Vallo, C. I. (2015). Enhanced degree of polymerization of methacrylate and epoxy resins by plasmonic heating of embedded silver nanoparticles. *Progress in Organic Coatings*, 88, 220–227. <https://doi.org/10.1016/j.porgcoat.2015.06.032>
- Awwad, A. M., Salem, N. M., & Abdeen, A. O. (2013). Green synthesis of silver nanoparticles using carob leaf extract and its antibacterial activity. *International Journal of Industrial Chemistry*, 4(29). <https://doi.org/10.1186/2228-5547-4-29>
- Azam, A., Ahmed, A. S., Oves, M., Khan, M. S., & Memic, A. (2012). Size-dependent antimicrobial properties of CuO nanoparticles against Gram-positive and -negative bacterial strains. *International Journal of Nanomedicine*, 7, 3527–3535. <https://doi.org/10.2147/IJN.S29020>
- Bankura, K. P., Maity, D., Mollick, M. M. R., Mondal, D., Bhowmick, B., Bain, M. K., ... Chattopadhyay, D. (2012). Synthesis, characterization and antimicrobial activity of dextran stabilized silver nanoparticles in aqueous medium. *Carbohydrate Polymers*, 89(4), 1159–1165. <https://doi.org/10.1016/j.carbpol.2012.03.089>
- Behra, R., Sigg, L., Clift, M. J. D., Herzog, F., Minghetti, M., Johnston, B., ... Petri-fink, A. (2013). Chemical and biochemical perspective Bioavailability of silver nanoparticles and ions : from a chemical and biochemical perspective. *Journal of the Royal Society*, 10(87), 20130396–20130396. <https://doi.org/10.1098/rsif.2013.0396>

- Bhardwaj, M., & Paul, S. (2016). Palladium nanoparticles onto ethylenediamine functionalized silica-cellulose substrates [Pd(0)-EDA/SCs]: An efficient and sustainable approach for hydrogenation of nitroarenes and carbonyl compounds under mild conditions. *Arabian Journal of Chemistry*, (0), 0–25. <https://doi.org/10.1016/j.arabjc.2016.05.008>
- Caro, C., M.Castillo, P., Klippstein, R., Pozo, D., & Zaderenko, A. P. (2010). Silver nanoparticles : sensing and imaging applications. *In Silver Nanoparticles*. InTech. 201–225 <https://doi.org/10.5772/8513>
- Chahal, S., Hussain, F. S. J., Kumar, A., Rasad, M. S. B. A., & Yusoff, M. M. (2016). Fabrication, characterization and in vitro biocompatibility of electrospun hydroxyethyl cellulose/poly (vinyl) alcohol nanofibrous composite biomaterial for bone tissue engineering. *Chemical Engineering Science*, 144, 17–29. <https://doi.org/10.1016/j.ces.2015.12.030>
- Chen, Y., Wang, C., Liu, H., Qiu, J., & Bao, X. (2005). Ag/SiO₂: a novel catalyst with high activity and selectivity for hydrogenation of chloronitrobenzenes. *Chemical Communications (Cambridge, England)*, 2(42), 5298–300. <https://doi.org/10.1039/b509595f>
- Cho, K.-H., Park, J.-E., Osaka, T., & Park, S.-G. (2005). The study of antimicrobial activity and preservative effects of nanosilver ingredient. *Electrochimica Acta*, 51(5), 956–960. <https://doi.org/10.1016/j.electacta.2005.04.071>
- Dealba-Montero, I., Guajardo-Pacheco, J., Morales-Sánchez, E., Araujo-Martínez, R., Loredó-Becerra, G. M., Martínez-Castañón, G.-A., ... Jasso, M. E. C. (2017). Antimicrobial Properties of Copper Nanoparticles and Amino Acid Chelated Copper Nanoparticles Produced by Using a Soya Extract. *Bioinorganic Chemistry and Applications*, 2017, 15–17. <https://doi.org/10.1155/2017/1064918>
- Dorjnamjin, D., Ariunaa, M., & Shim, Y. K. (2008). Synthesis of silver nanoparticles using hydroxyl functionalized ionic liquids and their antimicrobial activity. *International Journal of Molecular Sciences*, 9(5), 807–20. <https://doi.org/10.3390/ijms9050807>
- Duncan, T. V. (2011). Applications of nanotechnology in food packaging and food safety: barrier materials, antimicrobials and sensors. *Journal of Colloid and Interface Science*, 363(1), 1–24. <https://doi.org/10.1016/j.jcis.2011.07.017>

- Fayaz, A. M., Balaji, K., Girilal, M., Yadav, R., Kalaichelvan, P. T., & Venketesan, R. (2010). Biogenic synthesis of silver nanoparticles and their synergistic effect with antibiotics: a study against gram-positive and gram-negative bacteria. *Nanomedicine: Nanotechnology, Biology, and Medicine*, 6(1), 103–9. <https://doi.org/10.1016/j.nano.2009.04.006>
- Georgiev, P., Simeonova, S., Chanachev, A., Mihaylov, L., Nihtianova, D., & Balashev, K. (2016). Acceleration effect of copper(II) ions on the rate of citrate synthesis of gold nanoparticles. *Colloids and Surfaces A: Physicochemical and Engineering Aspects*, 494, 39–48. <https://doi.org/10.1016/j.colsurfa.2015.12.031>
- Ghaseminezhad, S. M., Hamed, S., & Shojaosadati, S. A. (2012). Green synthesis of silver nanoparticles by a novel method: Comparative study of their properties. *Carbohydrate Polymers*, 89(2), 467–472. <https://doi.org/10.1016/j.carbpol.2012.03.030>
- Grigorie, A. C., Muntean, C., & Stefanescu, M. (2015). Obtaining of γ -Fe₂O₃ nanoparticles by thermal decomposition of polyethyleneglycol-iron nitrate mixtures. *Thermochimica Acta*, 621, 61–67. <https://doi.org/10.1016/j.tca.2015.10.010>
- Haneda, M., & Towata, A. (2015). Catalytic performance of supported Ag nanoparticles prepared by liquid phase chemical reduction for soot oxidation. *Catalysis Today*, 242(PB), 351–356. <https://doi.org/10.1016/j.cattod.2014.05.044>
- Hassan, E. A., Hassan, M. L., Abou-zeid, R. E., & El-Wakil, N. A. (2015). Novel nanofibrillated cellulose/chitosan nanoparticles nanocomposites films and their use for paper coating. *Industrial Crops and Products*, 6–13. <https://doi.org/10.1016/j.indcrop.2015.12.006>
- Hebeish, A., Shaheen, T. I., & El-Naggar, M. E. (2016). Solid state synthesis of starch-capped silver nanoparticles. *International Journal of Biological Macromolecules*, 87, 70–76. <https://doi.org/10.1016/j.ijbiomac.2016.02.046>
- Hebeish, a. a., El-Rafie, M. H., Abdel-Mohdy, F. a., Abdel-Halim, E. S., & Emam, H. E. (2010). Carboxymethyl cellulose for green synthesis and stabilization of silver nanoparticles. *Carbohydrate Polymers*, 82(3), 933–941. <https://doi.org/10.1016/j.carbpol.2010.06.020>
- Hu, B., Wang, S.-B., Wang, K., Zhang, M., & Yu, S.-H. (2008). Microwave-Assisted Rapid Facile “Green” Synthesis of Uniform Silver Nanoparticles: Self-Assembly into Multilayered Films and Their Optical Properties. *Journal of Physical Chemistry C*, 112(30), 11169–11174. <https://doi.org/10.1021/jp801267j>

- Jamal, A., Rahman, M. M., Khan, S. B., Faisal, M., Akhtar, K., Rub, M. A., ... Al-Youbi, A. O. (2012). Cobalt doped antimony oxide nano-particles based chemical sensor and photo-catalyst for environmental pollutants. *Applied Surface Science*, *261*, 52–58. <https://doi.org/10.1016/j.apsusc.2012.07.066>
- Ju, J., Zhang, R., & Chen, W. (2016). Photochemical deposition of surface-clean silver nanoparticles on nitrogen-doped graphene quantum dots for sensitive colorimetric detection of glutathione. *Sensors and Actuators B: Chemical*, *228*, 66–73. <https://doi.org/10.1016/j.snb.2016.01.007>
- Kanmani, P., & Lim, S. T. (2013). Synthesis and characterization of pullulan-mediated silver nanoparticles and its antimicrobial activities. *Carbohydrate Polymers*, *97*(2), 421–8. <https://doi.org/10.1016/j.carbpol.2013.04.048>
- Karaođ, E., Kavas, H., Baykal, a, & Toprak, M. S. (2011). Effect of Hydrolyzing Agents on the Properties of Poly (Ethylene Glycol) -Fe₃O₄ Nanocomposite. *Nano-micro letters*, *3*(2), 79–85. <http://dx.doi.org/10.3786/nml.v3i2>.
- Katwal, R., Kaur, H., Sharma, G., Naushad, M., & Pathania, D. (2015). Electrochemical synthesized copper oxide nanoparticles for enhanced photocatalytic and antimicrobial activity. *Journal of Industrial and Engineering Chemistry*, *31*, 173–184. <https://doi.org/10.1016/j.jiec.2015.06.021>
- Khanna, P. K., Gaikwad, S., Adhyapak, P. V., Singh, N., & Marimuthu, R. (2007). Synthesis and characterization of copper nanoparticles. *Materials Letters*, *61*(25), 4711–4714. <https://doi.org/10.1016/j.matlet.2007.03.014>
- Kim, J. S., Kuk, E., Yu, K. N., Kim, J.-H., Park, S. J., Lee, H. J., ... Cho, M.-H. (2007). Antimicrobial effects of silver nanoparticles. *Nanomedicine : Nanotechnology, Biology, and Medicine*, *3*(1), 95–101. <https://doi.org/10.1016/j.nano.2006.12.001>
- Kim, Y. H., Lee, D. K., Jo, B. G., Jeong, J. H., & Kang, Y. S. (2006). Synthesis of oleate capped Cu nanoparticles by thermal decomposition. *Colloids and Surfaces A: Physicochemical and Engineering Aspects*, *284–285*, 364–368. <https://doi.org/10.1016/j.colsurfa.2005.10.067>
- Komeily-Nia, Z., Montazer, M., & Latifi, M. (2013). Synthesis of nano copper/nylon composite using ascorbic acid and CTAB. *Colloids and Surfaces A: Physicochemical and Engineering Aspects*, *439*, 167–175. <https://doi.org/10.1016/j.colsurfa.2013.03.003>
- Kora, A. J., Manjusha, R., & Arunachalam, J. (2009). Superior bactericidal activity of SDS capped silver nanoparticles: Synthesis and characterization. *Materials Science and Engineering: C*, *29*(7), 2104–2109. <https://doi.org/10.1016/j.msec.2009.04.010>

- Kora, A. J., Sashidhar, R. B., & Arunachalam, J. (2010). Gum kondagogu (*Cochlospermum gossypium*): A template for the green synthesis and stabilization of silver nanoparticles with antibacterial application. *Carbohydrate Polymers*, 82(3), 670–679. <https://doi.org/10.1016/j.carbpol.2010.05.034>
- Kumar, V. S., Nagaraja, B. M., Shashikala, V., Padmasri, a. H., Madhavendra, S. S., Raju, B. D., & Rao, K. S. R. (2004). Highly efficient Ag/C catalyst prepared by electro-chemical deposition method in controlling microorganisms in water. *Journal of Molecular Catalysis A: Chemical*, 223(1–2), 313–319. <https://doi.org/10.1016/j.molcata.2003.09.047>
- Li, Z., Lee, D., Sheng, X., Cohen, R. E., & Rubner, M. F. (2006). Two-Level Antibacterial Coating with Both Release-Killing and Contact-Killing Capabilities. *Langmuir*, 22(20), 9820–9823.
- Liz-marz, L. M. (2004). Nanometals formation and color. *Materials Today* 7(2), 26–31. [https://doi.org/10.1016/S1369-7021\(04\)00080-X](https://doi.org/10.1016/S1369-7021(04)00080-X)
- Lok, C.-N., Ho, C.-M., Chen, R., He, Q.-Y., Yu, W.-Y., Sun, H., ... Che, C.-M. (2007). Silver nanoparticles: partial oxidation and antibacterial activities. *Journal of Biological Inorganic Chemistry : JBIC : A Publication of the Society of Biological Inorganic Chemistry*, 12(4), 527–34. <https://doi.org/10.1007/s00775-007-0208-z>
- M. Awwad, A., & M. Salem, N. (2012). Green Synthesis of Silver Nanoparticles by Mulberry Leaves Extract. *Nanoscience and Nanotechnology*, 2(4), 125–128. <https://doi.org/10.5923/j.nn.20120204.06>
- Mochochoko, T., Oluwafemi, O. S., Jumbam, D. N., & Songca, S. P. (2013). Green synthesis of silver nanoparticles using cellulose extracted from an aquatic weed; water hyacinth. *Carbohydrate Polymers*, 98(1), 290–4. <https://doi.org/10.1016/j.carbpol.2013.05.038>
- Mogoşanu, G. D., Grumezescu, A. M., Bejenaru, C., & Bejenaru, L. E. (2016). Polymeric protective agents for nanoparticles in drug delivery and targeting. *International Journal of Pharmaceutics*, 510(2), 419–429. <https://doi.org/10.1016/j.ijpharm.2016.03.014>
- MubarakAli, D., Thajuddin, N., Jeganathan, K., & Gunasekaran, M. (2011). Plant extract mediated synthesis of silver and gold nanoparticles and its antibacterial activity against clinically isolated pathogens. *Colloids and Surfaces B: Biointerfaces*, 85(2), 360–365. <https://doi.org/10.1016/j.colsurfb.2011.03.009>

- Namasivayam, S. K. R., Samrat, K., & Ganesh, S. (2013). Preparation of Chitosan Stabilized Ofloxacin- Gold Nano Conjugate for the Improved Anti Bacterial Activity Against Human Pathogenic Bacteria. *Innovare Journal of Medical Science*, 1(2), 3–7. <http://innovareacademics.in/journals/index.php/ijms/article/view/154>
- Owusu-Ware, S. K., Boateng, J., Jordan, D., Portefaix, S., Tasseto, R., Ramano, C. D., & Antonijević, M. D. (2016). Molecular mobility of hydroxyethyl cellulose (HEC) films characterised by thermally stimulated currents (TSC) spectroscopy. *International Journal of Pharmaceutics*, 497(1–2), 222–227. <https://doi.org/10.1016/j.ijpharm.2015.11.052>
- Panacek, A., Kvítek, L., Pucek, R., Kolar, M., Vecerova, R., Pizúrova, N., ... Zboril, R. (2006). Silver colloid nanoparticles: synthesis, characterization, and their antibacterial activity. *The Journal of Physical Chemistry. B*, 110(33), 16248–53. <https://doi.org/10.1021/jp063826h>
- Pantidos, N., & Horsfall, L. E. (2014). Biological Synthesis of Metallic Nanoparticles by Bacteria , Fungi and Plants. *Journal of Nanomedicine & Nanotechnology*, 5(5), 10. <https://doi.org/10.4172/2157-7439.1000233>
- Park, Y., Hong, Y. N., Weyers, a, Kim, Y. S., & Linhardt, R. J. (2011). Polysaccharides and phytochemicals: a natural reservoir for the green synthesis of gold and silver nanoparticles. *IET Nanobiotechnology / IET*, 5(3), 69–78. <https://doi.org/10.1049/iet-nbt.2010.0033>
- Phong, N., Khuong, V., Tho, T., Du, C., & Minh, N. (2011). Green Synthesis of Copper Nanoparticles Colloidal Solutions and Used As Pink Disease Treatment Drug for Rubber Tree. *Proceedings of IWNA 2011*, 10–13.
- Pileni, M. P., Tanori, J., & Filankembo, A. (1997). Biomimetic strategies for the control of size, shape and self-organization of nanoparticles. *Colloids and Surfaces A: Physicochemical and Engineering Aspects*, 123–124(97), 561–573. [https://doi.org/10.1016/S0927-7757\(97\)03785-0](https://doi.org/10.1016/S0927-7757(97)03785-0)
- Pizzolato, E., Scaramuzza, S., Carraro, F., Sartori, A., Agnoli, S., Amendola, V., ... Sartorel, A. (2015). Water oxidation electrocatalysis with iron oxide nanoparticles prepared via laser ablation. *Journal of Energy Chemistry*, 0, 1–5. <https://doi.org/10.1016/j.jechem.2015.12.004>
- Porcaro, F., Battocchio, C., Antoccia, A., Fratoddi, I., Venditti, I., & Fracassi, A. (2016). Colloids and Surfaces B: Biointerfaces Synthesis of functionalized gold nanoparticles capped with 3-mercapto-1-propansulfonate and 1-thioglucose mixed thiols and “ in vitro ” bioresponse. *Colloids and Surfaces B: Biointerfaces*, 142, 408–416. <https://doi.org/10.1016/j.colsurfb.2016.03.016>

- Prabhu, S., & Poulouse, E. K. (2012). Silver nanoparticles: mechanism of antimicrobial action, synthesis, medical applications, and toxicity effects. *International Nano Letters*, 2(1), 32. <https://doi.org/10.1186/2228-5326-2-32>
- Prakash, P., Gnanaprakasam, P., Emmanuel, R., Arokiyaraj, S., & Saravanan, M. (2013). Green synthesis of silver nanoparticles from leaf extract of *Mimusops elengi*, Linn. for enhanced antibacterial activity against multi drug resistant clinical isolates. *Colloids and Surfaces. B, Biointerfaces*, 108, 255–9. <https://doi.org/10.1016/j.colsurfb.2013.03.017>
- Rai, M., Yadav, A., & Gade, A. (2009). Silver nanoparticles as a new generation of antimicrobials. *Biotechnology Advances*, 27(1), 76–83. <https://doi.org/10.1016/j.biotechadv.2008.09.002>
- Raj, D. R., Prasanth, S., Vineeshkumar, T. V, & Sudarsanakumar, C. (2016). Surface Plasmon Resonance based fiber optic sensor for mercury detection using gold nanoparticles PVA hybrid. *Optics Communications*, 367, 102–107. <https://doi.org/10.1016/j.optcom.2016.01.027>
- Ramyadevi, J., Jeyasubramanian, K., Marikani, A., Rajakumar, G., & Rahuman, A. A. (2012). Synthesis and antimicrobial activity of copper nanoparticles. *Materials Letters*, 71, 114–116. <https://doi.org/10.1016/j.matlet.2011.12.055>
- Raspolli Galletti, A. M., Antonetti, C., Marracci, M., Piccinelli, F., & Tellini, B. (2013). Novel microwave-synthesis of Cu nanoparticles in the absence of any stabilizing agent and their antibacterial and antistatic applications. *Applied Surface Science*, 280, 610–618. <https://doi.org/10.1016/j.apsusc.2013.05.035>
- Rastogi, V. K., Stanssens, D., & Samyn, P. (2016). Reaction efficiency and retention of poly(styrene-co-maleimide) nanoparticles deposited on fibrillated cellulose surfaces. *Carbohydrate Polymers*, 141, 244–252. <https://doi.org/10.1016/j.carbpol.2016.01.018>
- Ratyakshi, & Chauhan, R. P. (2009). Colloidal synthesis of silver nano particles. *Asian Journal of Chemistry*, 21(10), 113–116.
- Ren, G., Hu, D., Cheng, E. W. C., Vargas-Reus, M. a, Reip, P., & Allaker, R. P. (2009). Characterisation of copper oxide nanoparticles for antimicrobial applications. *International Journal of Antimicrobial Agents*, 33(6), 587–90. <https://doi.org/10.1016/j.ijantimicag.2008.12.004>
- Roco, M. C., Mirkin, C. A., & Hersam, M. C. (2010). Nanotechnology research directions for societal needs in 2020 – Retrospective and Outlook. *WTEC-World Technology Evaluation Center*, 476–477. <https://doi.org/10.1007/978-94-007-1168-6>

- Ruparelia, J. P., Chatterjee, A. K., Duttagupta, S. P., & Mukherji, S. (2008). Strain specificity in antimicrobial activity of silver and copper nanoparticles. *Acta Biomaterialia*, 4(3), 707–16. <https://doi.org/10.1016/j.actbio.2007.11.006>
- Sambhy, V., MacBride, M. M., Peterson, B. R., & Sen, A. (2006). Silver bromide nanoparticle/polymer composites: dual action tunable antimicrobial materials. *Journal of the American Chemical Society*, 128(30), 9798–808. <https://doi.org/10.1021/ja061442z>
- Sastry, M., Ahmad, A., Khan, M. I., & Kumar, R. (2003). Biosynthesis of metal nanoparticles using fungi and actinomycete. *Current Science*, 85(2). [https://doi.org/10.1016/S0927-7765\(02\)00174-1](https://doi.org/10.1016/S0927-7765(02)00174-1)
- Setua, P., Chakraborty, a., Seth, D., Bhatta, M. U., Satyam, P. V., & Sarkar, N. (2007). Synthesis, Optical Properties, and Surface Enhanced Raman Scattering of Silver Nanoparticles in Nonaqueous Methanol Reverse Micelles. *Journal of Physical Chemistry C*, 111(10), 3901–3907. <https://doi.org/10.1021/jp067475i>
- Shameli, K., Ahmad, M. Bin, Jazayeri, S. D., Shabanzadeh, P., Sangpour, P., Jahangirian, H., & Gharayebi, Y. (2012). Investigation of antibacterial properties silver nanoparticles prepared via green method. *Chemistry Central Journal*, 6(1), 73. <https://doi.org/10.1186/1752-153X-6-73>
- Shankar, S. S., Rai, A., Ahmad, A., & Sastry, M. (2004). Rapid synthesis of Au, Ag, and bimetallic Au core-Ag shell nanoparticles using Neem (*Azadirachta indica*) leaf broth. *Journal of Colloid and Interface Science*, 275(2), 496–502. <https://doi.org/10.1016/j.jcis.2004.03.003>
- Sharma, V. K., Yngard, R. a, & Lin, Y. (2009). Silver nanoparticles: green synthesis and their antimicrobial activities. *Advances in Colloid and Interface Science*, 145(1–2), 83–96. <https://doi.org/10.1016/j.cis.2008.09.002>
- Siddiqui, H., Qureshi, M. S., & Haque, F. Z. (2016). Effect of copper precursor salts: Facile and sustainable synthesis of controlled shaped copper oxide nanoparticles. *Optik*, 127(11), 4726–4730. <https://doi.org/10.1016/j.ijleo.2016.01.118>
- Singh, P., Singh, H., Ju, Y., Mathiyalagan, R., Wang, C., & Chun, D. (2016). Enzyme and Microbial Technology Extracellular synthesis of silver and gold nanoparticles by *Sporosarcina koreensis* DC4 and their biological applications. *Enzyme and Microbial Technology*, 86, 75–83. <https://doi.org/10.1016/j.enzmictec.2016.02.005>

- Sinha Roy, D., & Rohera, B. D. (2002). Comparative evaluation of rate of hydration and matrix erosion of HEC and HPC and study of drug release from their matrices. *European Journal of Pharmaceutical Sciences*, 16(3), 193–199. [https://doi.org/10.1016/S0928-0987\(02\)00103-3](https://doi.org/10.1016/S0928-0987(02)00103-3)
- Sirelkhatim, A., Mahmud, S., Seeni, A., Kaus, N. H. M., Ann, L. C., Bakhori, S. K. M., ... Mohamad, D. (2015). Review on zinc oxide nanoparticles: Antibacterial activity and toxicity mechanism. *Nano-Micro Letters*, 7(3), 219–242. <https://doi.org/10.1007/s40820-015-0040-x>
- Brownheim, S. V. (2011). *Characterization and in vitro toxicity of copper nanoparticles (Cu-NPs) in murine neuroblastoma (N2A) cells*. (Master thesis, Air Force Institute of Technology, Wright-Patterson Air Force Base, Ohio). Retrieved from <http://www.dtic.mil/get-tr-doc/pdf?AD=ADA539721>
- Sivaraj, R., Rahman, P. K. S. M., Rajiv, P., Salam, H. A., & Venckatesh, R. (2014). Biogenic copper oxide nanoparticles synthesis using *Tabernaemontana divaricate* leaf extract and its antibacterial activity against urinary tract pathogen. *Spectrochimica Acta Part A: Molecular and Biomolecular Spectroscopy*, 133, 178–181. <https://doi.org/10.1016/j.saa.2014.05.048>
- Soares, P. I. P., Machado, D., Laia, C., Pereira, L. C. J., Coutinho, J. T., Ferreira, I. M. M., ... Borges, J. P. (2016). Thermal and magnetic properties of chitosan-iron oxide nanoparticles. *Carbohydrate Polymers*, 149, 382–390. <https://doi.org/10.1016/j.carbpol.2016.04.123>
- Soloviev, M., & Gedanken, A. (2011). Coating a stainless steel plate with silver nanoparticles by the sonochemical method. *Ultrasonics Sonochemistry*, 18(1), 356–362. <https://doi.org/10.1016/j.ultsonch.2010.06.015>
- Sondi, I., & Salopek-Sondi, B. (2004). Silver nanoparticles as antimicrobial agent: a case study on *E. coli* as a model for Gram-negative bacteria. *Journal of Colloid and Interface Science*, 275(1), 177–82. <https://doi.org/10.1016/j.jcis.2004.02.012>
- Song, J. Y., & Kim, B. S. (2009). Rapid biological synthesis of silver nanoparticles using plant leaf extracts. *Bioprocess and Biosystems Engineering*, 32(1), 79–84. <https://doi.org/10.1007/s00449-008-0224-6>
- Song, Y., Zheng, Q., & Liu, C. (2008). Green biocomposites from wheat gluten and hydroxyethyl cellulose: Processing and properties. *Industrial Crops and Products*, 28(1), 56–62. <https://doi.org/10.1016/j.indcrop.2008.01.004>
- Soomro, R. A., Hussain Sherazi, S. T., Sirajuddin, Memon, N., Shah, M. R., Kalwar, N. H., ... Shah, A. (2014). Synthesis of air stable copper nanoparticles and their use in catalysis. *Advanced Materials Letters*, 5(4), 191–198. <https://doi.org/10.5185/amlett.2013.8541>

- Sousa, C., Botelho, C., & Oliveira, R. (2011). Nanotechnology applied to medical biofilms control. *Science against Microbial Pathogens: Communicating Current Research and Technological Advances*, 878–888. Retrieved from <http://repositorium.sdum.uminho.pt/handle/1822/22521>
- Swarnkar, R. K., Singh, S. C., & Gopal, R. (2012). Effect of aging on copper nanoparticles synthesized by pulsed laser ablation in water: structural and optical characterizations. *Bulletin of Materials Science*, 34(7), 1363–1369. <https://doi.org/10.1007/s12034-011-0329-4>
- Taghizadeh, M. T., & Seifi-Aghjekohal, P. (2015). Sonocatalytic degradation of 2-hydroxyethyl cellulose in the presence of some nanoparticles. *Ultrasonics Sonochemistry*, 26, 265–272. <https://doi.org/10.1016/j.ultsonch.2014.12.014>
- Tamilvanan, a., Balamurugan, K., Ponappa, K., & Kumar, B. M. (2014). Copper Nanoparticles: Synthetic Strategies, Properties and Multifunctional Application. *International Journal of Nanoscience*, 13(2), 1430001. <https://doi.org/10.1142/S0219581X14300016>
- Tanna, J. A., Chaudhary, R. G., Gandhare, N. V., Rai, A. R., Yerpude, S., & Juneja, H. D. (2016). Copper nanoparticles catalysed an efficient one-pot multicomponents synthesis of chromenes derivatives and its antibacterial activity. *Journal of Experimental Nanoscience*, 11(11), 884–900. <https://doi.org/10.1080/17458080.2016.1177216>
- Taveira, S. F., Nomizo, A., & Lopez, R. F. V. (2009). Effect of the iontophoresis of a chitosan gel on doxorubicin skin penetration and cytotoxicity. *Journal of Controlled Release*, 134(1), 35–40. <https://doi.org/10.1016/j.jconrel.2008.11.002>
- Theivasanthi, T., & Alagar, M. (2011). Studies of Copper Nanoparticles Effects on Micro-organisms. *Annals of Biological Research*, 2(3), 368–373
- Thirumurugan, A., Aswitha, P., Kiruthika, C., Nagarajan, S., & Christy, A. N. (2016). Green synthesis of platinum nanoparticles using *Azadirachta indica* – An eco-friendly approach. *Materials Letters*, 170, 175–178. <https://doi.org/10.1016/j.matlet.2016.02.026>
- Tran, Q. H., Nguyen, V. Q., & Le, A.-T. (2013). Silver nanoparticles: synthesis, properties, toxicology, applications and perspectives. *Advances in Natural Sciences: Nanoscience and Nanotechnology*, 4(3), 33001. <https://doi.org/10.1088/2043-6262/4/3/033001>

- Valodkar, M., Modi, S., Pal, A., & Thakore, S. (2011). Synthesis and anti-bacterial activity of Cu, Ag and Cu-Ag alloy nanoparticles: A green approach. *Materials Research Bulletin*, 46(3), 384–389. <https://doi.org/10.1016/j.materresbull.2010.12.001>
- Venditti, I., Hassanein, T. F., Fratoddi, I., Fontana, L., Battocchio, C., Rinaldi, F., ... Russo, M. V. (2015). Colloids and Surfaces B: Biointerfaces Bioconjugation of gold-polymer core – shell nanoparticles with bovine serum amine oxidase for biomedical applications. *Colloids and Surfaces B: Biointerfaces*, 134, 314–321. <https://doi.org/10.1016/j.colsurfb.2015.06.052>
- Vigneshwaran, N., Nachane, R. P., Balasubramanya, R. H., & Varadarajan, P. V. (2006). A novel one-pot “green” synthesis of stable silver nanoparticles using soluble starch. *Carbohydrate Research*, 341(12), 2012–8. <https://doi.org/10.1016/j.carres.2006.04.042>
- Vimala, K., Samba Sivudu, K., Murali Mohan, Y., Sreedhar, B., & Mohana Raju, K. (2009). Controlled silver nanoparticles synthesis in semi-hydrogel networks of poly(acrylamide) and carbohydrates: A rational methodology for antibacterial application. *Carbohydrate Polymers*, 75(3), 463–471. <https://doi.org/10.1016/j.carbpol.2008.08.009>
- Wan, C., & Li, J. (2016). Cellulose aerogels functionalized with polypyrrole and silver nanoparticles: in-situ synthesis, characterization and antibacterial activity. *Carbohydrate Polymers*, 146, 362–367. <https://doi.org/10.1016/j.carbpol.2016.03.081>
- Wei, D., Sun, W., Qian, W., Ye, Y., & Ma, X. (2009). The synthesis of chitosan-based silver nanoparticles and their antibacterial activity. *Carbohydrate Research*, 344(17), 2375–82. <https://doi.org/10.1016/j.carres.2009.09.001>
- Wong, K. K. Y., & Liu, X. (2010). Silver nanoparticles—the real “silver bullet” in clinical medicine? *MedChemComm*, 1(2), 125. <https://doi.org/10.1039/c0md00069h>
- Wu, C. L., & Chen, Y. (2015). Hydroxyethyl cellulose filled with M^{2+} chelate complexes with ethylenediaminetetraacetic acid (EDTA) as an effective electron-injection layer for polymer light-emitting diodes. *Organic Electronics: Physics, Materials, Applications*, 25, 156–164. <https://doi.org/10.1016/j.orgel.2015.06.035>
- Xiong, J., Wang, Y., Xue, Q., & Wu, X. (2011). Synthesis of highly stable dispersions of nanosized copper particles using l-ascorbic acid. *Green Chemistry*, 13(4), 900. <https://doi.org/10.1039/c0gc00772b>

- Xiong, R., Lu, C., Zhang, W., Zhou, Z., & Zhang, X. (2013). Facile synthesis of tunable silver nanostructures for antibacterial application using cellulose nanocrystals. *Carbohydrate Polymers*, 95(1), 214–9. <https://doi.org/10.1016/j.carbpol.2013.02.077>
- Xu, W., Jin, W., Lin, L., Zhang, C., Li, Z., Li, Y., ... Li, B. (2014). Green synthesis of xanthan conformation-based silver nanoparticles: Antibacterial and catalytic application. *Carbohydrate Polymers*, 101, 961–7. <https://doi.org/10.1016/j.carbpol.2013.10.032>
- Yi, W., Hong-ju, M. A. O., Guo-qing, Z., Hong-lian, Z., Qing-hui, J. I. N., & Jian-long, Z. (2010). Detection of Hepatitis B Virus Deoxyribonucleic Acid Based on Gold Nanoparticle Probe Chip. *Chinese Journal of Analytical Chemistry*, 38(8), 1133–1138. [https://doi.org/10.1016/S1872-2040\(09\)60062-1](https://doi.org/10.1016/S1872-2040(09)60062-1)
- Yoon, K. Y., Hoon Byeon, J., Park, J. H., & Hwang, J. (2007). Susceptibility constants of Escherichia coli and Bacillus subtilis to silver and copper nanoparticles. *Science of the Total Environment*, 373(2–3), 572–575. <https://doi.org/10.1016/j.scitotenv.2006.11.007>
- Zain, N. M., Stapley, a. G. F., & Shama, G. (2014). Green synthesis of silver and copper nanoparticles using ascorbic acid and chitosan for antimicrobial applications. *Carbohydrate Polymers*, 112, 195–202. <https://doi.org/10.1016/j.carbpol.2014.05.081>
- Zhang, L.-M. (1999). Synergistic blends from aqueous solutions of two cellulose derivatives. *Colloid Polymer Science*, 277, 886–890. <https://doi.org/10.1007/s003960050466>
- Zheng, W., Hu, L., Lee, L. Y. S., & Wong, K.-Y. (2016). Copper nanoparticles/polyaniline/graphene composite as a highly sensitive electrochemical glucose sensor. *Journal of Electroanalytical Chemistry*, 6–11. <https://doi.org/10.1016/j.jelechem.2016.08.004>