

THE ENVIRONMENTAL FATE
AND MOBILITY OF OESTROGENS
INTO SOILS

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SUPERVISOR'S DECLARATION

I hereby declare that I have checked this thesis and in my opinion, this thesis is adequate in terms of scope and quality for the award of the degree of Master of Engineering in Civil.

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I hereby declare that the work in this thesis is based on my original work except for quotations and citations which have been duly acknowledged. I also declare that it has not been previously or concurrently submitted for any other degree at Universiti Malaysia Pahang or any other institutions.

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ABSTRAK

Oestrogen merupakan gangguan endokrin terbesar di dalam alam sekitar yang menjadi satu kebimbangan. Oestrogen di laporkan sebagai compound yang berpotensi untuk mengganggu alam sekitar. Oleh itu, kajian ini dijalankan untuk mengetahui lebih lanjut tentang kadar serapan oestrogen di dalam tanah yang berbeza, hubungan antara pekali air-oktanol dan pekali octanol-tanah bagi oestrogen semulajadi dan buatan dan juga kesan ke atas alam sekitar. Kajian ini dimulai dengan menyiasat jenis serapan dan faktor-faktor yang mempengaruhi kadar serapan oestrogens di dalam tanah. Kadar serapan oestrogen menunjukkan penyerapan yang tinggi dengan karbon organik, peningkatan kapasiti serapan adalah selari dengan karbon organik. Nilai K_{ow} dan K_{oc} digunakan untuk mengenalpasti pergerakan oestrogen di dalam tanah dan hasilnya, E2 dan EE2 dikategorikan dengan perlahan dan tiada pergerakan manakala E1 dikategorikan dengan sedikit pergerakan dan pergerakan yang perlahan. Nilai karbon organik memainkan peranan yang penting bagi mengetahui kadar serapan oestrogen di dalam tanah. Selain itu, faktor kadar serapan oestrogen turut dipengaruhi oleh kepelbagaian suhu dan pH. Analisis yang dijalankan membuktikan bahawa serapan oestrogen didalam tanah disebabkan oleh proses 'physisorption'. Kesemua oestrogen menunjukkan ikatan yang lemah terhadap tanah disebabkan oleh kadar serapan oestrogen ke dalam tanah bergantung kepada 'hydrophobicity'. Hubungan antara pekali air-oktanol (K_{ow}) dan pekali air-tanah (K_d) di tentukan berdasarkan kadar serapan antara oestrogen dan juga nilai bagi pekali air-oktanol (K_{ow}). Nilai log K_{ow} yang tinggi menunjukkan kurang pergerakan berlaku di dalam tanah. Hal ini menunjukkan 'hydrophobicity' bagi oestrogen memainkan peranan penting bagi proses serapannya didalam tanah. Akhirnya, hasil kajian ini juga menunjukkan perbezaan yang ketara antara nilai-nilai eksperimen dan nilai-nilai yang dihasilkan oleh model pengiraan, 'Estimation Program Interface' (EPISUITE). EPISUITE menganggarkan nilai yang berkurangan bagi log K_{ow} estrone (E1) tetapi nilai yang lebih besar bagi log K_{ow} 17 β -estradiol (E2) dan 17 α -ethnylestradiol (EE2). EPISUITE menganggarkan nilai yang lebih besar bagi oestrogen di dalam keadaan alam sekitar-tanah. Ini menunjukkan, apabila nilai berdasarkan kajian dimasukkan ke dalam model pengiraan, nilai menunjukkan perbezaan disebabkan oleh keadaan persekitaran yang berlaku. Oleh itu, kajian ini membuktikan bahawa walaupun model pengiraan digunakan, namun ia tidak menilai dengan tepat kesan terhadap persekitaran oleh bahan pencemar. Eksperimen tetap perlu dilakukan untuk mendapatkan pembahagian sebenar oestrogen dalam persekitaran.

ABSTRACT

The behaviour of oestrogens in the environment is of great concern due to the endocrine disruption potential. The occurrence of oestrogens is reported as the most potent compounds in the environment, thus this study was conducted to determine the sorption affinity of oestrogens in different types of soils, relationship with between octanol-water partition coefficient and soil-water partition coefficient of natural and synthetic oestrogen and distribution of oestrogen in environment. The research started with the investigation of the sorption behaviour and factors that influences the sorption behaviour of oestrogens in different type of soils. The sorption behaviour of all oestrogens indicated high association with organic carbon of the soils as sorption capacity increased when organic carbon increased. The value of K_{ow} and K_{oc} used to classify the mobility of oestrogens into soils and as the result, while E2 and EE2 were classified as low to immobile and E1 was slight to low. The values of organic carbon plays an important role in adsorbing an amount of oestrogens in soils. The significant effect on oestrogens' sorption factors was observed in soils with varied temperature and pH. Based on the analysis, it was clear that the sorption mechanism of oestrogens into soils was physisorption. All the oestrogens indicated a weaker binding to all soils as sorption behaviour of oestrogens onto soils was dependent of their hydrophobicity. The relationship between octanol-water partitioning coefficient (K_{ow}) and soil-water partitioning coefficient (K_d) was being determine from the order of sorption among oestrogens was well correlated with their octanol-water partitioning coefficient (K_{ow}). The higher log K_{ow} values correlated to less mobile organic chemicals into soils. It is believed that hydrophobicity of oestrogens has play a major important role in regulating the sorption of oestrogens in soils. Finally, the result of this study also indicated a considerable difference between experimental values and values generated by the computational model, Estimation Program Interface (EPISUITE). The EPISUITE software underestimate the log K_{ow} value of estrone (E1) but overestimates log K_{ow} 17 β -estradiol (E2) and 17 α -ethnylestradiol (EE2). The result also shows that EPISUITE program was overestimated the oestrogens' partition in soils-environmental compartment. It can be concluded that by inputting the experimental values, the model changed remarkably and has given absolute values to the soil's ambient conditions. Thus, this study showed that although the computational model was indicative, it did not accurately assess the environmental impact of pollutant. The experiments need to be conducted to obtain the real partitioning of oestrogens in environment.

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LIST OF SYMBOLS

%	Percent
$\mu\text{g/g}$	Microgram per gram
$\mu\text{g/l}$	Microgram per litre
μm	Micro metre
cm	Centimetre
G	Gram
g/mol	Gram per mol
H	Hour
K	Kelvin
K_d	Soil water partition coefficient
K_F	Freundlich sorption coefficient
kg/l	Kilogram per litre
K_L	Langmuir sorption coefficient
km	Kilometre
K_{oc}	Organic carbon partition coefficient
K_{ow}	Octanol-water partition coefficient
K_P	Partition coefficient
l/d	Litre per day
l/kg	Litre per kilogram
l/mg	Litre per milligram
M	Molarity
m^3/mol	Cubic metre per mol
mg	Milligram
Mg/ha	Mega gram per hectare
mg/l	Milligram per litre
mg/m^3	Milligram per cubic metre
min^{-1}	Per minute
ml	Millilitre
ml/min	Milligram per minute
mm	Millimetre
mm Hg	Millimetre of mercury

mol/l	Mol per litre
mPa	Mega Pascal
ng/g	Nanogram per gram
ng/kg	Nanogram per kilogram
ng/l	Nanogram per litre
ng/m ³	Nanogram per cubic metre
Nm	Nano metre
°C	Degree Celsius
Pa	Pascal
ppb	Part per billion
ppt	Part per thousand
rpm	Revolution per minute
Sw	Water solubility

LIST OF ABBREVIATIONS

AACC	American Association For Clinical Chemistry
ACN	Acetonitrile
ASTM	American Society Of Testing And Materials
BBP	Butyl-Benzene Phthalate
BCF	Bio Concentration Factor
BPA	Bisphenol A
BS	British Standard
CAFO	Concentrated Animal Feeding Operations
CAS	Chemical Abstracts Service
DBP	Dibutyl Phthalate
DDT	Dichlorodiphenyltrichloroethane
DMP	Dimethyl Phthalate
DNA	Deoxyribonucleic Acid
DOP	Dioctyl Phthalate
DT	Dissipation Times
E.coli	Eshericia Coli
E1	Estrone
E2	17 β -Estradiol
E3	Estriol
EDCs	Endocrine Disrupting Compounds
EE2	17 α -Ethinylestradiol
EEDs	Environmental Endocrine Disruptors
EFMs	Environmental Fate Models
EJF	Environmental Justice Foundation
EPA	Environmental Protection Act
EPISUITE	Estimation Program Interface
EREs	Oestrogen Responsive Elements
FMT	Free Malaysia Today
FOA	The Fiber Optic Association
GF/C	Glass Fiber Filter
HCL	Acid Hydrochloride

HMW	High Molecular Weight
HOC	Hydrophobic Organic Chemicals
HPLC	High Performance Liquid Chromatography
HRT	Hormone Replacement Therapy
LH	Luteinizing Hormone
LMW	Low Molecular Weight
LOD	Limit Of Detection
MCL	Maximum Contaminant Level
MeEE2	Mestranol
MeOH	Methanol
MEPH	Monoethylexyl Phthalate
NaCl	Sodium Chloride
NaOH	Natrium Hydroxide
NWHN	The National Women's Health Network
OC	Organic Carbon
OECD	Organisation For Economic Co-Operation And Development
OM	Organic Matter
PCBs	Polychlorinated Biphenyl
PERs	Property Estimation Routine
RP-HPLC	Reverse Phase High Performance Liquid Chromatography
RSD	Relative Standard Deviation
SD	Standard Deviation
SHBG	Steroid Hormone-Binding Globulin
SPE	Solid Phase Extraction
SRC	Syracuse Research Corporation
SSA	Specific Surface Area
STPs	Sewerage Treatment Plants
TEBG	Testosterone-Oestrogen-Binding Globulin
UMP	Universiti Malaysia Pahang
USEPA	United State Environmental Protection Agency
UV	Ultraviolet
WHO	World Health Organization
WWTP's	Waste Water Treatment Plants

REFERENCES

- AACC. (2015). Steroid Hormone-Binding Globulin (SHBG). *Environment Science Technology*, 49(1), 570-579.
- Aaron, M., Nicholas, B., Ling, S., Stephen, C.W. and Min, L. (2016). Oestrogen and insulin transport through the blood-brain barrier. *Physiology & Behavior*, 163(1), 312-321.
- Akahori, Y., Nakai, M., Yamasaki, K., Takatsuki, M., Shimohigashi, Y. and Ohtaki, M. (2008). Relationship between the results of in vitro receptor binding assay to human oestrogen receptor α and in vivo uterotrophic assay: comparative study with 65 selected chemicals. *Toxicology In Vitro*, 22(1), 225–231.
- Aksglaede, A., Juul, H., Leffers, N.E., Skakkebaek, A. and Andersson, M. (2006). The sensitivity of the child to sex steroids: possible impact of exogenous oestrogens. *Human Reproduction*, 12(4), 341-349.
- Alcántara, M.I., Amaya, G., Matamoros, T. and Pde, G. (2017). Altered functionality of the corticotrophin-releasing hormone receptor-2 in the hypothalamic paraventricular nucleus of hyperphagic maternally separated rats. *Neuropeptides*, 63(1), 75-82.
- Alexander, V., Sirotkin, L. and Abdel, H. H. (2014). Phytoestrogens and their effects. *European Journal of Pharmacology*, 741(15), 230-236.
- Ali, S., Michael, J.A., John, D.W., and Bruce, B.J. (2006). Aqueous Solubilities of Estrone, 17β -Estradiol, 17α -Ethinylestradiol, and Bisphenol A. *Journal of Chemical Engineering and Data*, 51(3), 879–881.
- Amit, C. and Mario, A.B. (2016). Oestrogen repression of microRNA as a potential cause of cancer. *Biomedicine & Pharmacotherapy*, 78(1), 234-238.
- André, M.P.T., Pereira, L., Silveira, C., Laranjeira, L., M. Meiselb, C. and Lino, A.P. (2017). Human pharmaceuticals in Portuguese rivers: The impact of water scarcity in the environmental risk. *Science Total Environment*, 609(1), 1182-1191.
- Andrew, C.J. and Yihong, C. (2017). Does exposure to domestic wastewater effluent (including steroid oestrogens) harm fish populations in the UK?. *Science Total Environment*, 589(1), 89-96.
- Annika, I.S., Patrik, C.E., Stefan, M. and Willför, L. (2012). Content, composition, and stereochemical characterisation of lignans in berries and seeds. *Food Chemistry Reproductive Biology*, 134(4), 1991-1998.
- Arpita, B., Julien, F. and Gerber, A. (2017). Ecological distribution conflicts (EDCs) over mineral extraction in India: An overview. *The Extractive Industries and Society*, 4(3), 548-563.

- Ashok, K.S., Shveta, G., Kuldip, K., Satish, G., Yogesh, C., Arun, G. and Richa, S. (2013). Quantitative analysis of conjugated and free oestrogens in swine manure: Solutions to overcome analytical problems due to matrix effects. *Journal of Chromatography A*, 1305(13), 203-212.
- ASTM. (2000). Standard test method for moisture, ash and organic matter of peat and other organic soils.
- ASTM. (2001). Standard test method for determining a sorption constant (K_{oc}) for an organic chemical in soil and sediments.
- Bagnall, J.P., Ito, A., McAdam, E.J., Soares, A., Lester, J.N., and Cartmell, E. (2012). Resource dependent biodegradation of estrogens and the role of ammonia oxidising and heterotrophic bacteria. *Journal of Hazardous Materials*, 239(2), 56-63.
- Barbora, J., Jakub, J., Ondřej, A. and Klára, H. (2015). Phytoestrogens and mycoestrogens in surface waters - Their sources, occurrence, and potential contribution to oestrogenic activity. *Environment International*, 81(1), 26-44.
- Barel, C., Shore, L.S., Shemesh, M., Wenzel, A., Mueller, J. and Kronfeld, S. (2006). Monitoring of natural and synthetic hormones in a polluted river. *Journal of Environmental Manage*, 78(2), 16–23.
- Barron, S. and Stearns, L. (2009). No increase in breast cancer recurrence with concurrent use of tamoxifen and some CYP2D6-inhibiting medications. *breast diseases: A Year Book Quarterly*, 21(2), 166-167.
- Beauvais, M., Silva, H. and Powell, S. (2010). Human health risk assessment of endosulfan. Part IV: Occupational reentry and public non-dietary exposure and risk. *Regulatory Toxicology and Pharmacology*, 56(1), 38-50.
- Blokland, E.F., Tricht, L.A., Ginkel, S. and Sterk, S. (2017). Applicability of an innovative steroid-profiling method to determine synthetic growth promoter abuse in cattle. *The Journal of Steroid Biochemistry and Molecular Biology*, 174(9), 265-275.
- Casey, F.M., Hakk, H., Šimunek, J. and Larsen, G.L. (2003). Fate and transport of testosterone in agricultural soils. *Environmental Science and Technology*, 38(3), 790–798.
- Casey, F. M., Šimunek, J., Lee, J., Larsen, G.L. and Hakk, H. (2005). Sorption, mobility, and transformation of oestrogenic hormones in natural soil. *Journal of Environment Quality*, 34(4), 1372–1379.
- Chang, P.Y., Rula, A.D. and Kung, H. C. (2013). Microbial degradation of steroidal oestrogens. *Chemosphere*, 91(9), 1225-1235.

- Cheng, Z., Yao, F., Yuan, W.L., Hui, Q.C., Zhao, X.L. and Xue, W.Y. (2017). Uptake and translocation of organic pollutants in plants: A review. *Journal of Integrative Agriculture*, 16(8), 1659-1668.
- Cristina, L., Alia, T., Timothy, A.D., Stuart, P., and Martin, B. (2016). Soil nitrous oxide emissions in forage systems fertilized with liquid dairy manure and inorganic fertilizers. *Agriculture, Ecosystems & Environment*, 225(1), 160-172.
- Czajka, P., and Londry, K. (2006). Anaerobic biotransformation of estrogens. *Science of The Total Environment*, 367(2-3), 932-941.
- Dalel, B., Rim, B., Ikram, J., Jalel, B., Moneem, K., Habib, A. and John, L.Z. (2015). Fate of selected oestrogenic hormones in an urban sewage treatment plant in Tunisia (North Africa). *Science Total Environment*, 505(1), 154-160.
- Dan, X.S. and Lin, S.X. (2016). Mimicking postmenopausal steroid metabolism in breast cancer cell culture: Differences in response to DHEA or other steroids as hormone sources. *The Journal of Steroid Biochemistry and Molecular Biology*, 161(2), (92-100).
- D'Ascenzo, G., Di Corcia, A., Gentili, A., Mancini, R., Mastropasqua, R., Nazzari, M. and Samperi, R. (2003). Fate of natural oestrogen conjugates in municipal sewage transport and treatment facilities. *Science Total Environment*, 302(1-3), 199-209.
- David, J.F., Ekrem, K., William, A., Arnold, B.L., Barber, E.F., Kaufenberg, P.L., et al. (2015). Sediment–water distribution of contaminants of emerging concern in a mixed use watershed. *Science of The Total Environment*. 505(1), 896-904.
- Donald, W.P., Robert, T.R., Jill, E.S. and Geoffrey, A.H. (2018). Chapter 20 – Hormone receptors interact with other nuclear proteins to influence hormone responsiveness. *Principles of Hormone/Behavior Relations*, 18(2), 385–398.
- Dorota, E., Bronowicka, K., Margarita, L., Paweł, P. and Jagodziński, T. (2016). The role and impact of oestrogens and xenoestrogen on the development of cervical cancer. *Biomedicine & Pharmacotherapy*, 84(1). 1945-1953.
- Douglas, O., Pino, H., Yoan, P., David, H., Giovanni, E., Eric, D., Hullebusch, M. and Oturan, A. (2017). Removal mechanisms in aerobic slurry bioreactors for remediation of soils and sediments polluted with hydrophobic organic compounds: An overview. *Journal of Hazardous Materials*, 339(7), 427-449.
- Duarte, D. and Jones. W. (1996). Screening the environmental fate of organic contaminants in sewage sludge applied to agricultural soils: II. The potential for transfers to plants and grazing animals. *Science Total Environment*, 185(1–3), 59-70.
- EJF. (2002). Protecting people and wildlife from toxic pesticides. *Environment Science Technology*, 63(2), 441-448.

- Elke, D. and Shokoufeh, M. (2017). Targeted basic research to highlight the role of oestrogen and oestrogen receptors in the cardiovascular system. *Pharmacological Research*, 119(1), 27-35.
- Essandoh, H. K., Tizaoui, C. and Mohamed, H.A. (2012). Removal of estrone (E1), 17 β -estradiol (E2) and 17 α -Ethinylestradiol (EE2) during soil aquifer treatment of a model wastewater. *Separation Science and Technology*, 747(6), 777-787.
- Ewa, S., Lidy, M.B., Verburg, K., Magdalena, M. and Magdalena, C. (2017). Review article: Oestrogen-dependent seasonal adaptations in the immune response of fish. *Hormones and Behavior*, 88(1), 15-24.
- FAO. (2008). Experiences on implementation of the Rotterdam Convention from six countries in Asia. *Environment Science Technology*, 26(7), 471-477.
- Fisher, D.J., Staver, K.W. and Yonkos, L.T. (2005). Poultry litter-associated contaminants: environmental fate and effects on fish. *Maryland Center for Agro-Ecology*, 88, 52-66.
- FMT. (2015). Banned pesticides still being used in Cameron Highlands. *Environment Science Technology*, 69(3), 571-578.
- Getnet, D.B., Rehan, S., Kevin, A.M. and Solomon, T. (2015). Ecological risk assessment of acid rock drainage under uncertainty: The fugacity approach. *Environmental Technology & Innovation*, 4(1), 187-196.
- Gian, M.T. and Adalisa, P. (2016). Fetal safety profile of aromatase inhibitors: Animal data. *Reproductive Toxicology*, 66(1), 84-92.
- Gunnhild, P.O., Marianne, O.O., Sindre, A.P., Robert, J.L. and Augustine, A. (2014). Effects of elevated dissolved carbon dioxide and perfluorooctane sulfonic acid, given singly and in combination, on steroidogenic and biotransformation pathways of Atlantic cod. *Aquatic Toxicology*, 155(4), 222-235.
- Hall, J.M. and McDonnell, D.P. (2005). Coregulators in nuclear oestrogen receptor action: from concept to therapeutic targeting. *Molecular Interventions*, 5(6), 343-357.
- Hamed, S., and Mindy, S.K. (2015). Estrogen metabolism and breast cancer. *Cancer Letters*, 356(2), 231-243.
- Hines, M. (2017). Gonadal hormones and sexual differentiation of human brain and behavior. *Neuroscience and Biobehavioral Psychology Hormones, Brain and Behavior (Third Edition)*, 5(1), 247-278.
- Holthaus, K.I.E., Johnson, A.C., Jürgens, M.D., Williams, R.J., Smith, J.J.L. and Carter, J.E. (2002). The potential for estradiol and ethinylestradiol to sorb to suspended and bed sediments in some English rivers. *Environment Toxicology Chemistry*. 21(1), 2526- 2535.

- Hoogsteen, M.J.J., Lantinga, E.A., Bakker, E.J., Tittonell, P.A. (2015). Estimating soil organic carbon through loss on ignition: effects of ignition conditions and structural water loss. *European Journal of Soil Science*, 66(2), 320-328.
- Hui, J.X., Fu, W.G., Wei, T. and Shu, G.W. (2011). A short-term study on the interaction of bacteria, fungi and endosulfan in soil microcosm. *Science Total Environment*, 412(11), 375-379.
- Hui, Z., Jianghong, S., Xiaowei, L., Xinmin, Z. and Qingcai, C. (2014). Occurrence and removal of free oestrogens, conjugated oestrogens, and bisphenol A in manure treatment facilities in East China. *Water Research*, 58(4), 248-257.
- Ifelebuegu, J.N., Lester, J., Churchley, S. and Cart-mell, E. (2006). Removal of an endocrine disrupting chemical (17alpha-ethinyloestradiol) from wastewater effluent by activated carbon adsorption: *Journal of Environmental Technology*, 27(7), 1343-1349.
- Innes, M. (2013). Economics of agricultural residuals and overfertilization: Chemical fertilizer use, livestock waste, manure management, and environmental impacts. *Reference Module in Earth Systems and Environmental Sciences*, 2(6), 50-57.
- Isobe, H. N., Nakashima, A. and Takada, H. (2001). Distribution and behaviour of nonylphenol, octylphenol, and nonylphenol monoethoxylate in Tokyo metropolitan area: Their association with aquatic particles and sedimentary distributions. *Science Total Environment*, 35(6), 1041-1049.
- James, E.H. (2016). Interaction of the role of Concentrated Animal Feeding Operations (CAFOs) in Emerging Infectious Diseases (EIDS). *Infection, Genetics and Evolution*, 38(5), 44-46.
- James, L.G., Thomas, B., Edward, T.F., Jessica, G.D., Tracy, J.Y., Yun, Y.Y. and Dana, W.K. (2017). Rainfall-runoff of anthropogenic waste indicators from agricultural fields applied with municipal biosolids. *Science Total Environment*, 580(9), 83-89.
- Jian, C., Hu, Z., Yan, S., Yu, F.L. and Qing, R.Q. (2012). Synthesis of a dendritic oestrogen cluster: A potential tool for studies of nuclear *versus* extra nuclear pathways of oestrogen actions. *Chinese Chemical Letters*, 23(12), 1319-1322.
- Jian, G., Dandan, D., Yu, Y., Yong, R. and Diyun, C. (2016). Seasonal variation and partitioning of endocrine disrupting chemicals in waters and sediments of the Pearl River system, South China. *Environmental Pollution*, 219(6), 735-741.
- Jiho, L., Jaeweon, C., and Sang, D.K. (2011). Influence of 17 β -estradiol binding by dissolved organic matter isolated from wastewater effluent on oestrogenic activity. *Ecotoxicology and Environmental Safety*, 74(5), 1280-1287.
- Jiménez, O.M.M., Granados, L., Oliva, J., Quiroz, J. and Barrón, M. (2010). Nutritive value of *Brachiaria humidicola* with organic and inorganic fertilization in acid soils. *Archivos Zootecnia*, 59(228), 561-570.

- Jodi, L.S., Hugues, O., Yun, Z., Shannon, L. and Bartelt, H. (2015). The effect of particle size on sorption of oestrogens, androgens and progestagens in aquatic sediment. *Journal of Hazardous Materials*, 299(15), 1112-1121.
- Johnson, A.C. and Sumpter, J.P. (2001). Removal of endocrine-disrupting chemicals in activated sludge treatment works. *Environmental Science Technology*, 35(24), 4697–4703.
- Johnson, R., Aerni, A., Gerritsen, M., Gibert, W., Giger, K., Hylland, M., Jürgens, T. (2005). Comparing steroid oestrogen and nonylphenol content across a range of European sewage plants with different treatment and management practices. *Water Resource*, 39(21), 47-58.
- José, M.M., Anne, H., Isabelle, J., Daniel, Z., Jean, P.C., Mariana, F.F., Arnaud, P. et al. (2016). Steroid receptor profiling of vinclozolin and its primary metabolites. *Toxicology and Applied Pharmacology*, 216(8), 44-54.
- Kabir, M.S. and Rahman, I. (2015). A review on endocrine disruptors and their possible impacts on human health. *Environmental Toxicology Pharmacology*, 40(4), 241-258.
- Kancanamayoon, W. and Tatrahun, N. (2008). Determination of polycyclic aromatic hydrocarbons in water samples by solid phase extraction and gas chromatography. *World Journal of Chemistry*, 3(2), 51-54.
- Karin, B. and Loretta, L. (2015). Evaluation of low-cost materials for sorption of hydrophobic organic pollutants in storm water. *Journal of Environmental Management*, 159(24), 106-114.
- Khanal, S.K., Xie, M.L. Thompson, S., Sung, S.O. and van Leeuwen, J. (2006). Fate, transport, and biodegradation of natural oestrogens in the environment and engineered systems. *Environmental of Science and Technology*, 48(5), 6537-6546.
- Koh, Y.K.K., Chiu, T.Y., Boobis, A.R., Scrimshaw, M.D., Bagnall, J.P., Soares, A., Pollard, S. et al. (2009). The influence of operating conditions on the biodegradation of steroid oestrogens and nonylphenolic compounds during wastewater treatment processes. *Environmental Science and Technology*, 43(17), 6646-6654.
- Kolodziej, E.P., Harter, T. and Sedlak, D.L. (2004). Dairy wastewater, aquaculture, and spawning fish as sources of steroid hormones in the aquatic environment. *Environmental Science and Technology*, 38(23), 6377–6384.
- Kozarek, M.L., Wolfe, N.G., Love, K.F. and Knowlton, K. (2008). Sorption of oestrogen to three agricultural soils from Virginia, USA. *Transactions of the ASABE*, 51(5), 1591-1597.

- Kumar, S.V., Mohan, P. and Sarma, N. (2009). Sorptive removal of endocrine-disruptive compound (estriol, E3) from aqueous phase by batch and column studies: kinetic and mechanistic evaluation. *Journal of Hazardous Materials*, 164(2), 820-828.
- Kuster, S.P., Schindler, C., Rochat, M.K., Braun, J., Held, L., and Brandli, O. (2008). Reference equations for lung function screening of healthy never-smoking adults aged 18–80 years. *European Respiratory Journal*, 31, 860–868.
- Kyung, A., Hwang, K. and Chul, C. (2015). Chapter One – Endocrine disrupting chemicals with oestrogenicity posing the risk of cancer progression in oestrogen-responsive organs. *Advances in Molecular Toxicology*, 9, 1-33.
- Lahnsteiner, F., Berger, B., Kletzl, M. and Weismann, T. (2006). Effect of 17 β -estradiol on gamete quality and maturation in two salmonid species. *Aquatic Toxicology*, 79(2), 124–131.
- Lai, K.M., Johnson, K.L., Scrimshaw, M.D. and Lester, J.N. (2000). Binding of waterborne steroid oestrogens to solid phases in river and estuarine systems. *Environmental Science and Technology*, 34(18), 3890–3894.
- Lai, K.M., Scrimshaw, M.D. and Lester, J.N. (2002). The effects of natural and synthetic steroid oestrogens in relation to their environmental occurrence. *Critical Reviews in Toxicology*, 32, 113-132.
- Lateefa, A.K., Abdualah, Y.O., Najwa, A.A. and Mohamad, A.S. (2014). Adsorption behavior of oestrogenic compounds on carbon nanotubes from aqueous solutions: Kinetic and thermodynamic studies. *Journal of Industrial and Engineering Chemistry*, 20(3), 916–924.
- Lee, A.J., Cai, M.X., Thomas, P.E., Conney, A.H. and Zhu, B.T. (2003). Characterization of the oxidative metabolites of 17 β -estradiol and estrone formed by 15 selectively expressed human cytochrome p450 isoforms. *Endocrinology*, 144(8), 3382–3398.
- Li, M. and Scott, R.Y. (2017). Degradation and metabolite formation of oestrogen conjugates in an agricultural soil. *Journal of Pharmaceutical and Biomedical Analysis*, 145(13), 634-640.
- Liu, M., Edward, P., Lai, C. and Yu, Y. (2012). Removal of 17 β -estradiol by nylon filter membrane: adsorption kinetics and thermodynamics. *Journal of Recent Research and Applied Studies*, 11(1), 67-73.
- Liu, S. (2004). Transformation of MCF-10A human breast epithelial cells by zeranol and estradiol-17 β . *Journal of Breast*, 10(6), 514-21.
- Mackay, D. and Arnot, J.A. (2011). The application of fugacity and activity to simulating the environmental fate of organic contaminants. *Journal of Chemical & Engineering Data*, 56(4), 1348–1355.

- Marce, R.M. and Borrul, F. (2000). Review solid-phase extraction of polycyclic aromatic compounds. *Journal of Chromatography A*, 885(3), 273–290.
- María, J.B., Jimena, L., Lilián, E.C., Néstor, W., Soria, B. and García, A. (2016). Dynamics of expression of two vitellogenin genes in the Chagas' disease vector *Triatoma infestans*: Analysis throughout pre-vitellogenesis and vitellogenesis. *Acta Tropica*, 47(7), 547-553.
- Maria, N., Jennifer, D., Christel, C. and Anna, W. (2017). Freshwater ecotoxicity impacts from pesticide use in animal and vegetable foods produced in Sweden. *Science of The Total Environment*, 581-582(1), 448-459.
- Marie, C., Valera, P., Gourdy, F., Trémollières, J. and François, A. (2015). From the women's health initiative to the combination of oestrogen and selective oestrogen receptor modulators to avoid progestin addition. *Maturitas*, 82(3), 274-277.
- Marjeta, Č. and Ester, H. (2017). Disk-based solid phase extraction for the determination of diclofenac and steroidal oestrogens E1, E2 and EE2 listed in the WFD watch list by GC–MS. *Science Total Environment*, 590(5), 832-837.
- Mark, F. (2014). A new wastewater treatment technology for developing countries. *Filtration and Separation*, 51(5), 14-17.
- Mastrup, M., Jensen, R.L., Schfäer, A.I., and Khan, S. (2001). Fate modeling—An important tool for water recycling. The membranes research environment. *Environment Science Technology*, 16(7), 171-178.
- Matějček, A., Matějčková, J., Němcová, E., Jandurová, O.M., Štípková, M., Bouška, J. and Frelich, J. (2007). Joint effects of CSN3 and LGB genotypes and their relation to breeding values of milk production parameters in Czech Fleckvieh. *Czech Journal of Animal Science*, 52(11), 83–87.
- Matsumura, N., Ishibashi, H., Hirano, M., Nagao, Y., Watanabe, N., Shiratsuchi, H. (2005). Effects of nonylphenol and triclosan on production of plasma vitellogenin and testosterone in male South African clawed frogs (*Xenopus laevis*). *Biological and Pharmaceutical Bulletin*, 28(9), 1748-1751.
- Matteo, D., Dharni, V., Joseph, L., and Chittaranjan, R. (2014). Fate and transport of selected oestrogen compounds in Hawaii soils: Effect of soil type and macropores. *Journal of Contaminant Hydrology*, 166(7), 1-10.
- Mawuli, A., Faisal, K., Brian, V. and Ming, Y. (2015). Dynamic fugacity model for accidental oil release during Arctic shipping. *Marine Pollution Bulletin*, 111(1–2), 347-353.
- MERCK'S Index. (1996). The Merck Index: 12th edition. *Occupational Environment Medicine*, 54(4), 288.

- Mikael, G., Jörgen, M., Bethanie, C., Almroth, M., Eriksson, K., Joachim, S. and Thomas, B. (2017). Chemical monitoring of Swedish coastal waters indicates common exceedances of environmental thresholds, both for individual substances as well as their mixtures. *Marine Pollution Bulletin*, 122(1–2), 409-419.
- Ming, H.W., Chen, J.Q., Gang, X., Yan, F.S., Jing, M., Hui, X., Rui, S. et al. (2016). Occurrence, fate and interrelation of selected antibiotics in sewage treatment plants and their receiving surface water. *Ecotoxicology and Environmental Safety*, 132(1), 132-139.
- Mohamed, A.T., Mohamed, S.N., Gamal, A.E., Ibrahim, A. and Ali. I. (2017). Auspicious role of the steroidal heterocyclic derivatives as a platform for anti-cancer drugs. *Bioorganic Chemistry*, 73(1), 128-146.
- Mohan, S.V., Shailaja, S., Rama, K.M. and Sarma, P.N. (2007). Adsorptive removal of phthalate ester (di-ethyl phthalate) from aqueous phase by activated carbon: A kinetic study. *Journal of Hazardous Materials*, 146(1-2), 278-282.
- Mostofa, A., Christina, Ø.P., Anita, F., Tamie, L.V. and Mette, L. (2016). Influence of soil structure on contaminant leaching from injected slurry. *Journal of Environmental Management*, 184(2), 289-296.
- Mostofa, A., Jirka, Š. and Mette, L. (2014). Simulation of the redistribution and fate of contaminants from soil-injected animal slurry. *Agricultural Water Management*, 131(1), 17-29.
- Muhammad, A., Xiaoming, S., Yuanyuan, W., Dennis, F. and Yuesuo, Y. (2017). Environmental impact of oestrogens on human, animal and plant life: A critical review. *Environment International*, 99(5), 107-119.
- Nadali, A., Monavvar, D., Mohammad, S., Gholamreza, G., Abdolkazem, N. and Ali, A. B. (2017). Investigating the efficiency of co-composting and vermicomposting of vinasse with the mixture of cow manure wastes, bagasse, and natural zeolite. *Waste Management*, 69(1), 117-126.
- Nadine, G., Ishai, D. and Brian, B. (2015). Fate and transport of free and conjugated oestrogens during soil passage. *Environmental Pollution*, 206(4), 80-87.
- Nai, X.F., Jiao, Y., Hai M.Z., Yu, T.C., Ce, H.M., Quan, Y.C., Yan, W.L. et al. (2017). Efficient phytoremediation of organic contaminants in soils using plant–endophyte partnerships. *Science Total Environment*, 583(7), 352-368.
- Nghiem, L.D., McCutcheon, J., Schafer, A.I., and Elimelech, M. (2004). The role of endocrine disrupters in water recycling: risk or mania? *Water Science and Technology*, 50(9), 215–220.
- Nicole, S., Kast, M.S., Danielle, S., Jérôme, L., Antonia, P., Patrick, O. and Konrad, H. (2015). Addressing the complexity of water chemistry in environmental fate modeling for engineered nanoparticles. *Science Total Environment*, 535(15), 150-159.

- Noppe, T.V., Wulf, E.D., Verheyden, K., Monteyne, E. and Caeter, P.V. (2007). Occurrence of oestrogens in the Scheldt estuary: A 2- year survey. *Ecotoxicology and Environmental Safety*, 66(7), 1–8.
- NWHN. (2015). Herbs and Phytoestrogens. *Environment Science Technology*, 22(1), 377-383.
- OECD. (2004). Partition Coefficient (n-octanol/water), High Performance Liquid Chromatography (HPLC) Method.
- Plazinski, W., Dziuba, J. and Rudzinski, W. (2013). Modeling of sorption kinetics: The pseudo-second order equation and the sorbate intraparticle diffusivity. *Adsorption*, 19(5), 1055-1064.
- Rajasulochana, P., and Preethy, V. (2016). Comparison on efficiency of various techniques in treatment of waste and sewage water. *Resource Efficient Technologies*, 2(4), 175-184.
- Rajesh, K., Gaurav. H., Ashok, K.M., Abuzar, K. and Kenneth, G.F. (2014). Efficient analysis of selected oestrogens using fabric phase sorptive extraction and high performance liquid chromatography-fluorescence detection. *Journal of Chromatography A*, 1359(1), 16-25.
- Richard, J., Santen, W., Yue, J. and Wang, P. (2015). Oestrogen metabolites and breast cancer. *Steroids*, 99(6), 61-66.
- Ri-ping, H., Ze-hua, L., Su-fen, Y., Hua, Y., Zhi, D. and Ping-xiao, W. (2017). Worldwide human daily intakes of bisphenol A (BPA) estimated from global urinary concentration data (2000–2016) and its risk analysis. *Environmental Pollution*, 230(9), 143-152.
- Ritchie, T.J. and Macdonald, J.F. (2009). The impact of aromatic ring count on compound developability – are too many aromatic rings a liability in drug design?. *Drug Discovery Today*, 14(21–22), 1011-1020.
- Robert, J. H., and Michael, J.W. (2014). Gonadal steroid hormones and the hypothalamo–pituitary–adrenal axis. *Frontiers in Neuroendocrinology*, 35(2), 197-220.
- Robert, P.M.K. and Javier, A.T. (2016). Chapter 115 – Gonadotropin Releasing Hormones. *Endocrinology: Adult and Pediatric (Seventh Edition)*, 29(5), 2003–2022.
- Rong, Y., Na, L., Mei, M. and Zijian, W. (2014). Combined effects of oestrogenic chemicals with the same mode of action using an oestrogen receptor binding bioassay. *Environmental Toxicology and Pharmacology*, 3(8), 829–837.
- Ryohei, Y., Yoshinao, K., Satomi, K., Takeshi, M., Yukiko, O., Yasuhiko, O., Jan, M., et al. (2016). Characterization of evolutionary trend in squamate estrogen receptor sensitivity. *General and Comparative Endocrinology*, 238, 88-95.

- Saad, Y.J., Jayaprakash, S., Kavithaa, L., Oluwaseun, O. and Ogunbiyi, S. (2016). Reuse of treated sewage effluent (TSE) in Qatar. *Journal of Water Process Engineering*, 11(3), 174-182.
- Sabrina, G., Alberto, M., Andrea, C., Fabrizia, L., Sergio, F., and Giuseppe, P. (2017). Thyroid hormones in extreme longevity. *Mechanisms of Ageing and Development*, 165(1), 98-106.
- Samiullah, Y. (2010). Prediction of the environmental fate of chemicals. ISBN: 978-94-009-2211-2.
- Sarah, C., Virginie, B., Patrick, D., Nicolas, B., Patrick, B. and Guillermina, H.R. (2012). Fate of steroid hormones and endocrine activities in swine manure disposal and treatment facilities. *Water Research*, 46(3), 895-906.
- Sarah, H.D., Lisa, L.M., Welling, S. and Reeve, D. (2017). The influence of hormone replacement therapy on mating psychology among post-menopausal women. *Personality and Individual Differences*, 115(2), 13-18.
- Sarah, K.D., Corinne, J., Schuster, W., Manzoor, Q. and Katherine, P. (2016). A review of health risks and pathways for exposure to wastewater use in agriculture. *Environmental Health Perspective*, 124(7), 900-909.
- Sarmah, G.L., Northcott, F. and Scherr, F. (2008). Retention of oestrogenic steroid hormones by selected New Zealand soils. *Environment International*, 34(6), 749–755.
- Scherr, F.F., Ajit, K.S., Hong, J.D. and Keith, C.C. (2009). Degradation and metabolite formation of 17 β -estradiol-3-sulphate in New Zealand pasture soils. *Environment International*, 35(2), 291-297.
- Seema, P., Ahmad, H., Akondi, B.R., and Biswa, R. (2018). Estrogen: The necessary evil for human health, and ways to tame it. *Biomedicine & Pharmacotherapy*, 102, 403-411.
- Sergio, M.S., Spoel, E.R., and Spoel, D. (2014) Chemical properties, environmental fate, and degradation of seven classes of pollutants. *Chemical Research in Toxicology*, 27(5), 713-737.
- Shore, L.S. and Shemesh, M. (2003). Naturally produced steroid hormones and their release into the environment. *Pure and Applied Chemistry*, 75(11-12), 1859–1871.
- Shubhra, S., Riya, R.M., Sujaya, R. and Janardhana, R. (2017). Technology options for faecal sludge management in developing countries: Benefits and revenue from reuse. *Environmental Technology & Innovation*, 7(9), 203-218.
- Silva, C.P., Otero, M. and Esteves, V., (2012). Processes for the elimination of oestrogenic steroid hormones from water: A review. *Environmental Pollution*, 165(4), 38-58.

- Stanford, B.D. and Weinberg, H.S. (2007). Isotope dilution for quantitation of steroid oestrogens and nonylphenols by gas chromatography with tandem mass spectrometry in septic, soil, and groundwater matrices. *Journal of Chromatography A*, 1176(1-2), 26–36.
- Stephen, C.L., Watson, D.M., Paterson, A.M., Queirós, A.P., Rees, N.S. and Nicola, J.B. (2016). A conceptual framework for assessing the ecosystem service of waste remediation: In the marine environment. *Ecosystem Services*, 20(6), 69-81.
- Steven, P.G., Johnny, Y.N., Karensa, L.D., Andiliy, G.L. and Nicholas, D.S. (2015). Optimization of an indazole series of selective oestrogen receptor degraders: Tumor regression in a tamoxifen-resistant breast cancer xenograft. *Bioorganic & Medicinal Chemistry Letters*, 25(22), 5163-5167.
- Sun, J., Picht, E., Ginsburg, K.S., Bers, D.M., Steenbergen, C. and Murphy, E. (2006). Hypercontractile female hearts exhibit increased S-nitrosylation of the L-type Ca²⁺ channel α 1 subunit and reduced ischemia/reperfusion injury. *Science Total Environment*, 98(3), 403–411.
- Sun, Y., Huang, H., Wang, C., Shi, X., Hu, H., Kameya, T. and Fujie, K. (2014). Occurrence of oestrogenic endocrine disrupting chemicals concern in sewage plant effluent Front. *Environmental Science and Engineering*, 8(6), 18–26.
- Tatsanee, P., Kittima, T., Suphakde, J., Puttachart, P., Sasiprapa, T. and Suparat, W. (2016). Successful derivation of xeno-free mesenchymal stem cell lines from endometrium of infertile women. *Reproductive Biology*, 134(4), 1991-1998.
- Tedros, M.B., Jonathan, L., Mark, P.S., Krekeler, N. and Danielson, D. (2016). Adsorption of bisphenol A and ciprofloxacin by palygorskite-montmorillonite: Effect of granule size, solution chemistry and temperature. *Applied Clay Science*, 132(1), 518-527.
- Ternes, T.A., Meisenheimer, M., McDowell, D., Sacher, F., Brauch, H.J., Haist, G.B., Preuss, G. et al. (2002) Removal of pharmaceuticals during drinking water treatment. *Environmental of Science and Technology*, 36(17), 3855–3863.
- Tiffany, L., Torralba, S., Dave, T.F., Kuo, H.E., Allen, D.M. and Di, T. (2017). Bioconcentration factors and plant–water partition coefficients of munitions compounds in barley. *Chemosphere*, 189(1), 538-546.
- Timothy, J.E. (2017). Chapter 58 – Endocrine Disruption. *Reproductive and Developmental Toxicology (Second Edition)*, 32(8), 1091–1110.
- Ting, R., Dong, L., Shanjun, S., Maoyong, S., Hailin, W. and Guibin, J. (2015). Evaluation of the *in vitro* oestrogenicity of emerging bisphenol analogs and their respective oestrogenic contributions in municipal sewage sludge in China. *Chemosphere*, 124(6), 150-155.

- Ting, Y.F., Sarva, M.P., Clairede, B., Ahmad, Z.A., Sharifah, N.S.I. and Irniza, R. (2016). Analytical techniques for steroid oestrogens in water samples - A review. *Chemosphere*, 165(8), 358-368.
- Tizaoui, S., Ben. F. and Monser, L. (2017). Polyamide-6 for the removal and recovery of the oestrogenic endocrine disruptors estrone, 17 β -estradiol, 17 α -ethinylestradiol and the oxidation product 2-hydroxyestradiol in water. *Chemical Engineering Journal*, 328(15), 98-105.
- Tomohiro, K., Takumi, O., Souichiro, K., Manabu, S., Akifumi, O. and Kiminori, O. (2015). Oestrogenic activity of bis (4-hydroxyphenyl)methanes with cyclic hydrophobic structure. *Bioorganic & Medicinal Chemistry*, 23(21), 6900-6911.
- Travis, C.C., and Arms A.D. (1988). Bioconcentration of organics in beef, milk, and vegetation. *Environmental of Science and Technology*, 22(3), 271–274.
- Urban, Š., Marija, S.D. and Matjaž, J. (2016). In vitro impact of bisphenols BPA, BPF, BPAF and 17 β -estradiol (E2) on human monocyte-derived dendritic cell generation, maturation and function. *International Immunopharmacology*, 34(2), 146-154.
- Vimal, K., Andrew, C.J., Norihide, N., Naoyuki, Y. and Hiroaki, T. (2012). De-conjugation behavior of conjugated oestrogens in the raw sewage, activated sludge and river water. *Journal of Hazardous Materials*, 227(6), 49-54.
- Vimal, K., Norihide, N., Makoto, Y., Naoyuki, Y. and Hiroaki, T. (2011). The arrival and discharge of conjugated oestrogens from a range of different sewage treatment plants in the UK. *Chemosphere*, 82(8), 1124-1128.
- Wallace, J. M. (2012). Bone. *Environment Science Technology*, 37(1), 5471-5478.
- Watshara, S., Sunanta, N., Saw S., Virapong, P., Maris, L., Wikberg, J.E. and Chanin, N. (2016). Extending proteochemometric modeling for unraveling the sorption behavior of compound–soil interaction. *Chemometrics and Intelligent Laboratory Systems*, 151(7), 219-227.
- Weiwei, B., Bing, Z., Xiangjuan, Y., Yu, Z., Min, Y. and Zhimin, Q. (2017). Transformation and fate of natural oestrogens and their conjugates in wastewater treatment plants: Influence of operational parameters and removal pathways. *Water Research*, 124(11), 244-250.
- Welshons, W.V., Nagel, S.C., and Saal, F.S. (2006). Large effects from small exposures. III. Endocrine mechanisms mediating effects of bisphenol A at levels of human exposure. *Endocrinology*, 147(22), 56–69.
- Wilson, R., Duarte, D. and Jones. K.C. (1996). Screening the environmental fate of organic contaminants in sewage sludges applied to agricultural soils: I. The potential for downward movement to groundwaters. *Science Total Environment*, 185(9), 45–57.

- Xiaohua, Y., Jingchuan, X., Hong, Y., Qian, W., Arjun, K.V., Rolf, U.H., and Kurunthachalam, K. (2015). Occurrence and estrogenic potency of eight bisphenol analogs in sewage sludge from the U.S. EPA targeted national sewage sludge survey. *Journal of Hazardous Materials*, 299(1), 733-739.
- Xiaolin, L., Wei, Z., and Walton, R.K., (2013). Occurrence and removal of pharmaceutical and hormone contaminants in rural wastewater treatment lagoons. *Science of The Total Environment*, 445(3), 22-28.
- Xuelian, B., Francis, X.M., Casey, H.H., Thomas, M., De, S., Peter, G.O. and Eakalak, K. (2015). Sorption and degradation of 17 β -estradiol-17-sulfate in sterilized soil-water systems. *Chemosphere*, 119(12), 1322-1328.
- Xuelian, Z., Yanxia, L., Bei, L., Wang, J. and Chenghong, F. (2014). The effects of estrone and 17 β -estradiol on microbial activity and bacterial diversity in an agricultural soil: Sulfamethoxazole as a co-pollutant. *Ecotoxicology and Environmental Safety*, 89(23), 582-589.
- Yafeng, N., Zhimin, Q., Weiwei, B. and Junxin, L. (2014). Removal of endocrine-disrupting chemicals and conventional pollutants in a continuous-operating activated sludge process integrated with ozonation for excess sludge reduction. *Chemosphere*, 56(9), 1752-1759.
- Yalkowsky, S.H. and Sujit, B. (1992). Aqueous solubility: Methods of estimation for organic compounds. *Marcel Dekker*, 89(12), 128-148.
- Yen, J.Y., Peng, W.W., Chen, H.S., Tai, L.L., Cheng, Y.L. and Chih, H.K. (2017). Oestrogen levels, emotion regulation, and emotional symptoms of women with premenstrual dysphoric disorder: The moderating effect of oestrogen receptor 1 α polymorphism. *Progress in Neuro-Psychopharmacology and Biological Psychiatry*, 13(6), 142-149.
- Ying, G.G. and Kookana, R.S. (2005). Sorption and degradation of oestrogen-like-endocrine disrupting chemicals in soil. *Environmental Toxicology and Chemistry*, 24(10), 2640-2645.
- Yong, Qi., Asce, S.M. and Tian, C.Z. (2015). Sorption and desorption of testosterone at environmentally relevant levels: Effects of aquatic conditions and soil particle size fractions. *Journal of Environmental Engineering*, 142(1), 4015-4045.
- Yong, X.R., Kazunori, N., Munehiro, N., Nobuo, C. and Osamu, N. (2007). A thermodynamic analysis on adsorption of oestrogens in activated sludge process. *Water Research*, 41(1), 2341-2348.
- Yongnian, G., Junfeng, G., Jing, W., Shuangshuang, W., Qin, L., Shuhua, Z., and Ya, Z. (2017). Estimating the biomass of unevenly distributed aquatic vegetation in a lake using the normalized water-adjusted vegetation index and scale transformation method. *Science of The Total Environment*, 601(1), 998-1007.

- Yu, Z.Q., Xiao, B.H., Huang, W.L. and Peng, P.A., (2004). Sorption of steroid oestrogens to soils and sediments. *Environmental Toxicology and Chemistry*, 23(3), 531–539.
- Yun, Y.Y. (2010). Degradation and transport pathways of steroid hormones from human and animal waste. *Science Total Environment*, 83(7), 376-383.
- Zhongqi, H.E., Paulo, H.P. and Heidi, M.W. (2016). Applied and environmental chemistry of animal manure: A Review. *Pedosphere*, 26(6), 779-81.