

PRODUCTION OF FERULIC ACID VIA FERULOYL
POLYSACCHARIDE HYDROLYSIS OF BANANA
STEM WASTE USING SOIL MIXED CULTURE

NURUL SHAREENA AQMAR BINTI MOHD SHARIF

DOCTOR OF PHILOSOPHY
(BIO-PROCESS ENGINEERING)

UNIVERSITI MALAYSIA PAHANG

SUPERVISOR'S DECLARATION

We hereby declare that we have checked this thesis and in our opinion, this thesis is adequate in terms of scope and quality for the award of the degree of Doctor of Philosophy (Bio-process Engineering).



ASSOC. PROF. IR. DR. NORAZWINA BT. ZAINOL
DEPUTY DEAN
ACADEMIC & STUDENT AFFAIR
COLLEGE OF ENGINEERING
UNIVERSITI MALAYSIA PAHANG
LEBUHRAYA TUN RAZAK, 26300 KUANTAN
TEL : +609-549 2690 FAX : +609-549 2689

Full Name : IR. DR NORAZWINA BINTI ZAINOL

Position : ASSOCIATE PROFESSOR

Date : 7 MAY 2021



Prof. Madya Dr. Wan Mohd Hafizuddin bin Wan Yussof
Profesor Madya
Fakulti Teknologi Kejuruteraan Kimia dan Proses
Universiti Malaysia Pahang

Full Name : DR WAN MOHD HAFIZUDDIN BIN WAN YUSSOF

Position : ASSOCIATE PROFESSOR

Date : 7 MAY 2021



STUDENT'S DECLARATION

I hereby declare that the work in this thesis is based on my original work except for quotations and citations which have been duly acknowledged. I also declare that it has not been previously or concurrently submitted for any other degree at Universiti Malaysia Pahang or any other institutions.

A handwritten signature in black ink, appearing to be 'Nurul Shareena Aqmar Binti Mohd Sharif', is positioned above a horizontal line.

Full name : NURUL SHAREENA AQMAR BINTI MOHD SHARIF
ID number : PKB14003
Date : 7 MAY 2021

PRODUCTION OF FERULIC ACID VIA FERULOYL POLYSACCHARIDE
HYDROLYSIS OF BANANA STEM WASTE USING SOIL MIXED CULTURE

NURUL SHAREENA AQMAR BINTI MOHD SHARIF

Thesis submitted in fulfilment of the requirements
for the award of the degree of
Doctor of Philosophy
(Bio-process Engineering)

College of Engineering
UNIVERSITI MALAYSIA PAHANG

MAY 2021

Dedicated to

My parents, for constantly praying
My supervisor, for endless understanding
My husband, for never once doubting
My children, for evermore loving
My siblings, for always motivating

And to myself

For getting back up and start believing

ACKNOWLEDGEMENTS

In the name of Allah, the Most Gracious and Most Merciful.

All praise to Allah, without Him, this thesis would never be completed. Thank you, Allah, for all the people surrounding me. Thank you, Allah, for all the blood, sweat, and tears, making me stronger. Thank you, Allah, for opening the path and clearing up the way. Thank you, Allah, for easing the journey with a shower of fortune and blessings. Allah the Most Powerful.

My utmost appreciation goes to my supervisor, Associate Professor Dr. Ir. Norazwina Binti Zainol for her guidance from the beginning until the end of this study. Not once had she failed to encourage, giving me invaluable advice to keep on going. Her persistence inspired me and hoping this thesis would make her proud.

This research would not have been possible without the support from the administrative, lecturers, and laboratory staff of Fakulti Kejuruteraan Kimia dan Sumber Asli (FKKSA). My gratitude also goes to fellow postgraduates, especially Kamaliah, Zulsyazwan, Siti Natrah, Abu Hannifa, Zulhelmi, and Mah Kah Hong for their support and effort in any way possible, from day 1. Not forgetting, undergraduates Ee Su Thor and Joanne Tey as great team members, completing the objectives of this study together, no matter the time.

To my husband, there's no amount of gratitude I can express, for holding on, pulling me up when I fell so low, and pushing me to the top. But most importantly, for always believing the way no one could have. You are truly God-sent, and I am forever blessed. To my loving children, I could never repay your unconditional love for always be there even when I'm down. May both of you grow to be better than I am today.

Last but not least, a heartfelt thank you goes to all my parents, my siblings, including my in-laws for the undying motivation and assistance along the way. Without it, this journey would be utterly meaningless and impossible. To my mother, all credit goes to you, every letters in every page, every lesson I've ever learnt, and every doors done unlocked. Without your discipline, your hard work, and sacrifices, I wouldn't be who I am now. And to my father, I dedicate the title to you, for you are the reason I grew up decent, and you deserve all that I've achieved and more, Dr. M. Sharif.

ABSTRAK

Sumber lignoselulosa sangat mudah diperolehi daripada sisa-sisa pertanian terutamanya dari sisa tanaman dan sering dibiarkan mereput di kebun. Sisa tanaman ini bukan hanya boleh digunakan untuk menghasilkan kompaun organik, malah boleh juga digunakan sebagai sumber enzim. Kajian ini menggunakan kultur campuran tanah (SMC) bagi hidrolisis enzimatik *feruloyl-polysaccharide* daripada sisa batang pisang (BSW) untuk menghasilkan asid ferulik (FA). Kaedah analisis multivarian seperti model faktorial penuh (FFD) dan model komposit tengah (CCD) digunakan untuk mengkaji dan mengoptimumkan kesan dan interaksi antara lima faktor iaitu suhu fermentasi (A; 26 – 40 °C), pergolakan (B; 0 – 150 rpm), nisbah air kepada BSW (C; 1 – 2 v/v), nisbah substrat kepada inokulum (D; 1 – 2 v/v), dan masa inkubasi (E; 24 – 120 jam). Kemudian, kajian kinetik dijalankan berdasarkan model *Michaelis-Menten* untuk memerhati tindak balas proses. Akhir sekali, lima bakteria dominan dari SMC dikenalpasti dan keupayaan mereka dalam penghasilan FA dibandingkan. Eksperimen fermentasi secara berkelompok dijalankan sebanyak tiga kali berdasarkan nilai parameter yang diperolehi daripada model. Model garis lurus lekup 2^5 FFD didapati melengkapinya nilai korelasi $R^2 = 0.8019$ dengan impak faktor pada kedudukan $E > C > A > D > B$. Faktor E menyumbang 27.37% impak menunjukkan kepentingan aktiviti pertumbuhan sel, manakala interaksi DE adalah yang tertinggi membuktikan kepentingan masa dalam penggunaan substrat bagi penghasilan FA. Faktor E dan C telah dipilih sebagai pembolehubah untuk 5^2 CCD berdasarkan kaedah gerak-balas permukaan (RSM) dalam julat nilai diantara 12 sehingga 36 jam dan nisbah 0.5:1 sehingga 1.5:1. R^2 model kuadratik didapati melengkapinya nilai 0.8068. Seperti yang dijangkakan, interaksi antara dua faktor ini membuktikan bahawa kesan masa lebih tinggi berbanding nisbah air kepada BSW. Hasil maksimum FA pada kondisi optimum 27 jam masa inkubasi dan 1.1:1 nisbah air kepada BSW adalah 1.1657 mg FA/g BSW. Dengan nilai ini, eksperimen baharu dijalankan dengan memanjangkan masa inkubasi sehingga 60 jam dimana hasil FA diperhatikan pada setiap 6 jam untuk membentuk model kinetik *Michaelis-Menten*. Pemalar kinetik K_s (pemalar kadar tindak balas), K_m (pemalar *Michaelis-Menten*) dan V_{max} (kadar halaju maksimum) dari persamaan tindak balas kadar pertama dan model *Michaelis-Menten* telah dikenalpasti melalui jumlah ralat kuasa dua diantara data eksperimen dan teori menggunakan *Excel Solver*. Kepekatan awal biomas [S_0] dan hasil FA [P_0] adalah 7.4150 g biomas/L dan 1.3710 mg FA/g BSW. Nilai pemalar kinetik adalah $K_s = 0.0053 \text{ h}^{-1}$; $V_{max} = 4.1200 \times 10^{-5} \text{ } \mu\text{mol/min}$; $K_m = 0.0500 \text{ mmol/L}$. Nilai R^2 yang diperolehi adalah lebih dari 0.800 membuktikan kestabilan sistem. Kemudian, lima bakteria dominan telah dikenalpasti dan keupayaan masing-masing dalam penghasilan FA diperhatikan dimana: *Brevundimonas nasdae* s. W1-2B = 0.4535 mg FA/g BSW; *Pseudomonas monteilii* s. CIP 104883 (B) = 0.7919 mg FA/g BSW; *Pseudomonas monteilii* s. CIP 104883 (C) = 0.8302 mg FA/g BSW; *Lysinibacillus boronitolerans* s. 10a = 0.8249 mg FA/g BSW; dan *Bacillus anthracis* s. Ames = 0.8383 mg FA/g BSW. Walaubagaimanapun, hasil FA yang tertinggi dihasilkan dengan menggunakan SMC adalah pada 1.4597 mg FA/g BSW. Oleh itu, dapat disimpulkan bahawa BSW terbukti sangat berguna dan sesuai digunakan untuk menghasilkan produk seperti FA. Tambahan lagi, SMC dapat menjana enzim untuk hidrolisis BSW dalam penghasilan FA dengan lebih efisien.

ABSTRACT

The source of lignocellulose is abundantly available through agricultural wastes and often discarded in the landfill. These wastes are not just potentially produce organic compound, but also could be the source to generate crude enzyme. This study proposed the used of soil mixed culture (SMC) for feruloyl-polysaccharide hydrolysis from banana stem waste (BSW) to produce ferulic acid (FA). Method used was comprised of multivariate analysis through full factorial design (FFD) and central composite design (CCD) to investigate and optimize the effect and interaction of five factors; fermentation temperature (A; 26 – 40 °C), agitation (B; 0 – 150 rpm), water-to-BSW ratio (C; 1 – 2 v/v), substrate-to-inoculums ratio (D; 1 – 2 v/v), and incubation time (E; 24 – 120 h). Then, a study of kinetic modelling was performed based on Michaelis-Menten model to observe the process behaviour. Lastly, five most dominant bacteria from SMC were identified and later be compared on their ability to produce FA. Series of batch fermentation were conducted at triplicate according to the parameter values of experimental runs generated by the design. A 2^5 linear model of FFD was well fitted at $R^2=0.8019$ with factors contribution in the order of $E > C > A > D > B$. Factor E had 27.37% contribution with an optimum range at 12 – 36 h, indicating the significance of cell growth activities, while interaction of DE was highest revealing the importance of sufficient time for substrate utilization to get high FA yield. Parameter E and C were selected as the variables for a 5^2 CCD based on response surface methodology (RSM) at values range of 12 to 36 h and 0.5:1 to 1.5:1 v/v, respectively. The R^2 value for the quadratic model was fitted at 0.8068. Interaction between these factors revealed that effect of time was greater than ratio of water-to-BSW, as expected. The maximum FA produced was 1.1657 mg FA/g BSW at the optimum condition of 27 h incubation time and 1.1:1 water to BSW ratio. With this value, experiments were conducted at a prolonged of 60 h incubation time, where FA yield was observed at every 6 h time interval to develop the study of Michaelis-Menten kinetic model. The kinetic constant, K_s (reaction rate coefficient), K_m (Michaelis-Menten constant) and V_{max} (maximum forward velocity) from first-order reaction equation and Michaelis –Menten model were determined based on the sum squared error between experimental data and theoretical data by using Excel Solver. The initial concentration of biomass [S_0] and FA yield [P_0] were 7.4150 g biomass/L and 1.3710 mg FA/g BSW, respectively. The values of calculated kinetic parameters were: $K_s = 0.0053 \text{ h}^{-1}$; $V_{max} = 4.1200 \times 10^{-5} \text{ } \mu\text{mol/min}$; $K_m = 0.0500 \text{ mmol/L}$. The R^2 values obtained from squared error calculation were satisfactorily more than 0.800 which verified the stability of the system. Then, five most stand out pure strains were identified and their performance on FA yield were observed as follows: *Brevundimonas nasdae* s. W1-2B = 0.4535 mg FA/g BSW; *Pseudomonas monteilii* s. CIP 104883 (B) = 0.7919 mg FA/g BSW; *Pseudomonas monteilii* s. CIP 104883 (C) = 0.8302 mg FA/g BSW; *Lysinibacillus boronitolerans* s. 10a = 0.8249 mg FA/g BSW; and *Bacillus anthracis* s. Ames = 0.8383 mg FA/g BSW. Meanwhile, using SMC, highest FA yield was 1.4597 mg FA/g BSW. It can be concluded that, BSW was proven to be useful and highly feasible for producing FA, which is useful in pharmaceutical industry. Furthermore, using SMC as the source of inoculum to generate enzymes for hydrolysis of BSW was potentially abled to produce FA as efficiently.

TABLE OF CONTENT

DECLARATION	
TITLE PAGE	
DEDICATION	
ACKNOWLEDGEMENTS	ii
ABSTRAK	iii
ABSTRACT	iv
TABLE OF CONTENT	v
LIST OF TABLES	x
LIST OF FIGURES	xi
LIST OF SYMBOLS	xiii
LIST OF ABBREVIATIONS	xiv
CHAPTER 1 INTRODUCTION	1
1.1 Introduction	1
1.2 Background of Study	2
1.3 Problem Statement	5
1.4 Objectives of Study	6
1.5 Scopes of Study	6
1.6 Contribution of Study	8
1.7 Thesis Outline	9
CHAPTER 2 LITERATURE REVIEW	10
2.1 Introduction	10
2.1.1 Lignocellulosic Wastes	11
2.2 Banana Stem Waste as Substrate	13

2.3	Ferulic Acid	18
	2.3.1 Application of Ferulic Acid	19
2.4	Technique to Produce Ferulic Acid	21
	2.4.1 Feruloyl Polysaccharide Hydrolysis	26
2.5	Soil Mixed Culture as Ferulic Acid Producer	27
	2.5.1 Soil Microorganisms	29
2.6	Factors Affecting Ferulic Acid Production	32
	2.6.1 Effect of Temperature	35
	2.6.2 Effect of Agitation	36
	2.6.3 Effect of Water Content	36
	2.6.4 Effect of Substrate Amount	37
	2.6.5 Effect of Time	38
2.7	Optimization of Ferulic Acid Production	39
	2.7.1 Factorial Analysis	41
	2.7.2 Response Surface Methodology	43
2.8	Kinetic Study	45
	2.8.1 Michaelis-Menten	47
	2.8.2 Runge-Kutta Method	48
2.9	Mechanism Behaviour of Feruloyl Polysaccharide Hydrolysis	51
2.10	Chapter Summary	54
	CHAPTER 3 METHODOLOGY	56
3.1	Introduction	56
3.2	Banana Stem Waste	58
	3.2.1 BSW Characterization	58
	3.2.2 Substrate Preparation	58

3.3	Soil Mixed Culture	59
	3.3.1 Soil Characterization	59
	3.3.2 Inoculum Acclimatization	60
3.4	Experimental Design	60
	3.4.1 Factors Screening	62
	3.4.2 Optimum Parameters	63
	3.4.3 Data Validation	64
3.5	Kinetic Modelling	64
	3.5.1 Batch Fermentation	64
	3.5.2 Kinetic Coefficients	65
3.6	Analysis	66
	3.6.1 Ferulic Acid	66
	3.6.2 Biomass	67
	3.6.3 Feruloyl Esterase Assay	67
3.7	Strain Isolation	67
	3.7.1 Nutrient Agar Preparation	68
	3.7.2 Aseptic Serial Dilution	68
	3.7.3 Streaking Method	69
	3.7.4 Storage and Revival Technique	69
	3.7.5 Morphology Observation	70
3.8	Strain Identification	70
	3.8.1 MicroPlate System Measurement	70
	3.8.2 DNA Extraction	71
	3.8.3 PCR Amplification	71
3.9	Strain Performance	72

3.10	Chapter Summary	72
CHAPTER 4 RESULTS AND DISCUSSION		74
4.1	Introduction	74
4.2	Characterization	75
4.2.1	BSW Characteristics	75
4.2.2	Soil Characteristics	76
4.3	Factors Affecting FA Production	77
4.3.1	Contribution of Factors	77
4.3.2	Analysis of Variance for FFD	81
4.3.3	Interaction of Factors	83
4.3.4	Selection of Significant Factors	84
4.4	Optimization of FA Production	84
4.4.1	Analysis of Variance for CCD	85
4.4.2	Response Surface Plot	87
4.4.3	Optimum Point Selection and Validation	89
4.4.4	Comprehensive Review	90
4.5	Kinetic Study	91
4.5.1	Estimation of Kinetic Coefficients	92
4.5.2	Comprehensive Review	92
4.6	Mechanism Behaviour	95
4.6.1	Biomass Residues	95
4.6.2	FAE Activity	97
4.6.3	FA Production	98
4.7	Isolation of Soil Culture	99
4.8	Identification of Isolated Strains	101

4.8.1	Biochemical Test	101
4.8.2	Biomolecular Test	103
4.8.3	Strain Performance	106
4.9	Chapter Summary	107
CHAPTER 5 CONCLUSIONS		109
5.1	Introduction	109
5.2	Conclusions	109
5.3	Future Study Recommendations	111
REFERENCES		112
APPENDIX A EXPERIMENTAL PROCEDURES		134
APPENDIX B DATA AND FINDINGS		142
APPENDIX C LIST OF PUBLICATION		160

REFERENCES

- Abbasiliasi, S., Tan, J. S., Tengku Ibrahim, T. A., Bashokouh, F., Ramakrishnan, N. R., Mustafa, S., & Ariff, A. B. (2017). Fermentation factors influencing the production of bacteriocins by lactic acid bacteria: A review. *RSC Advances*, 7(47), 29395–29420. <https://doi.org/10.1039/c6ra24579j>
- Abdel-Latif, A., & Osman, G. (2017). Comparison of three genomic DNA extraction methods to obtain high DNA quality from maize. *Plant Methods*, 13(1), 1–9. <https://doi.org/10.1186/s13007-016-0152-4>
- Abdul Samad, K. (2017). *Synergistic Ferulic Acid Production From Banana Stem Waste By Co-Culture*.
- Abdullah, N., Sulaiman, F., & Taib, R. M. (2014). Characterization of banana (*Musa* spp.) plantation wastes as a potential renewable energy source. *AIP Conference Proceedings*, 1528(8), 325–330. <https://doi.org/10.1063/1.4803618>
- Abdullah, N., Sulaiman, F., Taib, R. M., & Miskam, M. A. (2015). Pyrolytic oil of banana (*Musa* spp.) pseudo-stem via fast process. *AIP Conference Proceedings*, 1657. <https://doi.org/10.1063/1.4915212>
- Abokitse, K., Wu, M., Bergeron, H., Grosse, S., & Lau, P. C. K. (2010). Thermostable feruloyl esterase for the bioproduction of ferulic acid from triticale bran. *Applied Microbiology and Biotechnology*, 87(1), 195–203. <https://doi.org/10.1007/s00253-010-2441-6>
- Abouelenien, F., Namba, Y., Kosseva, M. R., Nishio, N., & Nakashimada, Y. (2014). Enhancement of methane production from co-digestion of chicken manure with agricultural wastes. *Bioresource Technology*, 159, 80–87. <https://doi.org/10.1016/j.biortech.2014.02.050>
- Adegoke, K. A., & Bello, O. S. (2015). Dye sequestration using agricultural wastes as adsorbents. *Water Resources and Industry*, 12, 8–24. <https://doi.org/10.1016/j.wri.2015.09.002>
- Agrawal, R., Satlewal, A., Gaur, R., Mathur, A., Kumar, R., Gupta, R. P., & Tuli, D. K. (2015). Pilot scale pretreatment of wheat straw and comparative evaluation of commercial enzyme preparations for biomass saccharification and fermentation. *Biochemical Engineering Journal*, 102, 54–61. <https://doi.org/10.1016/j.bej.2015.02.018>
- AgriInfo.in. (2020). Soil Microbiology. Retrieved from <https://agriinfo.in/>
- Aguirre-Monroy, A. M., Santana-Martínez, J. C., & Dussán, J. (2019). *Lysinibacillus sphaericus* as a nutrient enhancer during fire-impacted soil replantation. *Applied and Environmental Soil Science*, 2019. <https://doi.org/10.1155/2019/3075153>
- Ahmed, I., Yokota, A., Yamazoe, A., & Fujiwara, T. (2007). Proposal of *Lysinibacillus boronitolerans* gen. nov. sp. nov., and transfer of *Bacillus fusiformis* to *Lysinibacillus fusiformis* comb. nov. and *Bacillus sphaericus* to *Lysinibacillus sphaericus* comb. nov. *International Journal of Systematic and Evolutionary Microbiology*, 57(5), 1117–1125. <https://doi.org/10.1099/ijs.0.63867-0>
- Akpinar, O., Erdogan, K., & Bostanci, S. (2009). Production of xylooligosaccharides by

- controlled acid hydrolysis of lignocellulosic materials. *Carbohydrate Research*, 344(5), 660–666. <https://doi.org/10.1016/j.carres.2009.01.015>
- Al-Zuhair, S., Hasan, M., & Ramachandran, K. B. (2003). Kinetics of the enzymatic hydrolysis of palm oil by lipase. *Process Biochemistry*, 38(8), 1155–1163. [https://doi.org/10.1016/S0032-9592\(02\)00279-0](https://doi.org/10.1016/S0032-9592(02)00279-0)
- Alias, L. M., Manoharan, S., Vellaichamy, L., Balakrishnan, S., & Ramachandran, C. R. (2009). Protective effect of ferulic acid on 7,12-dimethylbenz[a]anthracene-induced skin carcinogenesis in Swiss albino mice. *Experimental and Toxicologic Pathology*, 61(3), 205–214. <https://doi.org/10.1016/j.etp.2008.09.001>
- Altemimi, A., Lakhssassi, N., Baharlouei, A., Watson, D. G., & Lightfoot, D. A. (2017). Phytochemicals: Extraction, isolation, and identification of bioactive compounds from plant extracts. *Plants*, 6(4). <https://doi.org/10.3390/plants6040042>
- Álvarez, C., Reyes-Sosa, F. M., & Díez, B. (2016). Enzymatic hydrolysis of biomass from wood. *Microbial Biotechnology*, 9(2), 149–156. <https://doi.org/10.1111/1751-7915.12346>
- Amado, I. R., Franco, D., Sánchez, M., Zapata, C., & Vázquez, J. A. (2014). Optimisation of antioxidant extraction from *Solanum tuberosum* potato peel waste by surface response methodology. *Food Chemistry*, 165, 290–299. <https://doi.org/10.1016/j.foodchem.2014.05.103>
- Ana Plazonić, Bucar, F., Maleš, Željko, Mornar, A., Nigović, B., & Kujundžić, N. (2009). Identification and quantification of flavonoids and phenolic acids in burr parsley (*caucalis platycarpus* L.), using high-performance liquid chromatography with diode array detection and electrospray ionization mass spectrometry. *Molecules*, 14(7), 2466–2490. <https://doi.org/10.3390/molecules14072466>
- Arutchelvan, V., Kanakasabai, V., Elangovan, R., Nagarajan, S., & Muralikrishnan, V. (2006). Kinetics of high strength phenol degradation using *Bacillus brevis*. *Journal of Hazardous Materials*, 129(1–3), 216–222. <https://doi.org/10.1016/j.jhazmat.2005.08.040>
- Ascher, U. M., & Petzold, L. R. (1998). *Computer Methods for Ordinary Differential Equations and Differential-Algebraic Equations*. Society for Industrial and Applied Mathematics.
- Aybastier, Ö., Işık, E., Şahin, S., & Demir, C. (2013). Optimization of ultrasonic-assisted extraction of antioxidant compounds from blackberry leaves using response surface methodology. *Industrial Crops and Products*, 44, 558–565. <https://doi.org/10.1016/j.indcrop.2012.09.022>
- Azmir, J., Zaidul, I. S. M., Rahman, M. M., Sharif, K. M., Mohamed, A., Sahena, F., ... Omar, A. K. M. (2013). Techniques for extraction of bioactive compounds from plant materials: A review. *Journal of Food Engineering*, 117(4), 426–436. <https://doi.org/10.1016/j.jfoodeng.2013.01.014>
- Baharin, A., Fattah, N. A., Bakar, A. A., & Ariff, Z. M. (2016). Production of Laminated Natural Fibre Board from Banana Tree Wastes. *Procedia Chemistry*, 19, 999–1006. <https://doi.org/10.1016/j.proche.2016.03.149>
- Baiea, M. H. M., EL-Gioushy, S. ., & El-Sharony, T. F. (2015). Effect of Feldspar and

Bio-Fertilization on Growth, Productivity and Fruit Quality of Banana cv.
International Journal of Environment, (November).

- Bajaj, M., Gallert, C., & Winter, J. (2009). Phenol degradation kinetics of an aerobic mixed culture. *Biochemical Engineering Journal*, 46(2), 205–209. <https://doi.org/10.1016/j.bej.2009.05.021>
- Ballus, C. A., Meinhart, A. D., Bruns, R. E., & Godoy, H. T. (2011). Use of multivariate statistical techniques to optimize the simultaneous separation of 13 phenolic compounds from extra-virgin olive oil by capillary electrophoresis. *Talanta*, 83(4), 1181–1187. <https://doi.org/10.1016/j.talanta.2010.07.013>
- Ballus, C. A., Meinhart, A. D., De Souza Campos, F. A., Bruns, R. E., & Godoy, H. T. (2014). Doehlert design-desirability function multi-criteria optimal separation of 17 phenolic compounds from extra-virgin olive oil by capillary zone electrophoresis. *Food Chemistry*, 146, 558–568. <https://doi.org/10.1016/j.foodchem.2013.09.102>
- Barberousse, H., Kamoun, A., Chaabouni, M., Giet, J. M., Roiseux, O., Paquot, M., ... Blecker, C. (2009). Optimization of enzymatic extraction of ferulic acid from wheat bran, using response surface methodology, and characterization of the resulting fractions. *Journal of the Science of Food and Agriculture*, 89(10), 1634–1641. <https://doi.org/10.1002/jsfa.3630>
- Barros, I., Mendes, S., & Rosa, D. (2016). *Vibrio diabolicus* Immunomodulatory Effects on *Bathymodiolus azoricus* During Long-term Acclimatization at Atmospheric Pressure. *Journal of Aquaculture Research & Development*, 7(12). <https://doi.org/10.4172/2155-9546.1000464>
- Bartholomew, W. H., Karow, E. O., Sfat, M. R., & Wilhelm, R. H. (1950). Oxygen Transfer and Agitation in Submerged Fermentations. Effect of Air Flow and Agitation Rates upon Fermentation of *penicillium chrysogenum* and *Streptomyces griseus*. *Industrial & Engineering Chemistry*, 42(9), 1810–1815. <https://doi.org/10.1021/ie50489a033>
- Bartolomé, B., Faulds, C. B., Kroon, P. A., Waldron, K., Gilbert, H. J., Hazlewood, G., & Williamson, G. (1997). An *Aspergillus niger* esterase (ferulic acid esterase III) and a recombinant *Pseudomonas fluorescens* subsp. *cellulosa* esterase (Xy1D) release a 5-5' ferulic dehydrodimer (diferulic acid) from barley and wheat cell walls. *Appl. Environ. Microbiol.*, 63(1), 208–212.
- Baş, D., & Boyacı, I. H. (2007). Modeling and optimization i: Usability of response surface methodology. *Journal of Food Engineering*, 78(3), 836–845. <https://doi.org/10.1016/j.jfoodeng.2005.11.024>
- Béguin, P., & Aubert, J.-P. (1994). The biological degradation of cellulose. *FEMS Microbiology Reviews*, 13(1), 25–58. <https://doi.org/10.1111/j.1574-6976.1994.tb00033.x>
- Bell, K. S., Philp, J. C., Aw, D. W. J., & Christofi, N. (1998). A review: The genus *Rhodococcus*. *Journal of Applied Microbiology*, 85(2), 195–210. <https://doi.org/10.1046/j.1365-2672.1998.00525.x>
- Beyenal, H., & Babauta, J. T. (2015). *Biofilms in Bioelectrochemical Systems: From Laboratory Practice to Data Interpretation*. John Wiley & Sons Inc.

- Bilba, K., Arsene, M. A., & Ouensanga, A. (2007). Study of banana and coconut fibers. Botanical composition, thermal degradation and textural observations. *Bioresource Technology*, 98(1), 58–68. <https://doi.org/10.1016/j.biortech.2005.11.030>
- Bonifácio-Lopes, T., Teixeira, J. A., & Pintado, M. (2019). Current extraction techniques towards bioactive compounds from brewer's spent grain—A review. *Critical Reviews in Food Science and Nutrition*, 0(0), 1–12. <https://doi.org/10.1080/10408398.2019.1655632>
- Bonnina, E., Brunel, M., Gouy, Y., Lesage-Meessen, L., Asther, M., & Thibault, J. F. (2001). *Aspergillus niger* I-1472 and *Pycnoporus cinnabarinus* MUCL39533, selected for the biotransformation of ferulic acid to vanillin, are also able to produce cell wall polysaccharide-degrading enzymes and feruloyl esterases. *Enzyme and Microbial Technology*, 28(1), 70–80. [https://doi.org/10.1016/S0141-0229\(00\)00277-5](https://doi.org/10.1016/S0141-0229(00)00277-5)
- Boone, D. R., Castleholz, R. W., Brenner, D. J., Garrity, G. M., Krieg, N. R., & Staley, J. T. (2005). *Bergey's Manual of Systematic Bacteriology* (Vol. 2).
- Box, G. E. P., & Wilson, K. B. (1951). On the Experimental Attainment of Optimum Conditions. *Journal of the Royal Statistical Society: Series B (Methodological)*, 13(1), 1–38. <https://doi.org/10.1111/j.2517-6161.1951.tb00067.x>
- Briggs, G. E., & Haldane, J. B. S. (1925). A Note on the Kinetics of Enzyme Action. *Biochemical Journal*, 19(2), 338.
- Brown, K., & Lemon, J. (2021). Cations and Cation Exchange Capacity. Retrieved from soilquality.org.au website: <http://www.soilquality.org.au/factsheets/cation-exchange-capacity>
- Burrage, K., & Butcher, J. C. (1979). Stability Criteria for Implicit Runge-Kutta Methods. *SIAM Journal on Numerical Analysis*, 16(1), 46–57.
- Cai, Y., Luo, Q., Sun, M., & Corke, H. (2004). Antioxidant activity and phenolic compounds of 112 traditional Chinese medicinal plants associated with anticancer. *Life Sciences*, 74(17), 2157–2184. <https://doi.org/10.1016/j.lfs.2003.09.047>
- Canavar, P. E., Ekşin, E., & Erdem, A. (2015). Electrochemical monitoring of the interaction between mitomycin C and DNA at chitosan-carbon nanotube composite modified electrodes. *Turkish Journal of Chemistry*, 39(1), 1–12. <https://doi.org/10.3906/kim-1402-11>
- Chen, H. (2014). Biotechnology of lignocellulose: Theory and practice. In *Biotechnology of Lignocellulose: Theory and Practice*. <https://doi.org/10.1007/978-94-007-6898-7>
- Chen, H. P., Chow, M., Liu, C. C., Lau, A., Liu, J., & Eltis, L. D. (2012). Vanillin catabolism in *Rhodococcus jostii* RHA1. *Applied and Environmental Microbiology*, 78(2), 586–588. <https://doi.org/10.1128/AEM.06876-11>
- Crepin, V. F., Faulds, C. B., & Connerton, I. F. (2004). Functional classification of the microbial feruloyl esterases. *Applied Microbiology and Biotechnology*, 63(6), 647–652. <https://doi.org/10.1007/s00253-003-1476-3>
- Das, P. K., Nag, D., Debnath, S., & Nayak, L. K. (2010). Machinery for extraction and traditional spinning of plant fibres. *Indian Journal of Traditional Knowledge*, 9(2), 386–393.

- Davies, M. P., De Biasi, V., & Perrett, D. (2004). Approaches to the rational design of molecularly imprinted polymers. *Analytica Chimica Acta*, 504(1), 7–14. [https://doi.org/10.1016/S0003-2670\(03\)00812-2](https://doi.org/10.1016/S0003-2670(03)00812-2)
- De Ascensao, A. R. F. D. C., & Dubery, I. A. (2003). Soluble and wall-bound phenolics and phenolic polymers in *Musa acuminata* roots exposed to elicitors from *Fusarium oxysporum* f.sp. *cubense*. *Phytochemistry*, 63(6), 679–686. [https://doi.org/10.1016/S0031-9422\(03\)00286-3](https://doi.org/10.1016/S0031-9422(03)00286-3)
- de O. Buanafina, M. M. (2009). Feruloylation in grasses: Current and future perspectives. *Molecular Plant*, 2(5), 861–872. <https://doi.org/10.1093/mp/ssp067>
- de Paiva, L. B., Goldbeck, R., dos Santos, W. D., & Squina, F. M. (2013). Ferulic acid and derivatives: Molecules with potential application in the pharmaceutical field. *Brazilian Journal of Pharmaceutical Sciences*, 49(3), 395–411. <https://doi.org/10.1590/S1984-82502013000300002>
- Dean, A. C. R., & Hinshelwood, S. C. N. (1966). *Growth, Function and Regulation in Bacterial Cells*. Oxford University Press.
- Deepa, B., Abraham, E., Cordeiro, N., Mozetic, M., Mathew, A. P., Oksman, K., ... Pothan, L. A. (2015). Utilization of various lignocellulosic biomass for the production of nanocellulose: a comparative study. *Cellulose*, 22(2), 1075–1090. <https://doi.org/10.1007/s10570-015-0554-x>
- Deng, Y. ., & Wang, S. . (2016). Synergistic growth in bacteria depends on substrate complexity. *J Microbiol*, 54(1), 23–30.
- Department of Agriculture Malaysia. (2019). *Statistik Tanaman (Sub-sektor Tanaman Makanan) 2019*.
- Department of Statistics Malaysia. (2019). *Press Release: Selected Agricultural Indicators, Malaysia, 2019*.
- Dey, T. B., Chakraborty, S., Jain, K. K., Sharma, A., & Kuhad, R. C. (2016). Antioxidant phenolics and their microbial production by submerged and solid state fermentation process: A review. *Trends in Food Science & Technology*, 53(July), 60–74.
- Dimitrios, B. (2006). Sources of natural phenolic antioxidants. *Trends in Food Science and Technology*, 17(9), 505–512. <https://doi.org/10.1016/j.tifs.2006.04.004>
- Dobberstein, D., & Bunzel, M. (2010). Separation and detection of cell wall-bound ferulic acid dehydrodimers and dehydrotrimers in cereals and other plant materials by reversed phase high-performance liquid chromatography with ultraviolet detection. *Journal of Agricultural and Food Chemistry*, 58(16), 8927–8935. <https://doi.org/10.1021/jf101514j>
- Dodić, J. M., Vučurović, D. G., Dodić, S. N., Grahovac, J. A., Popov, S. D., & Nedeljković, N. M. (2012). Kinetic modelling of batch ethanol production from sugar beet raw juice. *Applied Energy*, 99, 192–197. <https://doi.org/10.1016/j.apenergy.2012.05.016>
- Donaghy, J., Kelly, P. F., & McKay, A. M. (1998). Detection of ferulic acid esterase production by *Bacillus* spp. and lactobacilli. *Applied Microbiology and Biotechnology*, 50(2), 257–260. <https://doi.org/10.1007/s002530051286>

- Egizabal, A., Zuloaga, O., Etxebarria, N., Fernández, L. A., & Madariaga, J. M. (1998). Comparison of microwave-assisted extraction and Soxhlet extraction for phenols in soil samples using experimental designs. *Analyst*, *123*(8), 1679–1684. <https://doi.org/10.1039/a802117a>
- Elomari, M., Coroler, L., Verhille, S., Izard, D., & Leclerc, H. (1997). *Pseudomonas monteilii* sp. nov., Isolated from Clinical Specimens. *International Journal of Systematic Bacteriology*, *47*(3), 846–852.
- Faulds, C. B., Zanichelli, D., Crepin, V. F., Connerton, I. F., Juge, N., Bhat, M. K., & Waldron, K. W. (2003). Specificity of feruloyl esterases for water-extractable and water-unextractable feruloylated polysaccharides: Influence of xylanase. *Journal of Cereal Science*, *38*(3), 281–288. [https://doi.org/10.1016/S0733-5210\(03\)00029-8](https://doi.org/10.1016/S0733-5210(03)00029-8)
- Fazary, A. E., Ismadji, S., & Ju, Y. H. (2010). Studies on temperature dependent kinetics of *Aspergillus awamori* feruloyl esterase in water solutions. *Kinetics and Catalysis*, *51*(1), 31–37. <https://doi.org/10.1134/S0023158410010064>
- Ferreira, L. M. A., Wood, T. M., Williamson, G., Faulds, C., Hazlewood, G. P., Black, G. W., & Gilbert, H. J. (1993). A modular esterase from *Pseudomonas fluorescens* subsp. *cellulosa* contains a non-catalytic cellulose binding domain. *Biochemical Journal*, *294*(2), 349–355. <https://doi.org/10.1042/bj2940349>
- Ferreira, P., Diez, N., Faulds, C. B., Soliveri, J., & Copa-Patiño, J. L. (2007). Release of ferulic acid and feruloylated oligosaccharides from sugar beet pulp by *Streptomyces tendae*. *Bioresource Technology*, *98*(8), 1522–1528. <https://doi.org/10.1016/j.biortech.2006.06.004>
- Ferreira, S., Duarte, A. P., Ribeiro, M. H. L., Queiroz, J. A., & Domingues, F. C. (2009). Response surface optimization of enzymatic hydrolysis of *Cistus ladanifer* and *Cytisus striatus* for bioethanol production. *Biochemical Engineering Journal*, *45*(3), 192–200. <https://doi.org/10.1016/j.bej.2009.03.012>
- Ferreira, S. L. C., Silva Junior, M. M., Felix, C. S. A., da Silva, D. L. F., Santos, A. S., Santos Neto, J. H., ... Souza, A. S. (2017). Multivariate optimization techniques in food analysis – A review. *Food Chemistry*, *273*, 3–8. <https://doi.org/10.1016/j.foodchem.2017.11.114>
- Ferulic Acid Catalogue. (2009). *Food & Cosmetic Material*. 0–13.
- Fogler, H. S. (2016). *Elements of Chemical Reaction Engineering Fifth Edition*. Prentice Hall.
- Foster, L. J. R., Zervas, S. J., Lenz, R. W., & Fuller, R. C. (1995). The biodegradation of poly-3-hydroxyalkanoates, PHAs, with long alkyl substituents by *Pseudomonas maculicola*. *Biodegradation*, *6*(1), 67–73. <https://doi.org/10.1007/BF00702301>
- Friedrich, M. T., Martins, M. L., Prestes, O. D., & Zanella, R. (2016). Use of Factorial Design in the Development of Multiresidue Method for Determination of Pesticide Residues in Wheat by Liquid Chromatography-Tandem Mass Spectrometry. *Food Analytical Methods*, *9*(9), 2541–2551. <https://doi.org/10.1007/s12161-016-0447-0>
- Furukawa, H., Zenno, S., Iwasawa, Y., Morita, H., Yoshida, T., & Nagasawa, T. (2003). Ferulic Acid Production from Clove Oil by *Pseudomonas fluorescens* E118. *Journal of Bioscience and Bioengineering*, *96*(4), 404–405.

<https://doi.org/10.1263/jbb.96.404>

- Gao, X., Wang, S., Xu, Y., Li, H., Zhao, H., & Pan, X. (2018). Ferulic acid and PDMS modified medical carbon materials for artificial joint prosthesis. *PLoS ONE*, *13*(9), 1–14. <https://doi.org/10.1371/journal.pone.0203542>
- García-Conesa, M. T., Crepin, V. F., Goldson, A. J., Williamson, G., Cummings, N. J., Connerton, I. F., ... Kroon, P. A. (2004). The feruloyl esterase system of *Talaromyces stipitatus*: Production of three discrete feruloyl esterases, including a novel enzyme, TsFaeC, with a broad substrate specificity. *Journal of Biotechnology*, *108*(3), 227–241. <https://doi.org/10.1016/j.jbiotec.2003.12.003>
- García-Pérez, J. A., Alarcón-Gutiérrez, E., Perroni, Y., & Barois, I. (2013). Earthworm communities and soil properties in shaded coffee plantations with and without application of glyphosate. *Applied Soil Ecology*, *83*, 230–237. <https://doi.org/10.1016/j.apsoil.2013.09.006>
- Garro, M. S., De Valdez, G. F., & De Giori, G. S. (2004). Temperature effect on the biological activity of *Bifidobacterium longum* CRL 849 and *Lactobacillus fermentum* CRL 251 in pure and mixed cultures grown in soymilk. *Food Microbiology*, *21*(5), 511–518. <https://doi.org/10.1016/j.fm.2004.01.001>
- Gavrilescu, M., & Tudose, R. Z. (1999). Modelling mixing parameters in concentric-tube airlift bioreactors. Part II: Axial dispersion. *Bioprocess Engineering*, *20*(6), 491–497. <https://doi.org/10.1007/s004490050620>
- Gervais, P., Marechal, P. A., & Molin, P. (1996). Water relations of solid state fermentation. *Journal of Scientific and Industrial Research*, *55*(5–6), 343–357.
- Gervais, Patrick, & Molin, P. (2003). The role of water in solid-state fermentation. *Biochemical Engineering Journal*, *13*(2–3), 85–101. [https://doi.org/10.1016/S1369-703X\(02\)00122-5](https://doi.org/10.1016/S1369-703X(02)00122-5)
- Godoy, L., Martínez, C., Carrasco, N., & Ganga, M. A. (2008). Purification and characterization of a p-coumarate decarboxylase and a vinylphenol reductase from *Brettanomyces bruxellensis*. *International Journal of Food Microbiology*, *127*(1–2), 6–11. <https://doi.org/10.1016/j.ijfoodmicro.2008.05.011>
- Goličnik, M. (2013). “Die kinetik der invertinwirkung” of L. Michaelis and M.L. Menten revisited after 100 years: Closed-form solutions of genuine invertase-reaction dynamics. *Match*, *70*(1), 63–72.
- Gopalan, N., Rodríguez-Duran, L. V., Saucedo-Castaneda, G., & Nampoothiri, K. M. (2015). Review on technological and scientific aspects of feruloyl esterases: A versatile enzyme for biorefining of biomass. *Bioresource Technology*, *193*, 534–544. <https://doi.org/10.1016/j.biortech.2015.06.117>
- Grajeda-González, U. F., Flores-Breceda, H., Aranda-Ruiz, J., Vidales-Contreras, J. A., Rodríguez-Fuentes, H., & Luna-Maldonado, A. I. (2016). Modeling of Corn Grain Drying by Runge- Kutta Method. *Journal of Experimental Biology and Agricultural Sciences*, *4*(5), 462–466. [https://doi.org/10.18006/2016.4\(5\).462.466](https://doi.org/10.18006/2016.4(5).462.466)
- Grosso, C., Ferreres, F., Gil-Izquierdo, A., Valentão, P., Sampaio, M., Lima, J., & Andrade, P. B. (2014). Box-Behnken factorial design to obtain a phenolic-rich extract from the aerial parts of *Chelidonium majus* L. *Talanta*, *130*, 128–136.

<https://doi.org/10.1016/j.talanta.2014.06.043>

- Guido, L. F., & Moreira, M. M. (2017). Techniques for Extraction of Brewer's Spent Grain Polyphenols: a Review. *Food and Bioprocess Technology*, *10*(7), 1192–1209. <https://doi.org/10.1007/s11947-017-1913-4>
- Gullón, B., Yáñez, R., Alonso, J. L., & Parajó, J. C. (2010). Production of oligosaccharides and sugars from rye straw: A kinetic approach. *Bioresource Technology*, *101*(17), 6676–6684. <https://doi.org/10.1016/j.biortech.2010.03.080>
- Hanrahan, G., & Lu, K. (2006). Application of factorial and response surface methodology in modern experimental design and optimization. *Critical Reviews in Analytical Chemistry*, *36*(3–4), 141–151. <https://doi.org/10.1080/10408340600969478>
- Harmsen, P., Huijgen, W. J. J., Bermudez, L., & Bakker, R. R. (2010). Literature review of physical and chemical pretreatment processes for lignocellulosic biomass. In *Wageningen UR-Food & Biobased Research*.
- Hatamoto, M., Imachi, H., Yashiro, Y., Ohashi, A., & Harada, H. (2007). Diversity of anaerobic microorganisms involved in long-chain fatty acid degradation in methanogenic sludges as revealed by RNA-based stable isotope probing. *Applied and Environmental Microbiology*, *73*(13), 4119–4127. <https://doi.org/10.1128/AEM.00362-07>
- Henri, V. (1901). Über das Gesetz der Wirkung des Invertins. *Zeitschrift Für Physikalische Chemie*, *39U*(1). <https://doi.org/10.1515/zpch-1902-3912>
- Herrick, J. B., Madsen, E. L., Batt, C. A., & Ghiorse, W. C. (1993). Polymerase chain reaction amplification of naphthalene-catabolic and 16S rRNA gene sequences from indigenous sediment bacteria. *Appl Environ Microbiol.*, *59*(3), 687–694.
- Hill, W. J., & Hunter, W. G. (1966). A Review of Response Surface Methodology: A Literature Survey. *Technometrics*, *8*(4), 571–590. <https://doi.org/10.1080/00401706.1966.10490404>
- Hodge, D. B., Karim, M. N., Schell, D. J., & McMillan, J. D. (2008). Soluble and insoluble solids contributions to high-solids enzymatic hydrolysis of lignocellulose. *Bioresource Technology*, *99*(18), 8940–8948. <https://doi.org/10.1016/j.biortech.2008.05.015>
- Höfte, H., & Whiteley, H. R. (1989). Insecticidal crystal proteins of *Bacillus thuringiensis*. *Microbiological Reviews*, *53*(2), 242–255. <https://doi.org/10.1128/mbr.53.2.242-255.1989>
- Howard, R. L., Abotsi, E., Van Rensburg, E. L. J., & Howard, S. (2003). Lignocellulose biotechnology: Issues of bioconversion and enzyme production. *African Journal of Biotechnology*, *2*(12), 702–733. <https://doi.org/10.5897/ajb2003.000-1115>
- Huang, W. C., Chen, S. J., & Chen, T. L. (2006). The role of dissolved oxygen and function of agitation in hyaluronic acid fermentation. *Biochemical Engineering Journal*, *32*(3), 239–243. <https://doi.org/10.1016/j.bej.2006.10.011>
- Humphreys, J. M., & Chapple, C. (2002). Rewriting the lignin roadmap. *Current Opinion in Plant Biology*, *5*(3), 224–229. [https://doi.org/10.1016/S1369-5266\(02\)00257-1](https://doi.org/10.1016/S1369-5266(02)00257-1)

- Ibn Abubakar, B. S. U., Abdullah, N., Idris, A., Zakaria, M. P., & Abdul Shokur, M. Y. (2012). Optimisation of pyrene removal by a mixed culture in a soil-slurry bioreactor using response surface methodology (RSM). *Procedia Engineering*, 50(Icasce), 786–799. <https://doi.org/10.1016/j.proeng.2012.10.087>
- Idi, A., Salleh, M. M., Ibrahim, Z., & Mohamad, S. E. (2012). Pretreatment of cocoa waste for bioethanol production using ionic liquid. *Jurnal Teknologi (Sciences and Engineering)*, 59(SUPPL.1), 49–56. <https://doi.org/10.11113/jt.v59.1584>
- Idris, A., & Suzana, W. (2006). Effect of sodium alginate concentration, bead diameter, initial pH and temperature on lactic acid production from pineapple waste using immobilized *Lactobacillus delbrueckii*. *Process Biochemistry*, 41(5), 1117–1123. <https://doi.org/10.1016/j.procbio.2005.12.002>
- Islam, M. U., Ullah, M. W., Khan, S., Shah, N., & Park, J. K. (2017). Strategies for cost-effective and enhanced production of bacterial cellulose. *International Journal of Biological Macromolecules*, 102, 1166–1173. <https://doi.org/10.1016/j.ijbiomac.2017.04.110>
- Ismail, H., Supri, & Yusof, A. M. M. (2004). Blend of waste poly(vinylchloride) (PVCw)/acrylonitrile butadiene-rubber (NBR): The effect of maleic anhydride (MAH). *Polymer Testing*, 23(6), 675–683. <https://doi.org/10.1016/j.polymertesting.2004.01.008>
- Ismail, S. N., & Zainol, N. (2014). Optimization of ferulic acid extraction from banana stem waste. *Asian Journal of Microbiology, Biotechnology and Environmental Sciences*, 16(3), 479–484.
- Jamaluddin, M F, Zainol, N., & Norsam, N. A. (2012). Isolation and identification of acetic acid producer from mixed culture of soil and banana stem waste in anaerobic condition. *International Journal*, 3(2), 1–5.
- Jamaluddin, Mohd Faizan, Salihon, J., Zainol, N., & Abdul-Rahman, R. (2011). Factorial Analysis on Acetic Acid Production Using Banana Stem Waste. *International Journal of Chemical and Environmental Engineering*, x(x), 1–7. Retrieved from http://umpir.ump.edu.my/2886/1/Factorial_Analysis_On_Acetic_Acid_Production_Using_Banana_Stem_Waste.pdf
- Johnson, K. A. (2013). A century of enzyme kinetic analysis, 1913 to 2013. *FEBS Letters*, 587(17), 2753–2766. <https://doi.org/10.1016/j.febslet.2013.07.012>
- Jönsson, L. J., & Martín, C. (2016). Pretreatment of lignocellulose: Formation of inhibitory by-products and strategies for minimizing their effects. *Bioresource Technology*, 199, 103–112. <https://doi.org/10.1016/j.biortech.2015.10.009>
- Kabenge, I., Omulo, G., Banadda, N., Seay, J., Zziwa, A., & Kiggundu, N. (2018). Characterization of Banana Peels Wastes as Potential Slow Pyrolysis Feedstock. *Journal of Sustainable Development*, 11(2), 14. <https://doi.org/10.5539/jsd.v11n2p14>
- Kanski, J., Aksenova, M., Stoyanova, A., & Butterfield, D. A. (2002). Ferulic acid antioxidant protection against hydroxyl and peroxy radical oxidation in synaptosomal and neuronal cell culture systems in vitro: Structure-activity studies. *Journal of Nutritional Biochemistry*, 13(5), 273–281. [https://doi.org/10.1016/S0955-2863\(01\)00215-7](https://doi.org/10.1016/S0955-2863(01)00215-7)

- Karacabey, E., & Mazza, G. (2008). Optimization of solid-liquid extraction of resveratrol and other phenolic compounds from milled grape canes (*Vitis vinifera*). *Journal of Agricultural and Food Chemistry*, 56(15), 6318–6325. <https://doi.org/10.1021/jf800687b>
- Kareem, S. A., Kefas, H. M., Chior, T. J., & Latinwo, G. K. (2011). Kinetics of Fermentation by Enzymes : A Mathematical Approach. *Au. J.T*, 15(2), 109–114.
- Katsoni, A., Frontistis, Z., Xekoukoulotakis, N. P., Diamadopoulou, E., & Mantzavinos, D. (2008). Wet air oxidation of table olive processing wastewater: Determination of key operating parameters by factorial design. *Water Research*, 42(14), 3591–3600. <https://doi.org/10.1016/j.watres.2008.05.007>
- Khuri, A. I., & Mukhopadhyay, S. (2010). Response surface methodology. *Wiley Interdisciplinary Reviews: Computational Statistics*, 2(2), 128–149. <https://doi.org/10.1002/wics.73>
- Khushairi, Z. A., Samad, K. A., Rahman, N. A. A., Yussof, H. W., & Zainol, N. (2020). Application of Michaelis–Menten in the kinetics of oil palm frond enzymatic hydrolysis for ferulic acid production. *SN Applied Sciences*, 2(2). <https://doi.org/10.1007/s42452-020-2062-3>
- Khushairi, Z. A., Yussof, H. W., Rodzri, N. A., Abu Samah, R., & Zainol, N. (2016). A factorial analysis study on factors contribution to ferulic acid production from oil palm frond waste. *Jurnal Teknologi*, 78(10), 147–152. <https://doi.org/10.11113/jt.v78.7339>
- Khushairi, Z. A., Yussof, H. W., & Zainol, N. (2018). Optimization of Ferulic Acid Production from Oil Palm Frond Bagasse. *Sustainable Technologies for the Management of Agricultural Wastes*, 57–69. https://doi.org/10.1007/978-981-10-5062-6_5
- Kim, J. S., Lee, Y. Y., & Kim, T. H. (2016). A review on alkaline pretreatment technology for bioconversion of lignocellulosic biomass. *Bioresource Technology*, 199, 42–48. <https://doi.org/10.1016/j.biortech.2015.08.085>
- Kim, K. H., Tsao, R., Yang, R., & Cui, S. W. (2006). Phenolic acid profiles and antioxidant activities of wheat bran extracts and the effect of hydrolysis conditions. *Food Chemistry*, 95(3), 466–473. <https://doi.org/10.1016/j.foodchem.2005.01.032>
- Kim, M. J., Choi, S. J., Lim, S. T., Kim, H. K., Heo, H. J., Kim, E. K., ... Shin, D. H. (2007). Ferulic acid supplementation prevents trimethyltin-induced cognitive deficits in mice. *Bioscience, Biotechnology and Biochemistry*, 71(4), 1063–1068. <https://doi.org/10.1271/bbb.60564>
- Koehler, T. M. (2009). *Bacillus anthracis* physiology and genetics. *Molecular Aspects of Medicine*, 30(6), 386–396. <https://doi.org/10.1016/j.mam.2009.07.004>
- Koh, M. P., & Hoi, W. K. (2003). Sustainable biomass production for energy in Malaysia. *Biomass and Bioenergy*, 25(5), 517–529. [https://doi.org/10.1016/S0961-9534\(03\)00088-6](https://doi.org/10.1016/S0961-9534(03)00088-6)
- Kolb, M., Schieder, D., Faulstich, M., & Sieber, V. (2013). Analysis of lignocellulose derived phenolic monomers by headspace solid-phase microextraction and gas chromatography. *Journal of Chromatography A*, 1307, 144–157.

<https://doi.org/10.1016/j.chroma.2013.07.094>

- Kucharska, K., Rybarczyk, P., Hołowacz, I., Łukajtis, R., Glinka, M., & Kamiński, M. (2018). Pretreatment of lignocellulosic materials as substrates for fermentation processes. *Molecules*, *23*(11), 1–32. <https://doi.org/10.3390/molecules23112937>
- Kumar, A. K., & Sharma, S. (2017). Recent updates on different methods of pretreatment of lignocellulosic feedstocks: a review. *Bioresources and Bioprocessing*, *4*(1). <https://doi.org/10.1186/s40643-017-0137-9>
- Kumar, M., & Kumar, D. (2011). Comparative study of pulping of banana stem. *International Journal of Fiber and Textile Research*, *1*(1), 1–5.
- Kurasawa, S. I., Sugahara, T., & Hayashi, J. (1982). Studies on dietary fiber of mushrooms and edible wild plants. *Nutrition Reports International (USA)*.
- Laidler, Keith J., & Meiser, J. H. (1982). *Physical Chemistry*. Benjamin-Cummings Pub Co.
- Laidler, Keith James. (1978). *Physical Chemistry with Biological Applications*. Benjamin/Cummings Pub. Co.
- Larkin, M. J., De Mot, R., Kulakov, L. A., & Nagy, I. (1998). Applied aspects of Rhodococcus genetics. *Antonie van Leeuwenhoek, International Journal of General and Molecular Microbiology*, *74*(1–3), 133–153. <https://doi.org/10.1023/A:1001776500413>
- Laszlo, J. A., Compton, D. L., & Li, X. L. (2006). Feruloyl esterase hydrolysis and recovery of ferulic acid from jojoba meal. *Industrial Crops and Products*, *23*(1), 46–53. <https://doi.org/10.1016/j.indcrop.2005.03.005>
- Latha, C. D. S., & Rao, D. B. (2007). *Principles of Biotechnology*. Discovery Publishing House New Delhi.
- Lattanzio, V. (2013). Phenolic Compounds: Introduction. In *Natural Products: Phytochemistry, Botany and Metabolism of Alkaloids, Phenolics and Terpenes*. <https://doi.org/10.1007/978-3-642-22144-6>
- Lau, T., Harbourne, N., & Oruña-Concha, M. J. (2019). Optimization of enzyme-assisted extraction of ferulic acid from sweet corn cob by response surface methodology. *Journal of the Science of Food and Agriculture*, (November). <https://doi.org/10.1002/jsfa.10155>
- Laura Ilzarbe, María Jesús Álvarez, Elisabeth Viles, M. T. (2008). Practical Applications of Design of Experiments in the Field of Engineering: A Bibliographical Review. *Quality and Reliability Engineering International*, *24*(25 September 2007), 229–248. <https://doi.org/10.1002/qre>
- Lee, Z. M. P., Bussema, C., & Schmidt, T. M. (2009). rrn DB: Documenting the number of rRNA and tRNA genes in bacteria and archaea. *Nucleic Acids Research*, *37*(SUPPL. 1), 489–493. <https://doi.org/10.1093/nar/gkn689>
- Lehninger, A. L., Nelson, D. L., & Cox, M. M. (2005). *Lehninger Principles of Biochemistry Fourth Edition*. New York : W.H. Freeman.
- Li, C., Liu, G., Nges, I. A., Deng, L., Nistor, M., & Liu, J. (2016). Fresh banana pseudo-stems as a tropical lignocellulosic feedstock for methane production. *Energy*,

- Li, Y., Kawamura, Y., Fujiwara, N., Naka, T., Liu, H., Huang, X., ... Ezaki, T. (2004). *Sphingomonas yabuuchiae* sp. nov. and *Brevundimonas nasdae* sp. nov., isolated from Russian space laboratory Mir. *International Journal of Systematic and Evolutionary Microbiology*, 54(3), 819–825. <https://doi.org/10.1099/ijs.0.02829-0>
- Liao, W., Wen, Z., Hurley, S., Liu, Y., Liu, C., & Chen, S. (2005). Effects of hemicellulose and lignin on enzymatic hydrolysis of cellulose from dairy manure. *Applied Biochemistry and Biotechnology - Part A Enzyme Engineering and Biotechnology*, 124(1–3), 1017–1030. <https://doi.org/10.1385/ABAB:124:1-3:1017>
- Lin, D., Xiao, M., Zhao, J., Li, Z., Xing, B., Li, X., ... Chen, S. (2016). An overview of plant phenolic compounds and their importance in human nutrition and management of type 2 diabetes. *Molecules*, 21(10). <https://doi.org/10.3390/molecules21101374>
- Lin, F. H., Lin, J. Y., Gupta, R. D., Tournas, J. A., Burch, J. A., Selim, M. A., ... Pinnell, S. R. (2005). Ferulic acid stabilizes a solution of vitamins C and E and doubles its photoprotection of skin. *Journal of Investigative Dermatology*, 125(4), 826–832. <https://doi.org/10.1111/j.0022-202X.2005.23768.x>
- Liyana-Pathirana, C., & Shahidi, F. (2005). Optimization of extraction of phenolic compounds from wheat using response surface methodology. *Food Chemistry*, 93(1), 47–56. <https://doi.org/10.1016/j.foodchem.2004.08.050>
- Longobardi, F., Ventrella, A., Napoli, C., Humpfer, E., Schütz, B., Schäfer, H., ... Sacco, A. (2012). Classification of olive oils according to geographical origin by using 1H NMR fingerprinting combined with multivariate analysis. *Food Chemistry*, 130(1), 177–183. <https://doi.org/10.1016/j.foodchem.2011.06.045>
- Lu, W., Gao, S., Xiao, Y., Zhang, L., Li, J., & Chen, W. (2011). A liquid chromatographic-tandem mass spectrometric method for the quantitation of eight components involved in lithospermic acid B biosynthesis pathway in *Salvia miltiorrhiza* hairy root cultures. *Journal of Medicinal Plants Research*, 5(9), 1664–1672.
- Ma, T. S., & Zaga, G. Z. (1942). Micro-Kjeldahl Determination of Nitrogen. A New Indicator and An Improved Rapid Method. *Industrial and Engineering Chemistry - Analytical Edition*, 14(3), 280–282. <https://doi.org/10.1021/i560103a035>
- Maleque, M. A., Belal, F. Y., & Sapuan, S. M. (2007). Mechanical properties study of pseudo-stem banana fiber reinforced epoxy composite. *Arabian Journal for Science and Engineering*, 32(2 B), 359–364.
- Mann, J. (1987). *Secondary Metabolism Second Edition*. Oxford University Press Inc., New York.
- Manter, D. K., Weir, T. L., & Vivanco, J. M. (2010). Negative effects of sample pooling on PCR-Based estimates of soil microbial richness and community structure. *Applied and Environmental Microbiology*, 76(7), 2086–2090. <https://doi.org/10.1128/AEM.03017-09>
- Mathew, S., & Abraham, T. E. (2004). Ferulic acid: An antioxidant found naturally in plant cell walls and feruloyl esterases involved in its release and their applications. *Critical Reviews in Biotechnology*, 24(2–3), 59–83.

<https://doi.org/10.1080/07388550490491467>

- Max, B., Torrado, A. M., Moldes, A. B., Converti, A., & Domínguez, J. M. (2009). Ferulic acid and p-coumaric acid solubilization by alkaline hydrolysis of the solid residue obtained after acid prehydrolysis of vine shoot prunings: Effect of the hydroxide and pH. *Biochemical Engineering Journal*, 43(2), 129–134. <https://doi.org/10.1016/j.bej.2008.09.015>
- Menon, V., Rao, M., & Prakash, G. (2010). Value added products from hemicellulose - Biotechnological perspective. In *Global Journal of Biochemistry* (Vol. 1). Retrieved from http://ncl.csircentral.net/1138/1/Value_added_products_from_hemicellulose.pdf
- Michaelis, L., & Menten, M. L. (1913). *Die Kinetik der Invertinwirkung*.
- Min, J.-Y., Kang, S.-M., Park, D.-J., Kim, Y.-D., Jung, H.-N., Yang, J.-K., ... Choi, M.-S. (2006). Enzymatic Release of Ferulic Acid from *Ipomoea batatas* L. (Sweet Potato) Stem. *Biotechnology and Bioprocess Engineering*, 11(4), 372–376.
- Mo, X., & Xu, Y. (2010). Ferulic acid release and 4-vinylguaiacol formation during Chinese rice wine brewing and fermentation. *Journal of the Institute of Brewing*, 116(3), 304–311. <https://doi.org/10.1002/j.2050-0416.2010.tb00435.x>
- Mohd Taib, R. (2019). *Pyrolysis of Banana Pseudo-stem and Leaf Through Fast and Slow Processes in Producing Bio-char and Bio-oil*. Universiti Sains Malaysia.
- Monod, J. (1949). The Growth of Bacterial Cultures. *Annual Reviews in M*, 3(XI), 371–394.
- Morgan, M. C., Boyette, M., Goforth, C., Sperry, K. V., & Greene, S. R. (2009). Comparison of the Biolog OmniLog Identification System and 16S ribosomal RNA gene sequencing for accuracy in identification of atypical bacteria of clinical origin. *Journal of Microbiological Methods*, 79(3), 336–343. <https://doi.org/10.1016/j.mimet.2009.10.005>
- Mousavi, M., Noroozian, E., Jalali-Heravi, M., & Mollahosseini, A. (2007). Optimization of solid-phase microextraction of volatile phenols in water by a polyaniline-coated Pt-fiber using experimental design. *Analytica Chimica Acta*, 581(1), 71–77. <https://doi.org/10.1016/j.aca.2006.08.001>
- Mukerjee, R., & Wu, C. F. J. (2006). *A Modern Theory of Factorial Design*.
- Mukherjee, G., Singh, R. K., Mitra, A., & Sen, S. K. (2007). Ferulic acid esterase production by *Streptomyces* sp. *Bioresource Technology*, 98(1), 211–213. <https://doi.org/10.1016/j.biortech.2005.12.001>
- Mussatto, S. I., Dragone, G., & Roberto, I. C. (2007). Ferulic and p-coumaric acids extraction by alkaline hydrolysis of brewer's spent grain. *Industrial Crops and Products*, 25(2), 231–237. <https://doi.org/10.1016/j.indcrop.2006.11.001>
- Mussatto, S., & Teixeira, J. (2010). Lignocellulose as raw material in fermentation processes. *Applied Microbiology and Microbial Biotechnology*, 2, 897–907. <https://doi.org/10.1016/j.jrras.2014.02.003>
- Myers, R. H. (1999). Response surface methodology - Current status and future directions. *Journal of Quality Technology*, 31(1), 30–44.

<https://doi.org/10.1080/00224065.1999.11979891>

- Nagai, K., Ohta, S., Zenda, H., Matsumoto, H., & Makino, M. (1996). Biochemical characterization of a *Pseudomonas fluorescens* strain isolated from a benzalkonium chloride solution. *Chemical Pharmaceutical Bulletin*, 19(6), 873–875.
- Najafpour, G. D., Tajallipour, M., Komeili, M., & Mohammadi, M. (2009). Kinetic model for an up-flow anaerobic packed bed bioreactor: Dairy wastewater treatment. *African Journal of Biotechnology*, 8(15), 3590–3596. <https://doi.org/10.5897/AJB2009.000-9355>
- Najafpour, Ghasem D. (2007). *Biochemical Engineering and Biotechnology*.
- Nasim Ahmad Khan, S. (2004). Elimination of Heavy Metals from Wastewater Using Agricultural Wastes as Adsorbents. *Malaysian Journal of Science*, Volume 23(Issue 1), 43–51.
- Navarro-Villoslada, F., San Vicente, B., & Moreno-Bondi, M. C. (2004). Application of multivariate analysis to the screening of molecularly imprinted polymers for bisphenol a. *Analytica Chimica Acta*, 504(1), 149–162. [https://doi.org/10.1016/S0003-2670\(03\)00766-9](https://doi.org/10.1016/S0003-2670(03)00766-9)
- Noratiqah, K., Madihah, M. S., Aisyah, B. S., Eva, M. S., Suraini, A. A., & Kamarulzaman, K. (2013). Statistical optimization of enzymatic degradation process for oil palm empty fruit bunch (OPEFB) in rotary drum bioreactor using crude cellulase produced from *Aspergillus niger* EFB1. *Biochemical Engineering Journal*, 75, 8–20. <https://doi.org/10.1016/j.bej.2013.03.007>
- Noremberg, B. S., Silva, R. M., Paniz, O. G., Alano, J. H., Gonçalves, M. R. F., Wolke, S. I., ... Carreño, N. L. V. (2017). From banana stem to conductive paper: A capacitive electrode and gas sensor. *Sensors and Actuators, B: Chemical*, 240, 459–467. <https://doi.org/10.1016/j.snb.2016.09.014>
- Noura, Salih, K. M., Jusuf, N. H., Hamid, A. A., & Yusoff, W. M. W. (2009). High Prevalence of *Pseudomonas* Species in Soil Samples from Ternate Island-Indonesia. *Pakistan Journal of Biological Sciences*, 12, 1036–1040.
- Oh, S. E., Kim, K. S., Choi, H. C., Cho, J., & Kim, I. S. (2000). Kinetics and physiological characteristics of autotrophic denitrification by denitrifying sulfur bacteria. *Water Science and Technology*, 42(3–4), 59–68. <https://doi.org/10.2166/wst.2000.0359>
- Okpokwasili, G. C., & Nweke, C. O. (2005). Microbial growth and substrate utilization kinetics. *African Journal of Biotechnology*, 5(4), 305–317. <https://doi.org/10.5897/AJB2006.000-5041>
- Ou, S., & Kwok, K. C. (2004). Ferulic acid: Pharmaceutical functions, preparation and applications in foods. *Journal of the Science of Food and Agriculture*, 84(11), 1261–1269. <https://doi.org/10.1002/jsfa.1873>
- Padam, B. S., Tin, H. S., Chye, F. Y., & Abdullah, M. I. (2014). Banana by-products: an under-utilized renewable food biomass with great potential. *Journal of Food Science and Technology*, 51(12), 3527–3545. <https://doi.org/10.1007/s13197-012-0861-2>
- Palani Swamy, S. kumaran, & Govindaswamy, V. (2015). Therapeutical properties of ferulic acid and bioavailability enhancement through feruloyl esterase. *Journal of Functional Foods*, 17, 657–666. <https://doi.org/10.1016/j.jff.2015.06.013>

- Panagiotou, G., Olavarria, R., & Olsson, L. (2007). *Penicillium brasilianum* as an enzyme factory; the essential role of feruloyl esterases for the hydrolysis of the plant cell wall. *Journal of Biotechnology*, 130(3), 219–228. <https://doi.org/10.1016/j.jbiotec.2007.04.011>
- Peng, X., Misawa, N., & Harayama, S. (2003). Isolation and Characterization of Thermophilic Bacilli Degrading. *Applied and Environmental Microbiology*, 69(3), 1417–1427. <https://doi.org/10.1128/AEM.69.3.1417>
- Pepper, I. L., & Gentry, T. J. (2002). INCIDENCE OF BACILLUS ANTHRACIS IN SOIL. *Soil Science*, 167(10), 627–635.
- Pereira, A. L. S., do Nascimento, D. M., Souza, M. de S. M., Cassales, A. R., Saraiva Morais, J. P., de Paula, R. C. M., ... Feitosa, J. P. A. (2014). Banana (*Musa* sp. cv. Pacovan) pseudostem fibers are composed of varying lignocellulosic composition throughout the diameter. *BioResources*, 9(4), 7749–7763. <https://doi.org/10.15376/biores.9.4.7749-7763>
- Plaggenborg, R., Overhage, J., Loos, A., Archer, J. A. C., Lessard, P., Sinskey, A. J., ... Priefert, H. (2006). Potential of Rhodococcus strains for biotechnological vanillin production from ferulic acid and eugenol. *Applied Microbiology and Biotechnology*, 72(4), 745–755. <https://doi.org/10.1007/s00253-005-0302-5>
- Puri, M., Sharma, D., & Barrow, C. J. (2012). Enzyme-assisted extraction of bioactives from plants. *Trends in Biotechnology*, 30(1), 37–44. <https://doi.org/10.1016/j.tibtech.2011.06.014>
- Ramdhoney, A., & Jeetah, P. (2017). Production of wrapping paper from banana fibres. *Journal of Environmental Chemical Engineering*, 5(5), 4298–4306. <https://doi.org/10.1016/j.jece.2017.08.011>
- Rao, R. S. P. (2005). *STUDIES ON WATER EXTRACTABLE FERULOYL POLYSACCHARIDES FROM NATIVE AND GERMINATED RICE (Oryza sativa) AND RAGI (Eleusine coracana)*. (September).
- Reimer, L. C., Vetcinina, A., Carbasse, J. S., Söhngen, C., Gleim, D., Ebeling, C., & Overmann, J. (2019). BacDive in 2019: Bacterial phenotypic data for High-throughput biodiversity analysis. *Nucleic Acids Research*, 47(D1), D631–D636. <https://doi.org/10.1093/nar/gky879>
- Rillig, M. C., & Mummey, D. L. (2006). Mycorrhizas and soil structure. *New Phytologist*, 171(1), 41–53. <https://doi.org/10.1111/j.1469-8137.2006.01750.x>
- Rodríguez-León, J. A., de Carvalho, J. C., Pandey, A., Soccol, C. R., & Rodríguez-Fernández, D. E. (2018). Kinetics of the Solid-State Fermentation Process. *Current Developments in Biotechnology and Bioengineering*, 57–82. <https://doi.org/10.1016/b978-0-444-63990-5.00004-9>
- Rosazza, J. P. N., Huang, Z., Dostal, L., Volm, T., & Rousseau, B. (1995). Biocatalytic transformations of ferulic acid: An abundant aromatic natural product. *Journal of Industrial Microbiology*, 15(6), 472–479. <https://doi.org/10.1007/BF01570017>
- Roshchina, V. V., & Narwal, S. S. (2007). *Cell Diagnostics: Images, Biophysical and Biochemical Processes in Allelopathy*. Science Publishers.
- Ryan, D., Antolovich, M., Prenzler, P., Robards, K., & Lavee, S. (2002).

- Biotransformations of phenolic compounds in *Olea europaea* L. *Scientia Horticulturae*, 92(2), 147–176. [https://doi.org/10.1016/S0304-4238\(01\)00287-4](https://doi.org/10.1016/S0304-4238(01)00287-4)
- Ryan, M. P., & Pembroke, J. T. (2018). Brevundimonas spp: Emerging global opportunistic pathogens. *Virulence*, 9(1), 480–493. <https://doi.org/10.1080/21505594.2017.1419116>
- Sadeghi, O., Fazeli, A., Bakhtiarinejad, M., & Che Sidik, N. A. (2014). An Overview of Waste-to-Energy in Malaysia. *Applied Mechanics and Materials*, 695, 792–796. <https://doi.org/10.4028/www.scientific.net/amm.695.792>
- Saija, A., Tomaino, A., Trombetta, D., De Pasquale, A., Uccella, N., Barbuzzi, T., ... Bonina, F. (2000). In vitro and in vivo evaluation of caffeic and ferulic acids as topical photoprotective agents. *International Journal of Pharmaceutics*, 199(1), 39–47. [https://doi.org/10.1016/S0378-5173\(00\)00358-6](https://doi.org/10.1016/S0378-5173(00)00358-6)
- Saikia, S., Mahnot, N. K., & Mahanta, C. L. (2015). Optimisation of phenolic extraction from *Averrhoa carambola* pomace by response surface methodology and its microencapsulation by spray and freeze drying. *Food Chemistry*, 171, 144–152. <https://doi.org/10.1016/j.foodchem.2014.08.064>
- Salleh, N. H. M., Daud, M. Z. M., Arbain, D., Ahmad, M. S., & Ismail, K. S. K. (2011). Optimization of alkaline hydrolysis of paddy straw for ferulic acid extraction. *Industrial Crops and Products*, 34(3), 1635–1640. <https://doi.org/10.1016/j.indcrop.2011.06.010>
- Samad, K. A., & Zainol, N. (2016). Release of ferulic acid from banana stem waste by ferulic acid esterase from co-culture. *International Research Journal of Engineering and Technology*, 3(8), 819–823.
- Samad, K. A., & Zainol, N. (2017a). Effects of agitation and volume of inoculum on ferulic acid production by co-culture. *Biocatalysis and Agricultural Biotechnology*, 10(August 2016), 9–12. <https://doi.org/10.1016/j.bcab.2017.01.010>
- Samad, K. A., & Zainol, N. (2017b). The use of factorial design for ferulic acid production by co-culture. *Industrial Crops and Products*, 95, 202–206. <https://doi.org/10.1016/j.indcrop.2016.10.028>
- Samad, K. A., Zainol, N., & Mohd Syukri, N. S. (2016). Development of soil co-culture system for ferulic acid production. *ARPN Journal of Engineering and Applied Sciences*, 11(4), 2253–2257.
- Sands, D. C., & Rovira, A. D. (1971). *Pseudomonas fluorescens* Biotype G, the Dominant Fluorescent Pseudomonad in South Australian Soils and Wheat Rhizospheres. *Journal of Applied Bacteriology*, 34(1), 261–275. <https://doi.org/10.1111/j.1365-2672.1971.tb02285.x>
- Sango, T., Cheumani Yona, A. M., Duchatel, L., Marin, A., Kor Ndikontar, M., Joly, N., & Lefebvre, J. M. (2018). Step-wise multi-scale deconstruction of banana pseudo-stem (*Musa acuminata*) biomass and morpho-mechanical characterization of extracted long fibres for sustainable applications. *Industrial Crops and Products*, 122(October), 657–668. <https://doi.org/10.1016/j.indcrop.2018.06.050>
- Sarkar, N., Ghosh, S. K., Bannerjee, S., & Aikat, K. (2012). Bioethanol production from agricultural wastes: An overview. *Renewable Energy*, 37(1), 19–27.

<https://doi.org/10.1016/j.renene.2011.06.045>

- Sathasivam, K., & Mas Haris, M. R. H. (2010). Adsorption kinetics and capacity of fatty acid-modified banana trunk fibers for oil in water. *Water, Air, and Soil Pollution*, 213(1–4), 413–423. <https://doi.org/10.1007/s11270-010-0395-z>
- Schroyen, M., Vervaeren, H., Vandepitte, H., Van Hulle, S. W. H., & Raes, K. (2015). Effect of enzymatic pretreatment of various lignocellulosic substrates on production of phenolic compounds and biomethane potential. *Bioresource Technology*, 192, 696–702. <https://doi.org/10.1016/j.biortech.2015.06.051>
- Schutyser, W., Renders, T., Van den Bossche, G., Van den Bosch, S., Koelewijn, S.-F., Ennaert, T., & Sels, B. F. (2017). Catalysis in Lignocellulosic Biorefineries: The Case of Lignin Conversion. In *Nanotechnology in Catalysis*. <https://doi.org/10.1002/9783527699827.ch23>
- Seber, G. A. F., & Lee, A. J. (2003). *Linear Regression Analysis Second Edition*. John Wiley & Sons Inc.
- Senesi, S., & Ghelardi, E. (2010). Production, Secretion and Biological Activity of *Bacillus cereus* Enterotoxins. *Toxins*, 2(7), 1690–1703. <https://doi.org/10.3390/toxins2071690>
- Sgarbossa, A., Giacomazza, D., & Di Carlo, M. (2015). Ferulic acid: A hope for Alzheimer's disease therapy from plants. *Nutrients*, 7(7), 5764–5782. <https://doi.org/10.3390/nu7075246>
- Shin, H.-D., McClendon, S., Le, T., Taylor, F., & Chen, R. R. (2006). A Complete Enzymatic Recovery of Ferulic Acid From Corn Residues With Extracellular Enzymes From *Neosartorya spinosa* NRRL185 Hyun-Dong. *Biotechnology and Bioengineering*, 95(6).
- Shin, H. D., & Chen, R. R. (2006). Production and characterization of a type B feruloyl esterase from *Fusarium proliferatum* NRRL 26517. *Enzyme and Microbial Technology*, 38(3–4), 478–485. <https://doi.org/10.1016/j.enzmictec.2005.07.003>
- Shuler, M. L., Kargi, F., & DeLisa, M. (2017). *Bioprocess Engineering : Basic Concepts* (Third Edit). Pearson Education (US).
- Silhavy, T. J., Kahne, D., & Walker, S. (2010). The bacterial cell envelope. *Cold Spring Harbor Perspectives in Biology*, 2(5). <https://doi.org/10.1101/cshperspect.a000414>
- Silverstein, R. A., Chen, Y., Sharma-Shivappa, R. R., Boyette, M. D., & Osborne, J. (2007). A comparison of chemical pretreatment methods for improving saccharification of cotton stalks. *Bioresource Technology*, 98(16), 3000–3011. <https://doi.org/10.1016/j.biortech.2006.10.022>
- Sindhu, R., Binod, P., & Pandey, A. (2016). Biological pretreatment of lignocellulosic biomass - An overview. *Bioresource Technology*, 199, 76–82. <https://doi.org/10.1016/j.biortech.2015.08.030>
- Song, J. M., Nam, K., Sun, Y. U., Kang, M. H., Kim, C. G., Kwon, S. T., ... Lee, Y. H. (2010). Molecular and biochemical characterizations of a novel arthropod endo- β -1,3-glucanase from the Antarctic springtail, *Cryptopygus antarcticus*, horizontally acquired from bacteria. *Comparative Biochemistry and Physiology - B Biochemistry and Molecular Biology*, 155(4), 403–412.

<https://doi.org/10.1016/j.cbpb.2010.01.003>

- Sousa, F., Jus, S., Erbel, A., Kokol, V., Cavaco-Paulo, A., & Gubitz, G. M. (2007). A novel metalloprotease from *Bacillus cereus* for protein fibre processing. *Enzyme and Microbial Technology*, *40*(7), 1772–1781. <https://doi.org/10.1016/j.enzmictec.2006.12.017>
- Srinivasan, M., Sudheer, A. R., & Menon, V. P. (2007). Ferulic acid: Therapeutic potential through its antioxidant property. *Journal of Clinical Biochemistry and Nutrition*, *40*(2), 92–100. <https://doi.org/10.3164/jcbn.40.92>
- Stanbury, P., Whitaker, A., & Hall, S. (2016). *Principles of Fermentation Technology Third Edition*.
- Stat-Ease Inc. (2020). Stat-Ease. Retrieved from <https://www.statease.com/>
- Stratil, P., Klejdus, B., & Kubáň, V. (2007). Determination of phenolic compounds and their antioxidant activity in fruits and cereals. *Talanta*, *71*(4), 1741–1751. <https://doi.org/10.1016/j.talanta.2006.08.012>
- Süli, E., & Mayers, D. F. (2003). *An Introduction to Numerical Analysis*. Cambridge University Press.
- Sumathi, S., Chai, S. P., & Mohamed, A. R. (2008). Utilization of oil palm as a source of renewable energy in Malaysia. *Renewable and Sustainable Energy Reviews*, *12*(9), 2404–2421. <https://doi.org/10.1016/j.rser.2007.06.006>
- Sun, S., Sun, S., Cao, X., & Sun, R. (2016). The role of pretreatment in improving the enzymatic hydrolysis of lignocellulosic materials. *Bioresource Technology*, *199*, 49–58. <https://doi.org/10.1016/j.biortech.2015.08.061>
- Suzuki, A., Kagawa, D., Fujii, A., Ochiai, R., Tokimitsu, I., & Saito, I. (2002). Ferulic Acid on Blood Pressure in. *Change*, *7061*(01), 351–357.
- Swatsitang, P., Tucker, G., Robards, K., & Jardine, D. (2000). Isolation and identification of phenolic compounds in *Citrus sinensis*. *Analytica Chimica Acta*, *417*(2), 231–240. [https://doi.org/10.1016/S0003-2670\(00\)00937-5](https://doi.org/10.1016/S0003-2670(00)00937-5)
- Szwajgier, D., & Jakubczyk, A. (2011). *Production of extracellular ferulic acid esterases by*. *10*(3), 287–302.
- Teoh, Y. P., & Mat Don, M. (2011). Kinetic model for the hydrolysis of sterilized palm press fibre. *Chemical Engineering Science*, *66*(15), 3523–3530. <https://doi.org/10.1016/j.ces.2011.04.011>
- The Expresswire. (2019, November). Ferulic Acid Market 2019 Industry Size and Share Evolution to 2026 by Growth Insight , Key Development , Growth Status , Top Key Players , Trends and Forecast by Market Reports World. *The Expresswire*. Retrieved from <https://www.marketwatch.com/press-release/ferulic-acid-market-2019-industry-size-and-share-evolution-to-2026-by-growth-insight-key-development-growth-status-top-key-players-trends-and-forecast-by-market-reports-world-2019-11-20>
- Tilay, A., Bule, M., Kishenkumar, J., & Annapure, U. (2008). Preparation of ferulic acid from agricultural wastes: Its improved extraction and purification. *Journal of Agricultural and Food Chemistry*, *56*(17), 7644–7648.

<https://doi.org/10.1021/jf801536t>

- Toms, A., & Wood, J. M. (1969). The Degradation of trans-Ferulic Acid. *Biochemistry*, 9(2), 337–343.
- Topakas, E, & Christakopoulos, P. (2004). *Enzymic release of phenolic antioxidants from plant cell-wall material*. (February).
- Topakas, Evangelos, Vafiadi, C., & Christakopoulos, P. (2007). Microbial production, characterization and applications of feruloyl esterases. *Process Biochemistry*, 42(4), 497–509. <https://doi.org/10.1016/j.procbio.2007.01.007>
- Tripathi, S., Singh, S., Gangwar, A., Mishra, O. P., Chakrabarti, S. K., Bhardwaj, N. K., & Varadhan, R. (2013). Blending of banana stem with wheat straw and bagasse to enhance physical strength properties of paper. *IPPTA: Quarterly Journal of Indian Pulp and Paper Technical Association*, 25(2), 121–125.
- Vaithanomsat, P., & Apiwatanapiwat, W. (2009). Feasibility study on vanillin production from *Jatropha curcas* stem using steam explosion as a pretreatment. *Inter J Chem Biolo Engr*, 3(5), 839–842. Retrieved from <http://www.waset.org/publications/15930>
- Van Alphen, B. J., & Stoorvogel, J. J. (2000). A functional approach to soil characterization in support of precision agriculture. *Soil Science Society of America Journal*, 64(5), 1706–1713. <https://doi.org/10.2136/sssaj2000.6451706x>
- Van Soeste, P. J., & Wine, R. H. (1967). Method for Determination of Lignin Cellulose and Silica. *Journal of Animal Science*, 26(4), 940.
- Vanderghem, C., Brostaux, Y., Jacquet, N., Blecker, C., & Paquot, M. (2012). Optimization of formic/acetic acid delignification of *Miscanthus×giganteus* for enzymatic hydrolysis using response surface methodology. *Industrial Crops and Products*, 35(1), 280–286. <https://doi.org/10.1016/j.indcrop.2011.07.014>
- Vatankhah, A. R. (2014). Discussion of “application of excel solver for parameter estimation of the nonlinear Muskingum models” by Reza Barati. *KSCE Journal of Civil Engineering*, 19(1), 332–336. <https://doi.org/10.1007/s12205-014-1422-1>
- Ventura, S. P. M., E Silva, F. A., Quental, M. V., Mondal, D., Freire, M. G., & Coutinho, J. A. P. (2017). Ionic-Liquid-Mediated Extraction and Separation Processes for Bioactive Compounds: Past, Present, and Future Trends. *Chemical Reviews*, 117(10), 6984–7052. <https://doi.org/10.1021/acs.chemrev.6b00550>
- Wan Ab Karim Ghani, W. A., Moghadam, R. A., & Mohd Salleh, M. A. (2011). Air Gasification of Malaysia Agricultural Waste in a Fluidized Bed Gasifier: Hydrogen Production Performance. *Sustainable Growth and Applications in Renewable Energy Sources*. <https://doi.org/10.5772/26111>
- Wang, J., Sun, B., Cao, Y., Tian, Y., & Li, X. (2008). Optimisation of ultrasound-assisted extraction of phenolic compounds from wheat bran. *Food Chemistry*, 106(2), 804–810. <https://doi.org/10.1016/j.foodchem.2007.06.062>
- Wang, X., Wu, Y., Chen, G., Yue, W., Liang, Q., & Wu, Q. (2013). Optimisation of ultrasound assisted extraction of phenolic compounds from *Sparganii rhizoma* with response surface methodology. *Ultrasonics Sonochemistry*, 20(3), 846–854. <https://doi.org/10.1016/j.ultsonch.2012.11.007>

- Whetten, R., & Sederoff, R. (1995). Lignin biosynthesis. *Plant Cell*, 7(7), 1001–1013. <https://doi.org/10.1105/tpc.7.7.1001>
- Wiebe, W. L., & Campbell, R. . (1993). Characterization of *Pseudomonas syringae* pv. *maculicola* and Comparison with *P. s. tomato*. *Plant Disease*, Vol. 77, p. 414. <https://doi.org/10.1094/pd-77-0414>
- William, R., & Husain, M. (2010). *Status of Banana Cultivation and Disease Incidences in Malaysia*.
- Williamson, K., & McCarty, P. L. (1976). A Model of Substrate Utilization by Bacterial Films. *Water Pollution Control Federation*, 48(1), 9–24.
- Wilson, T. A., Nicolosi, R. J., Woolfrey, B., & Kritchevsky, D. (2007). Rice bran oil and oryzanol reduce plasma lipid and lipoprotein cholesterol concentrations and aortic cholesterol ester accumulation to a greater extent than ferulic acid in hypercholesterolemic hamsters. *Journal of Nutritional Biochemistry*, 18(2), 105–112. <https://doi.org/10.1016/j.jnutbio.2006.03.006>
- Wisker, E., Feldheim, W., Pomeranz, Y., & Meuser, F. (1985). *Advances in Cereal Science and Technology*.
- Wong, W. H., Lee, W. X., Ramanan, R. N., Tee, L. H., Kong, K. W., Galanakis, C. M., ... Prasad, K. N. (2015). Two level half factorial design for the extraction of phenolics, flavonoids and antioxidants recovery from palm kernel by-product. *Industrial Crops and Products*, 63, 238–248. <https://doi.org/10.1016/j.indcrop.2014.09.049>
- Wu, H., Li, H., Xue, Y., Luo, G., Gan, L., Liu, J., ... Long, M. (2017). High efficiency co-production of ferulic acid and xylooligosaccharides from wheat bran by recombinant xylanase and feruloyl esterase. *Biochemical Engineering Journal*, 120, 41–48. <https://doi.org/10.1016/j.bej.2017.01.001>
- Xie, C. yan, Gu, Z. xin, You, X., Liu, G., Tan, Y., & Zhang, H. (2010). Screening of edible mushrooms for release of ferulic acid from wheat bran by fermentation. *Enzyme and Microbial Technology*, 46(2), 125–128. <https://doi.org/10.1016/j.enzmictec.2009.10.005>
- Xiros, C., Katapodis, P., & Christakopoulos, P. (2009). Evaluation of *Fusarium oxysporum* cellulolytic system for an efficient hydrolysis of hydrothermally treated wheat straw. *Bioresource Technology*, 100(21), 5362–5365. <https://doi.org/10.1016/j.biortech.2009.05.065>
- Xiros, C., Moukouli, M., Topakas, E., & Christakopoulos, P. (2009). Factors affecting ferulic acid release from Brewer's spent grain by *Fusarium oxysporum* enzymatic system. *Bioresource Technology*, 100(23), 5917–5921. <https://doi.org/10.1016/j.biortech.2009.06.018>
- Xu, F., Sun, R. C., Sun, J. X., Liu, C. F., He, B. H., & Fan, J. S. (2005). Determination of cell wall ferulic and p-coumaric acids in sugarcane bagasse. *Analytica Chimica Acta*, 552(1–2), 207–217. <https://doi.org/10.1016/j.aca.2005.07.037>
- Xu, Y., Hanna, M. A., & Isom, L. (2008). “Green” Chemicals from Renewable Agricultural Biomass - A Mini Review. *The Open Agriculture Journal*, 2(1), 54–61. <https://doi.org/10.2174/1874331500802010054>

- Yan, L., Chen, P., Zhang, S., Li, S., Yan, X., Wang, N., ... Li, H. (2016). Biotransformation of ferulic acid to vanillin in the packed bed-stirred fermentors. *Scientific Reports*, 6(September), 1–12. <https://doi.org/10.1038/srep34644>
- Yoon, S.-H., Li, C., Lee, Y.-M., Lee, S.-H., Kim, S.-H., Choi, M.-S., ... Kim, S.-W. (2005). Production of Vanillin from Ferulic Acid Using Recombinant Strains of *Escherichia coli*. *Biotechnology and Bioprocess Engineering*, 10(4), 378–384.
- Zafar, S. (2015). Agricultural Biomass in Malaysia - BioEnergy Consult. *BioEnergy Consult*, 0–1. Retrieved from <http://www.bioenergyconsult.com/tag/biomass-wastes-in-malaysia/>
- Zainal, Z. A., Ali, R., Lean, C. H., & Seetharamu, K. N. (2001). Prediction of performance of a downdraft gasifier using equilibrium modeling for different biomass materials. *Energy Conversion and Management*, 42(12), 1499–1515. [https://doi.org/10.1016/S0196-8904\(00\)00078-9](https://doi.org/10.1016/S0196-8904(00)00078-9)
- Zainol, N., & Salihon, J. (2009). *Biogas Production from Waste using Biofilm Reactor : Factor Analysis in Two Stages System*. 3(6), 30–34.
- Zainol, N., Salihon, J., & Abdul-Rahman, R. (2008). Biogas production from banana stem waste: Optimisation of 10 l sequencing batch reactor. *2008 IEEE International Conference on Sustainable Energy Technologies, ICSET 2008*, 357–359. <https://doi.org/10.1109/ICSET.2008.4747032>
- Zake, J., Pietsch, S. A., Friedel, J. K., & Zechmeister-Boltenstern, S. (2015). Can agroforestry improve soil fertility and carbon storage in smallholder banana farming systems? *Journal of Plant Nutrition and Soil Science*, 178(2), 237–249. <https://doi.org/10.1002/jpln.201400281>
- Zhang, H., Kallimanis, A., Koukkou, A. I., & Drainas, C. (2004). Isolation and characterization of novel bacteria degrading polycyclic aromatic hydrocarbons from polluted Greek soils. *Applied Microbiology and Biotechnology*, 65(1), 124–131. <https://doi.org/10.1007/s00253-004-1614-6>
- Zhang, Z. Y., Pan, L. P., & Li, H. H. (2010). Isolation, identification and characterization of soil microbes which degrade phenolic allelochemicals. *Journal of Applied Microbiology*, 108(5), 1839–1849. <https://doi.org/10.1111/j.1365-2672.2009.04589.x>
- Zheng, H. Z., Hwang, I. W., & Chung, S. K. (2009). Enhancing polyphenol extraction from unripe apples by carbohydrate- hydrolyzing enzymes. *Journal of Zhejiang University: Science B*, 10(12), 912–919. <https://doi.org/10.1631/jzus.B0920186>
- Zheng, Y., Wu, X. M., Branford-White, C., Ning, X., Quan, J., & Zhu, L. M. (2009). Enzymatic synthesis and characterization of novel feruloylated lipids in selected organic media. *Journal of Molecular Catalysis B: Enzymatic*, 58(1–4), 65–71. <https://doi.org/10.1016/j.molcatb.2008.11.005>
- Zhou, X., & Wu, F. (2012). Effects of amendments of ferulic acid on soil microbial communities in the rhizosphere of cucumber (*Cucumis sativus* L.). *European Journal of Soil Biology*, 50, 191–197. <https://doi.org/10.1016/j.ejsobi.2012.03.001>
- Zhu, L., Zhang, Z., Xia, N., Zhang, W., Wei, Y., Huang, J., ... Yang, L. (2019). Anti-arthritis activity of ferulic acid in complete Freund's adjuvant (CFA)-induced

- arthritis in rats: JAK2 inhibition. *Inflammopharmacology*, 28(2), 463–473.
<https://doi.org/10.1007/s10787-019-00642-0>
- Zularisam, A. W., Ismail, A. F., Salim, M. R., Sakinah, M., & Matsuura, T. (2009). Application of coagulation-ultrafiltration hybrid process for drinking water treatment: Optimization of operating conditions using experimental design. *Separation and Purification Technology*, 65(2), 193–210.
<https://doi.org/10.1016/j.seppur.2008.10.018>
- Zuorro, A. (2015). Optimization of polyphenol recovery from espresso coffee residues using factorial design and response surface methodology. *Separation and Purification Technology*, 152, 64–69. <https://doi.org/10.1016/j.seppur.2015.08.016>
- Zwietering, M. H., Rombouts, F. M., & Van 't Riet, K. (1992). Comparison of definitions of the lag phase and the exponential phase in bacterial growth. *Journal of Applied Bacteriology*, 72, 139–145. <https://doi.org/https://doi.org/10.1111/j.1365-2672.1992.tb01815.x>