

REINFORCEMENT AND OPTIMIZATION OF
HYDROXYETHYL CELLULOSE / POLY
(VINYL ALCOHOL) WITH CELLULOSE
NANOCRYSTAL AS A BONE TISSUE
ENGINEERING SCAFFOLD

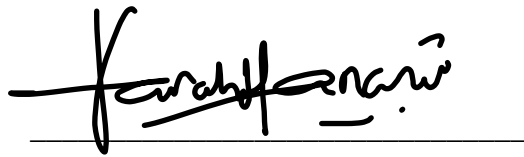
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STUDENT'S DECLARATION

I hereby declare that the work in this thesis is based on my original work except for quotations and citations which have been duly acknowledged. I also declare that it has not been previously or concurrently submitted for any other degree at Universiti Malaysia Pahang or any other institutions.

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ABSTRAK

Bahan bio adalah istilah perubatan yang digunakan untuk menggambarkan semua sumber semulajadi atau sintetik seperti polimer yang berguna dalam pengenalan tisu hidup sebagai sebahagian daripada peranti perubatan atau implan tanpa menyebabkan sebarang tindak balas imun. Selulosa telah diterokai secara meluas sejak beberapa dekad sebagai salah satu daripada bahan bio yang digunakan dalam aplikasi kejuruteraan tisu kerana sifatnya yang unik yang kos rendah, biokompatibilitas yang baik dan sifat mekanikal yang baik. Penyediaan kristal nano selulosa (CNC) dari pulpa selulosa adalah cara alternatif untuk memenuhi permintaan CNC. Dalam kejuruteraan tisu, penggantian atau regenerasi tulang yang rosak adalah satu cabaran utama dalam pembedahan ortopedik. Oleh itu, reka bentuk kejuruteraan tisu tulang berasaskan perancah untuk mengatasi kecacatan tulang ini. Laporan ini terdiri daripada dua bahagian. Bahagian pertama adalah mengenai fabrikasi dan pencirian CNC manakala bahagian kedua adalah fabrikasi dan pencirian perancah termasuk degradasi secara vitro dan kajian hidup sel. Dalam kerja-kerja ini, CNC yang dihasilkan daripada tandan buah kosong (EFB) berjaya dihasilkan oleh hidrolisis asid. Pulpa selulosa dipanaskan pada suhu 85 °C dalam 65 % asid sulfurik. Pulpa selulosa telah dicairkan, sonikasi, dan kemudian mengalami pengeringan beku untuk mendapatkan CNC. CNC bertindak sebagai pengisi nano dalam perancah dan akan dicirikan secara fizikal, kimia dan termal dicirikan dengan menggunakan mikroskop elektron pengimbasan pelepasan medan (FESEM), keseluruhan reflektansi dilemahkan-Fourier mengubah spektroskopi inframerah (ATR-FTIR) dan calorimetry scanning differential (DSC). Keputusan FESEM menunjukkan bahawa CNC muncul dalam bentuk sfera dengan dimensi zarah dalam diameter 5 hingga 30 nm. Spektrum penyerapan CNC muncul dalam kumpulan tertentu yang berada pada 1045, 1346, 1637, 2903, dan 3391 cm^{-1} . Termometer DSC menunjukkan bahawa suhu lebur, T_m berlaku pada 336.4 °C, manakala suhu peralihan kaca, T_g ialah 43.5 °C. Seterusnya, perancah tiga dimensi (3D) HEC / PVA dan HEC / PVA / CNC telah berjaya direka oleh teknik pengeringan beku. HEC (5 % berat) dan PVA (15 % berat) telah dibubarkan dan dicampur pada nisbah 50:50 dan digabungkan dengan pelbagai kepekatan CNC (1, 3, 5 dan 7 % berat). Ciri-ciri morfologi, mekanikal dan terma perancah dicirikan oleh SEM, ATR-FTIR, DSC, thermogravimetric (TGA), dan mesin tegangan sejagat (UTM). Tingkah laku perancah scaffolds dicirikan oleh satu siri analisis termasuk nisbah pembengkakan, penurunan berat dan perubahan pH. Sementara itu, kajian sitotoksikiti pada kedua-dua bahan bio perancah dilakukan dengan menggunakan sel osteoblast janin manusia (hFOB) menggunakan ujian MTT dan kajian morfologi perancah sel. Gabungan HEC/PVA dengan CNC dipamerkan kefungsiannya yang lebih baik yang mengakibatkan penurunan ukuran liang rata-rata dan terdapat sedikit perubahan dalam struktur kimia seperti yang ditentukan oleh spektrum FTIR. Kajian termal menunjukkan bahawa suhu lebur perancah HEC/PVA/CNC beralih kepada nilai yang lebih tinggi. Selain itu, dapat dilihat bahawa penambahan CNC menghasilkan peningkatan tekanan muktamad (dari 0.18 hingga 0.92) dan tegangan muktamad (dari 5.83 hingga 11.03). Oleh itu, ia menawarkan prestasi mekanikal yang sangat baik. Kajian kultur sel mendedahkan bahawa sel-sel hFOB dapat melekat dan menyebarkan ke semua perancah dan menyokong perekatan sel dan percambahan sel. Kepekatan optimum CNC yang paling baik didapati pada 3 dan 5 % berat dan tambahan CNC mengurangkan daya tahan sel. Oleh kerana ciri-ciri bio serasi dan terbiodegradasi, perancah yang sangat baru ini boleh menghasilkan matriks perancah alternatif yang menjanjikan untuk pertumbuhan semula kejuruteraan tisu tulang.

ABSTRACT

Biomaterial is a medical terminology that is used to describe all natural or synthetic resources such as polymers that are useful in the introduction of living tissue as part of medical devices or implants without causing any adverse immune rejection reactions. Cellulose has been extensively explored over many decades as one of the biomaterials used in tissue engineering applications due to their unique properties which are low cost, good biocompatibility and good mechanical properties. Preparation of cellulose nanocrystals (CNC) from cellulose pulp is an alternative way to fulfil the demand for CNC. In tissue engineering, replacement or regeneration of damaged bone is a major challenge in orthopaedic surgery. Hence, scaffold-based bone tissue engineering is designed to overcome these bone defects. This report is comprised of two parts. The first part is about the fabrication and characterization of CNC while the second part is the fabrication and characterization of scaffolds including in vitro degradation and cell culture studies. In this present work, CNC produced from empty fruit bunch (EFB) was successfully fabricated by acid hydrolysis. Cellulose pulps were heated at 85 °C in 65 % of sulphuric acid. The cellulose suspension was diluted, centrifuged, sonicated, and then freeze-dried to obtain the CNC. The CNC acted as nanofillers in scaffolds and was physically, chemically and thermally characterized by using field emission scanning electron microscope (FESEM), attenuated total reflectance-Fourier transform infrared spectroscopy (ATR-FTIR) and differential scanning calorimetry (DSC). FESEM results showed that CNC appeared in a spherical shape with particle dimensions in the range of 5 to 30 nm in diameter. The absorption spectra of CNC appeared in specific bands which were at 1045, 1346, 1637, 2903, and 3391 cm^{-1} . DSC thermograms shows that the melting temperature, T_m was at 336.4 °C, while the glass transition temperature, T_g was 43.5 °C. Next, a porous three-dimensional (3D) scaffold of HEC/PVA and HEC/PVA/CNC were successfully fabricated by freeze-drying technique. HEC (5 wt%) and PVA (15 wt%) were dissolved and blended at a ratio of 50:50 and incorporated with various concentrations of CNC (1, 3, 5 and 7 wt%). The morphology, mechanical and thermal properties of scaffolds were characterized by SEM, ATR-FTIR, DSC, thermogravimetric analysis (TGA), and universal tensile machine (UTM). The degradation behaviours of scaffolds were characterized by a series of analyses including swelling ratio, weight loss and pH changes. Meanwhile, cytotoxicity studies on both porous scaffold biomaterials were carried out by utilizing human fetal osteoblast (hFOB) cells using MTT assays and cell-scaffold morphological study. HEC/PVA incorporated with CNC exhibited superior functionality which resulted in decreased average pore size and there were some slight changes in the chemical structure as determined by FTIR spectra. Thermal studies revealed that the melting temperatures of HEC/PVA/CNC scaffold were slightly shifted to a higher value. Furthermore, it can be seen that the addition of CNC resulted in increases in the ultimate tensile stress (from 0.18 to 0.92) and ultimate tensile strain (from 5.83 to 11.03). Hence, it offers a very good mechanical performance. The cell culture study revealed that the hFOB cells were able to attach and spread on all scaffolds and supported the cell adhesion and proliferation. The optimum concentration of CNC was at 3 and 5 wt% while further addition of CNC reduced the cell viability. Due to its biocompatible and biodegradable properties, these newly developed highly porous scaffolds may provide a promising alternative scaffolding matrix for bone tissue engineering regeneration.

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REFERENCES

- Abd-Khorsand, S., Saber-Samandari, S., & Saber-Samandari, S. (2017). Development of nanocomposite scaffolds based on TiO₂ doped in grafted chitosan/hydroxyapatite by freeze drying method and evaluation of biocompatibility. *Int J Biol Macromol*, 101, 51-58.
- Ali, M. E., & Lamprecht, A. (2017). Spray freeze drying as an alternative technique for lyophilization of polymeric and lipid-based nanoparticles. *Int J Pharm*, 516(1-2), 170-177.
- Alizadeh, M., Abbasi, F., Khoshfetrat, A. B., & Ghaleh, H. (2013). Microstructure and characteristic properties of gelatin/chitosan scaffold prepared by a combined freeze-drying/leaching method. *Mater Sci Eng C Mater Biol Appl*, 33(7), 3958-3967.
- Anandan, D., Madhumathi, G., Nambiraj, N. A., & Jaiswal, A. K. (2019). Gum based 3D composite scaffolds for bone tissue engineering applications. *Carbohydr Polym*, 214, 62-70.
- Bhardwaj, N., Chakraborty, S., & Kundu, S. C. (2011). Freeze-gelled silk fibroin protein scaffolds for potential applications in soft tissue engineering. *Int J Biol Macromol*, 49(3), 260-267.
- Bhat, S., & Kumar, A. (2013). Biomaterials and bioengineering tomorrow's healthcare. *Biomatter*, 3(3).
- Borkotoky, S. S., Dhar, P., & Katiyar, V. (2018). Biodegradable poly (lactic acid)/Cellulose nanocrystals (CNCs) composite microcellular foam: Effect of nanofillers on foam cellular morphology, thermal and wettability behavior. *Int J Biol Macromol*, 106, 433-
- Butylina, S., Geng, S., & Oksman, K. (2016). Properties of as-prepared and freeze-dried hydrogels made from poly(vinyl alcohol) and cellulose nanocrystals using freeze-thaw technique. *European Polymer Journal*, 81, 386-396.
- Caló, E., & Khutoryanskiy, V. V. (2015). Biomedical applications of hydrogels: A review of patents and commercial products. *European Polymer Journal*, 65, 252-267.
- Chocholata, P., Kulda, V., & Babuska, V. (2019). Fabrication of Scaffolds for Bone-Tissue Regeneration. *Materials (Basel)*, 12(4).
- Clarke, B. (2008). Normal bone anatomy and physiology. *Clin J Am Soc Nephrol*, 3 Suppl 3, S131-139.
- Daniel A. Osorio, B. E. J. L., Jacek M. Kwiecien, Xiaoyue Wang, Iflah Shahid, Ariana L. Hurley, Emily D. Cranston, Kathryn Grandfield. (2019). Cross-linked Cellulose Nanocrystal Aerogels as Viable Bone Tissue Scaffolds.
- Edwards, J. V., Prevost, N., Sethumadhavan, K., Ullah, A., & Condon, B. (2013). Peptide conjugated cellulose nanocrystals with sensitive human neutrophil

- elastase sensor activity. *Cellulose*, 20(3), 1223-1235.
- Fereshteh, Z., Fathi, M., Bagri, A., & Boccaccini, A. R. (2016). Preparation and characterization of aligned porous PCL/zein scaffolds as drug delivery systems via improved unidirectional freeze-drying method. *Mater Sci Eng C Mater Biol Appl*, 68, 613-622.
- Fonte, P., Reis, S., & Sarmiento, B. (2016). Facts and evidences on the lyophilization of polymeric nanoparticles for drug delivery. *J Control Release*, 225, 75-86.
- Gavasane, A. J. (2014). Synthetic Biodegradable Polymers Used in Controlled Drug Delivery System: An Overview. *Clinical Pharmacology & Biopharmaceutics*, 3(2).
- Geetha, B., Premkumar, J., Pradeep, J. P., & Krishnakumar, S. (2019). Synthesis and characterization of bioscaffolds using freeze drying technique for bone regeneration. *Biocatalysis and Agricultural Biotechnology*, 20.
- Ghafari, R., Jonoobi, M., Amirabad, L. M., Oksman, K., & Taheri, A. R. (2019). Fabrication and characterization of novel bilayer scaffold from nanocellulose based aerogel for skin tissue engineering applications. *Int J Biol Macromol*, 136, 796-803.
- Ghorbani, F., Nojehdehian, H., & Zamanian, A. (2016). Physicochemical and mechanical properties of freeze cast hydroxyapatite-gelatin scaffolds with dexamethasone loaded PLGA microspheres for hard tissue engineering applications. *Mater Sci Eng C Mater Biol Appl*, 69, 208-220.
- Ghorbani, F., Zamanian, A., & Nojehdehian, H. (2017). Effects of pore orientation on in-vitro properties of retinoic acid-loaded PLGA/gelatin scaffolds for artificial peripheral nerve application. *Mater Sci Eng C Mater Biol Appl*, 77, 159-172.
- Gokmen, F. O., Rzayev, Z. M., Salimi, K., Bunyatova, U., Acar, S., Salamov, B., & Turk, M. (2015). Novel multifunctional colloidal carbohydrate nanofiber electrolytes with excellent conductivity and responses to bone cancer cells. *Carbohydr Polym*, 133, 624-636.
- Henry J. Donahue, Z. L., Zhiyi Zhou, and Clare E. Yellowley. (2000). Differentiation of human fetal osteoblastic cells and gap junctional intercellular communication. *American Physiological Society*, 278, C315-C322.
- Hirano, Y., & Mooney, D. J. (2004). Peptide and Protein Presenting Materials for Tissue Engineering. *Advanced Materials*, 16(1), 17-25.
- Hoggatt, J., & Pelus, L. M. (2013). Hematopoiesis. In *Brenner's Encyclopedia of Genetics* (pp. 418-421).
- Huang, C., Bhagia, S., Hao, N., Meng, X., Liang, L., Yong, Q., & Ragauskas, A. J. (2019). Biomimetic composite scaffold from an in situ hydroxyapatite coating on cellulose nanocrystals. *RSC Advances*, 9(10), 5786-5793.
- Jia An, C. K. C., Ting Yu, Huaqiong Li & Lay Poh Tan. (2013). *Advanced*

nanobiomaterial strategies for the development of organized tissue engineering constructs. *Nanomedicine*, 8(4), 591-602.

- Kai, D., Prabhakaran, M. P., Stahl, B., Eblenkamp, M., Wintermantel, E., & Ramakrishna, S. (2012). Mechanical properties and in vitro behavior of nanofiber-hydrogel composites for tissue engineering applications. *Nanotechnology*, 23(9), 095705.
- Kanimozhi, K., Khaleel Basha, S., & Sugantha Kumari, V. (2016). Processing and characterization of chitosan/PVA and methylcellulose porous scaffolds for tissue engineering. *Mater Sci Eng C Mater Biol Appl*, 61, 484-491.
- Kavitha Sankar, P. C., Rajmohan, G., & Rosemary, M. J. (2017). Physico-chemical characterisation and biological evaluation of freeze dried chitosan sponge for wound care. *Materials Letters*, 208, 130-132.
- Khoshkava, V., & Kamal, M. R. (2014). Effect of drying conditions on cellulose nanocrystal (CNC) agglomerate porosity and dispersibility in polymer nanocomposites. *Powder Technology*, 261, 288-298.
- Kordjamshidi, A., Saber-Samandari, S., Ghadiri Nejad, M., & Khandan, A. (2019). Preparation of novel porous calcium silicate scaffold loaded by celecoxib drug using freeze drying technique: Fabrication, characterization and simulation. *Ceramics International*, 45(11), 14126-14135.
- Kousaku Ohkawa, D. C., Hakyong Kim, Ayako Nishida, Hiroyuki Yamamoto. (2004). Electrospinning of Chitosan. *Macromolecular Rapid Communications*, 1600-16005.
- Kumar, A., Mandal, S., Barui, S., Vasireddi, R., Gbureck, U., Gelinsky, M., & Basu, B. (2016). Low temperature additive manufacturing of three dimensional scaffolds for bone-tissue engineering applications: Processing related challenges and property assessment. *Materials Science and Engineering: R: Reports*, 103, 1-39.
- Lamaming, J., Hashim, R., Leh, C. P., Sulaiman, O., Sugimoto, T., & Nasir, M. (2015). Isolation and characterization of cellulose nanocrystals from parenchyma and vascular bundle of oil palm trunk (*Elaeis guineensis*). *Carbohydr Polym*, 134, 534-540.
- Li, J., Wang, Y., Wei, X., Wang, F., Han, D., Wang, Q., & Kong, L. (2014). Homogeneous isolation of nanocelluloses by controlling the shearing force and pressure in microenvironment. *Carbohydr Polym*, 113, 388-393.
- MacNeil, S. (2007). Progress and opportunities for tissue-engineered skin. *Nature*, 445(7130), 874-880.
- Maji, S., Agarwal, T., Das, J., & Maiti, T. K. (2018). Development of gelatin/carboxymethyl chitosan/nano-hydroxyapatite composite 3D macroporous scaffold for bone tissue engineering applications. *Carbohydr Polym*, 189, 115-125.
- Mohammadi, Z., Mesgar, A. S., & Rasouli-Disfani, F. (2016). Reinforcement of freeze-

- dried chitosan scaffolds with multiphasic calcium phosphate short fibers. *J Mech Behav Biomed Mater*, 61, 590-599.
- N. J. Cao, Q. X., and L. F. Chen. (1995). Acid Hydrolysis of Cellulose in Zinc Chloride Solution. *Applied Biochemistry and Biotechnology*, 51/52.
- Nasution, H., Harahap, H., Afandy, Y., & Fath, M. T. A. (2017). The effect of cellulose nanocrystal (CNC) from rattan biomass as filler and citric acid as co-plasticizer on tensile properties of sago starch biocomposite.
- Preethi Soundarya, S., Sanjay, V., Haritha Menon, A., Dhivya, S., & Selvamurugan, N. (2018). Effects of flavonoids incorporated biological macromolecules based scaffolds in bone tissue engineering. *Int J Biol Macromol*, 110, 74-87.
- Qasim, S. B., Husain, S., Huang, Y., Pogorielov, M., Deineka, V., Lyndin, M., . . . Rehman, I. U. (2017). In- vitro and in -vivo degradation studies of freeze gelled porous chitosan composite scaffolds for tissue engineering applications. *Polymer Degradation and Stability*, 136, 31-38.
- Reys, L. L., Silva, S. S., Pirraco, R. P., Marques, A. P., Mano, J. F., Silva, T. H., & Reis, R. L. (2017). Influence of freezing temperature and deacetylation degree on the performance of freeze-dried chitosan scaffolds towards cartilage tissue engineering. *European Polymer Journal*, 95, 232-240.
- Saber-Samandari, S., Saber-Samandari, S., Kiyazar, S., Aghazadeh, J., & Sadeghi, A. (2016). In vitro evaluation for apatite-forming ability of cellulose-based nanocomposite scaffolds for bone tissue engineering. *Int J Biol Macromol*, 86, 434-442.
- Salehpour, S., Rafieian, F., Jonoobi, M., & Oksman, K. (2018). Effects of molding temperature, pressure and time on polyvinyl alcohol nanocomposites properties produced by freeze drying technique. *Industrial Crops and Products*, 121, 1-9.
- Sayed, M., Mahmoud, E. M., Bondioli, F., & Naga, S. M. (2019). Developing porous diopside/hydroxyapatite bio-composite scaffolds via a combination of freeze-drying and coating process. *Ceramics International*, 45(7), 9025-9031.
- Schardosim, M., Soulie, J., Poquillon, D., Cazalbou, S., Duployer, B., Tenailleau, C., . . . Combes, C. (2017). Freeze-casting for PLGA/carbonated apatite composite scaffolds: Structure and properties. *Mater Sci Eng C Mater Biol Appl*, 77, 731-738.
- Schieker, M., Seitz, H., Drosse, I., Seitz, S., & Mutschler, W. (2006). Biomaterials as Scaffold for Bone Tissue Engineering. *European Journal of Trauma*, 32(2), 114-124.
- Shahbazarab, Z., Teimouri, A., Chermahini, A. N., & Azadi, M. (2018). Fabrication and characterization of nanobiocomposite scaffold of zein/chitosan/nanohydroxyapatite prepared by freeze-drying method for bone tissue engineering. *Int J Biol Macromol*, 108, 1017-1027.
- Shazni, Z. A., Mariatti, M., Nurazreena, A., & Razak, K. A. (2016). Properties of

Calcium Phosphate Scaffolds Produced by Freeze-Casting. *Procedia Chemistry*, 19, 174-180. doi:10.1016/j.proche.2016.03.090

- Sofla, M. R. K., Brown, R. J., Tsuzuki, T., & Rainey, T. J. (2016). A comparison of cellulose nanocrystals and cellulose nanofibres extracted from bagasse using acid and ball milling methods. *Advances in Natural Sciences: Nanoscience and Nanotechnology*, 7(3).
- Sommerfeldt, D. W., & Rubin, C. T. (2001). Biology of bone and how it orchestrates the form and function of the skeleton. *Eur Spine J*, 10 Suppl 2, S86-95.
- Sultan, S., & Mathew, A. P. (2018). 3D printed scaffolds with gradient porosity based on a cellulose nanocrystal hydrogel. *Nanoscale*, 10(9), 4421-4431.
- Tabuchi, R., Azuma, K., Izumi, R., Tanou, T., Okamoto, Y., Nagae, T., . . . Anraku, M. (2016). Biomaterials based on freeze dried surface-deacetylated chitin nanofibers reinforced with sulfobutyl ether beta-cyclodextrin gel in wound dressing applications. *Int J Pharm*, 511(2), 1080-1087.
- Vestena, M., Gross, I. P., Müller, C. M. O., & Pires, A. T. N. (2015). Nanocomposite of Poly(Lactic Acid)/Cellulose Nanocrystals: Effect of CNC Content on the Polymer Crystallization Kinetics. *Journal of the Brazilian Chemical Society*.
- Yan, L., Wu, J., Zhang, L., Liu, X., Zhou, K., & Su, B. (2017). Pore structures and mechanical properties of porous titanium scaffolds by bidirectional freeze casting. *Mater Sci Eng C Mater Biol Appl*, 75, 335-340.
- Yang, W., Xu, H., Lan, Y., Zhu, Q., Liu, Y., Huang, S., . . . Guo, R. (2019). Preparation and characterisation of a novel silk fibroin/hyaluronic acid/sodium alginate scaffold for skin repair. *Int J Biol Macromol*, 130, 58-67.
- Ye, M., Mohanty, P., & Ghosh, G. (2014). Morphology and properties of poly vinyl alcohol (PVA) scaffolds: impact of process variables. *Mater Sci Eng C Mater Biol Appl*, 42, 289-294.
- Yılmaz, P., öztürk Er, E., Bakırdere, S., ülgen, K., & özbek, B. (2019). Application of supercritical gel drying method on fabrication of mechanically improved and biologically safe three-component scaffold composed of graphene oxide/chitosan/hydroxyapatite and characterization studies. *Journal of Materials Research and Technology*.
- Youssef Habibi, L. A. L., and Orlando J. Rojas. (2009). Cellulose Nanocrystals: Chemistry, Self-Assembly, and Applications. *Chem. Rev*, 110, 3479–3500.
- Zhang, C., Salick, M. R., Cordie, T. M., Ellingham, T., Dan, Y., & Turng, L. S. (2015). Incorporation of poly(ethylene glycol) grafted cellulose nanocrystals in poly(lactic acid) electrospun nanocomposite fibers as potential scaffolds for bone tissue engineering. *Mater Sci Eng C Mater Biol Appl*, 49, 463-471.
- Zhang, H., Nie, H., Li, S., White, C. J. B., & Zhu, L. (2009). Crosslinking of electrospun polyacrylonitrile/hydroxyethyl cellulose composite nanofibers. *Materials Letters*, 63(13-14), 1199-1202.

- Zhang, J., Zhou, A., Deng, A., Yang, Y., Gao, L., Zhong, Z., & Yang, S. (2015). Pore architecture and cell viability on freeze dried 3D recombinant human collagen-peptide (RHC)-chitosan scaffolds. *Mater Sci Eng C Mater Biol Appl*, 49, 174-182.
- Zhou, X. H., Wei, D. X., Ye, H. M., Zhang, X., Meng, X., & Zhou, Q. (2016). Development of poly(vinyl alcohol) porous scaffold with high strength and well ciprofloxacin release efficiency. *Mater Sci Eng C Mater Biol Appl*, 67, 326-335.
- Zulkifli, F. H., Hussain, F. S., Rasad, M. S., & Mohd Yusoff, M. (2014). Nanostructured materials from hydroxyethyl cellulose for skin tissue engineering. *Carbohydr Polym*, 114, 238-245.
- Zulkifli, F. H., Hussain, F. S. J., Harun, W. S. W., & Yusoff, M. M. (2019). Highly porous of hydroxyethyl cellulose biocomposite scaffolds for tissue engineering. *Int J Biol Macromol*, 122, 562-571.