

THE STUDY ON PLANT HORMONES AND
SPENT MUSHROOM COMPOST ON THE
GROWTH OF *Ficus carica*

SITI HAMIDAH RADIYAH BINTI SHIEKH
MAHMUD

MASTER OF SCIENCE

UNIVERSITI MALAYSIA PAHANG



SUPERVISOR'S DECLARATION

I hereby declare that I have checked this thesis and in my opinion, this thesis is adequate in terms of scope and quality for the award of the degree of Master of Science.

(Supervisor's Signature)

Full Name : ASSOC. PROF. DR. TAN SUAT HIAN

Position : ASSOCIATE PROFESSOR

Date :



STUDENT'S DECLARATION

I hereby declare that the work in this thesis is based on my original work except for quotations and citations which have been duly acknowledged. I also declare that it has not been previously or concurrently submitted for any other degree at Universiti Malaysia Pahang or any other institutions.

(Student's Signature)

Full Name : SITI HAMIDAH RADIYAH BINTI SHIEKH MAHMUD

ID Number : MKT16002

Date :

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SITI HAMIDAH RADIYAH BINTI SHIEKH MAHMUD

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ABSTRAK

Ficus carica (*F. carica*) atau pokok ara dari keluarga mulberi (Moraceae) dikenali sebagai 'buah tin' di kalangan rakyat Malaysia tinggi permintaannya dari bidang farmaseutikal. Oleh kerana nilai nutrisinya. Objektif penyelidikan ini adalah untuk mengetahui teknik pensterilan dan kepekatan pengatur pertumbuhan tanaman dan penggunaan SMC yang optimum. Penanaman *in vitro* tanaman ara di bawah persekitaran terkawal diperkenalkan untuk memenuhi permintaan penyebaran besar bahan penanaman berkualiti tinggi dan kompos cendawan *Pleurotus ostreatus* (SMC) yang merupakan substrat sisa digunakan sebagai pembekal nutrien alternatif. Pada penanaman *in vitro* tanaman ara, Ujian fitokimia dan spektroskopi Fourier transformasi inframerah (FTIR) juga dijalankan untuk mengenal pasti kumpulan metabolit sekunder yang berkaitan dengan tumbuhan ini yang dapat memberi manfaat kepada bidang perubatan. Bahagian pertama dalam penanaman *in vitro* tanaman ara adalah penghasilan anak pokok aseptik melalui proses pensterilan. Teknik pensterilan terbaik adalah rawatan 3 (T3) 70% etanol selama 10 minit, 30% clorox ditambahkan dengan dua titis Tween 20 selama 10 minit dan 0.01% perak asetat selama 5 minit. Bahagian kedua adalah induksi pucuk dan akar yang dilakukan dalam berbagai jenis media yang dilengkapi dengan pengatur pertumbuhan tanaman (PGR), SMC dan gabungan PGR dan SMC. Proliferasi tunas tertinggi dengan adanya hormon tumbuhan dicapai pada 15 μM Kinetin. Untuk percambahan akar, bilangan akar tertinggi terdapat pada kultur 30 μM IAA. Untuk penanaman *in vitro* di media yang dilengkapi dengan SMC, jumlah pucuk tertinggi diperoleh pada media SMC 10%. Walau bagaimanapun, akar baru gagal berkembang di media yang dilengkapi dengan SMC sahaja. PGR (15 μM Kinetin dan 30 μM IAA) dan SMC (10%) kemudian digabungkan dengan proliferasi pucuk tertinggi dalam media gabungan 15 μM Kinetin dengan SMC 10% dengan bilangan percambahan maksimum yang dicapai (3.00 ± 1.27) dengan panjang pucuk $0.23 \text{ cm} \pm 0.08$. Bilangan maksimum akar (0.50 ± 0.84) dengan panjang akar 0.10 ± 0.16 dicapai dalam kultur 30 μM IAA + 10% SMC. Ujian fitokimia yang dilakukan untuk ekstrak daun *F. carica* menunjukkan adanya kumpulan metabolit sekunder yang berfungsi seperti amina, aromatik, nitril, karbonil, alkena, nitro dan alkil halide selari dengan keputusan FTIR. Kesimpulannya, penggunaan pengatur pertumbuhan tanaman dan SMC yang optimum memungkinkan untuk mempromosikan tunas dan akar baru untuk kultur nodal. SMC dapat berinovasi dari bahan buangan untuk menyediakan karbon dan nutrien dan meminimumkan penggunaan hormon tumbuhan dalam mikropropagasi tanaman ara.

ABSTRACT

Ficus carica (*F. carica*) or fig tree from mulberry family (Moraceae) known as ‘buah tin’ among Malaysians highly demand from pharmaceutical area for its nutritional value. The objective of this research is to determine the optimize sterilization technique and concentrations of plant growth regulator and SMC use. *In vitro* culture of fig plant under controlled environment was introduced in fulfilling the demand for mass propagation of high quality planting material and *Pleurotus ostreatus* spent mushroom compost (SMC) which is a waste substrate was used as an alternative nutrient supplier. Upon *in vitro* culture of fig plant, phytochemical test and Fourier transform infrared spectrophotometer (FTIR) analysis were also conducted to highlight the important functional groups of secondary metabolites related to this plant that can give benefits to medicinal area. The first part is production of aseptic plantlets through sterilization process for *in vitro* fig’s culture. The best sterilization technique was treatment 3 (T3) of 70% ethanol for 10 minutes, 30% clorox added with two drops of Tween 20 for 10 minutes and 0.01% silver acetate for 5 minutes. The second part is the shoots and root inductions conducted in different types of media that supplemented with plant growth regulator, SMC and combination of PGR and SMC. The highest shoot proliferations in presence of plant hormone were achieved at 15 μM Kinetin. For roots proliferation, the highest number of root was found in 30 μM IAA cultures. For media supplemented with SMC, the highest number of shoots was obtained in 10% SMC media. However, new roots were failed to grow in media supplemented with SMC only. PGR (15 μM Kinetin and 30 μM IAA) and SMC (10%) were combine with highest proliferations of shoots in combination media were 15 μM Kinetin with 10% SMC media with maximum number of shoot proliferation achieved (3.00 ± 1.27) and length of shoots ($0.23 \text{ cm} \pm 0.08$). The maximum number of root (0.50 ± 0.84) and length of root (0.10 ± 0.16) was achieved in 30 μM IAA + 10% SMC culture. The phytochemical test conducted for extracts of *F.carica* leaves indicates the presence of functional group of secondary metabolites such as amine, aromatic, nitrile, carbonyl, alkene, nitro and alkyl halide which is align with FTIR result. In conclusion, optimal use of plant growth regulator and SMC enables the promoting of new shoots and roots for fig nodal culture. SMC can be innovate from waste material to provide carbon and nutrient and minimizes the usage of plant hormone in micropropagation of fig plant.

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REFERENCES

- Abbas, H., & Qaiser, M. (2012). *In vitro* response of *Ruellia linearibracteolata* to different growth hormones- an attempt to conserve an endangered species. Pak. J. Bot, 791-794. [http://www.pakbs.org/pjbot/PDFs/44\(2\)/49.pdf](http://www.pakbs.org/pjbot/PDFs/44(2)/49.pdf)
- Abraham, A., Afewerki, B., Tsegay, B., Ghebremedhin, H., Teklehaimanot, B., & Reddy, K.S. (2018). Extraction of agar and alginate from marine seaweeds in red sea region. International Journal of Marine Biology and Research, 3(2), 1-8. <http://dx.doi.org/10.15226/24754706/3/2/00126>
- Afaf, M., Afaf, A., & Mohamed, A. H. (2015). Evaluation of antioxidant and antimicrobial potential of different leaves crude extracts of Omani *Ficus carica* against food borne pathogenic bacteria. Asian Pacific Journal of Tropical Disease, 13-16. [https://doi.org/10.1016/S2222-1808\(14\)60619-8](https://doi.org/10.1016/S2222-1808(14)60619-8)
- Afagh, H.V., Saadatmand, S., Riahi, H., & Nejad, R.A.K. (2019). Influence of spent mushroom compost (SMC) as an organic fertilizer on nutrient, growth, yield, and essential oil composition of german chamomile (*Matricaria Recutita* L.). Journal of Communications in Soil Science and Plant Analysis, 538-548. <https://doi.org/10.1080/00103624.2019.1568450>
- Ahmad, S., Abdel-Salam, N.M., & Ullah, R. (2016). *In vitro* antimicrobial bioassays, DPPH radical scavenging activity, and FTIR spectroscopy analysis of *Heliotropium bacciferum*. Biomed Research International, 12pp. <https://doi.org/10.1155/2016/3818945>
- Aljane, F., & Nahdi, S. (2014). Propagation of some local fig (*Ficus carica* L.) cultivars by hardwood cuttings under the field conditions in Tunisia. International Scholarly Research Notices, 5 pp. <http://doi.org/10.1155/2014/809450>
- Ali, B., Mujeeb, M., Aeri, V., Mir, S. R., Faiyazuddin, M., & Shakeel, F. (2012). Anti-inflammatory and antioxidant activity of *Ficus carica* Linn, leaves. Natural Product Research, 26(5), 460-465. <http://doi.org/10.1080/14786419.2010.488236>
- Altaf, H., Qarbi, I. A., & Hummera, N. (2012). Plant tissue culture: current status and opportunities. Intech Open. <http://doi.org/10.5772/50568>
- Altman, A., & Loberant, B. (1997). Micropropagation: clonal plant propagation *in vitro*. In: Agricultural Biotechnology, Marcel Dekker, Inc., 19-42. <http://doi.org/isbn:-0-8247-9439-7>
- Anuar, W.N.H.W., Tan, S.H., Mahmud, S.H.R.S., Norazmi, W.N.M., & Safruddin, E.F.S.E. (2019). Enhancement effect of BAP and spent mushroom compost in micro-propagation of sabah snake grass. American Journal of Biochemistry and Biotechnology, 15 (4), 190-197. <http://doi.org/10.3844/ajbbsp.2019.190.197>
- Ashokkumar, R., & Ramaswamy, M. (2014). Phytochemical screening by ftir spectroscopic analysis of leaf extracts. International Journal of Current Microbiology and Applied Sciences, 3(1), 395-406. <http://doi.org/issn-2319-7706>

- Avneesh, D. S., Abdullah, N., & Vikineswary, S. (2003). Optimization of extraction of bulk enzymes from spent mushroom compost. *Journal of Chemical Technology and Biotechnology*, 743-752. <http://doi.org/10.1002/jctb.852>
- Barolo, M., Ruiz, M., & Lopez, S. (2014). *Ficus carica* L. (Moracea): an ancient source of food and health. *Food Chemistry*, 119-127. <http://doi.org/10.1016/j.foodchem.2014.04.112>.
- Beyl, C. A., & Trigiano, R. N. (2015). *Plant propagation concepts and laboratory exercises*. CRC Press. <http://doi.org/isbn-9781466503878>
- Bhatia, S., Sharma, K., Dahiya, R., & Bera, T. (2015). *Modern applications of plant biotechnology in pharmaceutical sciences*. Academic Press, 439. <http://doi.org/isbn-9780128022214>
- Biswas, K.K., Foster, A.J., Aung, T., & Mahmoud, S.S. (2009). Essential oil production : relationship with abundance of glandular trichomes in aerial surface of plants. *Acta Physiol Plant*, 31(1), 13-19. <http://doi.org/10.1007/s11738-008-0214-y>
- Borah, A., Anbumalarmathi, J., & Aruna, S. (2019). *In vitro* propagation of *Coccinia indica* (L.)Voight from internodal segments. *Indian Journal of Agriculture Research*, 53, 202-207. <http://doi.org/10.18805/IJARE.A-5020>
- Bouhouce, N., & Ksiksi, T.(2007). An efficient *in vitro* plant regeneration system for medicinal plant *Teucrium stocksianum* Boiss. *Plant Biotechnology Report*, 179-184. <https://doi.org/10.1007/s11816-007-0033-4>
- Buck, G.B., Castro, G.F.D., Mattiello, E., & Zotarelli, L. (2020). Applications of gypsum and ammonium sulfate change soil chemical properties of a salt-affected agricultural soil. *Journal of Agricultural Science*, 12(7). <http://doi.org/10.5539/jas.v12n7p1>
- Chawla, H. S. (2002). *Introduction to Plant Biotechnology: Second Edition*. Science Publisher. <http://doi.org/isbn-1578082285>
- Chithiraichelvan, R., Kurian, R.M., Awachare, C.M., & Laxman, H. (2017).Performance of fig (*Ficus carica* l.) under different planting densities.*International Journal of Current Microbiology and Applied Sciences*,6(6):2603-2610. <http://doi.org/10.20546/ijcmas.2017.606.311>
- Chun, C. & Kozai, T. (2001). A closed-type transplant production system. *Progress in Biotechnology*, 18, 375-384. [http://doi.org/10.1016/S0921-0423\(01\)80094-0](http://doi.org/10.1016/S0921-0423(01)80094-0)
- Condit, I.J. (1955). Fig varieties: a monograph . *Journal of Agricultural Science*, 323-511.<http://doi.org/10.3733/hilg.v23n11p323>
- Cristie, J.A., Kozai, T., & Smith, M.A.L. (1995). *Automation and environmental control in plant tissue culture*. Springer -Science + Business Media. <http://doi.org/10.1007/978-94-015-8461-6>
- Crisosto, H., Ferguson, L., Bremer, V., Stover, E., & Colleli, G. (2011). *Postharvest biology and technology of tropical and subtropical fruits*. Woodhead Publishing Series in Food Science, Technology and Nutrition, 134-158. [isbn/9780857092885](http://doi.org/isbn/9780857092885)

- Cullen, M., T. Electronic sources: Food and agriculture organization of the United Nations (FAOSTAT). <http://www.fao.org/faostat/en/#data/TP> (20/10/2020)
- Danial, G., Ibrahim, D., Brkat, S., & Khalil, B. (2014). Multiple shoots production from shoot tips of fig tree (*Ficus carica* L.) and callus induction from leaf segments. *International Journal of Pure and Applied Sciences and Technology*, 20(1), 117-124. <http://doi.org/ISSN 2229 - 6107>
- Daud, N., Shashita, J., & Rozi, M. (2012). An improved surface sterilization technique for introducing leaf, nodal and seed explant of *Aquilaria malaccensis* from field sources into tissue culture. *Journal of Molecular Biotechnology*, 20(2), 55-58. <http://doi.org/issn-0128-7451>
- Darwesh, H., Bazaid, S., & Samra, B. (2014). *In vitro* propagation method of *Ficus carica* at Taif governorate using tissue culture technique. *International Journal of Advanced Research*, 756-761. <http://doi.org/issn-2320-5407>
- Debnath, M., Malik, C.P., & Bisen, P.S. (2006). Micropropagation: a tool for the production of high quality plant-based medicines. *Current Pharmaceutical Biotechnology*, 7, 33-49. <https://doi.org/10.2174/138920106775789638>
- Dessoky, E., Attia, A., & Mohamed, E. (2016). An efficient protocol for *in vitro* propagation of fig (*Ficus carica* sp) and evaluation of genetic fidelity using RAPD and ISSR markers. *Journal of Applied Biology & Biotechnology*, 4(4),57-63. <http://doi.org/10.7324/JABB.2016.40406>
- Djilianov, D., Genova, G., Parvanova, D., Zapryanora, N., Konstantinova, T., & Atanassov, A. (2005). *In vitro* culture of resurrection plant *Haberlea rhodopensis*. *Plant Cell, Tissue and Organ Culture*, 115-118. <https://doi.org/10.1007/s11240-004-8835-3>.
- Dolgun, O., & Tekintas, F.E. (2008). Production of fig (*Ficus carica* L.) nursery plants by stem layering method. *Agriculturae Conspectus Scientificus*, 73(3), 157-160. <http://doi.org/issn-1331-7776>
- Ebrahimi, E., Asadi, G., & von Fragstein und Niemsdorff, P. (2019). A field study on the effect of organic soil conditioners with different placements on dry matter and yield of tomato (*Lycopersicon esculentum* L.). *International Journal of Recycling of Organic Waste in Agriculture*, 59-66.<http://doi.org/10.1007/s40093-018-0228-4>
- Fabiano, C.C., Tezotto, T., Favarin, J.L., Polacco, J.C., & Mazzafera, P. (2015). Essentiality of nickel in plants: a role in plant stresses, *Frontier Plant Science*. <https://doi.org/10.3389/fpls.2015.00754>
- Figueiredo, A. C., Barroso, J.G., Pedro, L.G., & Scheffer, J.C. (2008). Factors affecting secondary metabolites production in plants: volatile components and essential oils. *Flavour & Fragrance Journal*, 23, 213-226. <http://doi.org/10.1002/ffj.1875>
- Flaishman, M., Rodov, V., & Stover, E. (2008). The Fig: Botany, Horticulture and Breeding. *Horticulture reviews*, 113-196.<http://doi.org/10.1002/9780470380147.ch2>
- Fleming, F.F., Yao, L., Ravikumar, P.C., Funk, L., & Shook, B.C. (2011). Nitrile-containing pharmaceuticals: efficacious roles of the nitrile pharmacophore. *Medical Chemistry* 53(22), 7902-7917. <https://doi.org/10.1021/jm100762r>

- Foo, P. C., Lee, Z. H., Chin, C. K., Subramaniam, S., & Chew, B. L. (2018). Shoot induction in white eggplant (*Solanum melongena* L. Cv. Bulat putih) using 6-Benzylaminopurine and Kinetin. *Tropical Life Sciences Research*, 119–129. <https://doi.org/10.21315/tlsr2018.29.2.9>
- Gál, B., Bucher, C., & Burns, N.Z. (2016). Chiral alkyl halides: underexplored motifs in medicine. *Mar Drugs*, 14(11), 206. <http://doi.org/10.3390/md14110206>
- George, E. F., Hall, M. A., & Klerk, G. D. (2007). The components of plant tissue culture media I: macro- and micro-nutrients. *Plant propagation by tissue culture 3rd edition*, Netherlands: Springer, 1-28. <http://doi.org/isbn-978-1-4020-5005-3>
- George, P., & Manuel, J. (2013). Low cost tissue culture technology for the regeneration of some economically important plants for developing countries. *International Journal of Agricultural, Environment and Biotechnology*, 6, 703-711. <http://doi.org/issn-2230-732X>
- Gharbia, H. D., Diaa, A. I., Sazan, A. B., & Belan, M. K. (2014). Multiple shoots production from shoot tips of fig tree (*Ficus carica* L.) and callus induction from leaf segment. *International Journal of Pure Sciences and Technology*, 117-124. <http://doi.org/212500233>
- Gilani, A., Mehmood, M., Janbaz, K., Khan, A., & Saeed, S. (2008). Ethnopharmacological studies on antispasmodic and antiplatelet activities of *Ficus carica*. *Journal of Ethnopharmacology*, 119(1), 1-5. <http://doi.org/10.1016/j.jep.2008.05.040>
- Hassanpour, S., Maheri-Sis, N., Eshratkhah, B., & Mehmandar, F. B. (2011). Plants and secondary metabolites (Tannins): a review. *International Journal of Forest, Soil and Erosion*, 47-53. <http://www.ijfse.com/issn/2251-6387>
- Hasanuzzaman, M., Bhuyan, M.H.M.B., Nahar, K., Hossain, M.S., Mahmud, J.A., Hossen, M.S., Masud, A.A.C., Moumita, & Fujita, M. (2018). Potassium: a vital regulator of plant responses and tolerance to abiotic stresses. *Agronomy*, 8 (31). <http://doi.org/10.3390/agronomy8030031>
- Hameed, E. (2009). Total phenolic contents and free radical scavenging activity of certain egyptian *Ficus* species leaf samples. *Food Chemistry*, 114(4), 1271-1277. <http://doi.org/10.1016/j.foodchem.2008.11.005>
- Han, G.K., Sang, H.P., Hyuk, S. H., Seong, H.K, Hyuk, G.P., & Won, M. (2005). Detection and recovery of hydrolytic enzymes from spent compost of four mushroom species. *Folia Microbiol*, 103-106. <https://doi.org/10.1007/bf02931456>
- He, X. P., Zeng, Y., Zang, Y., Li, J., Field, R. A., & Chen, G. (2016). Carbohydrate CuAAC click chemistry for therapy and diagnosis. *Carbohydrate Research*, 429, 1-22. <https://doi.org/10.1016/j.carres.2016.03.022>
- Hepler, P.K. (2005). Calcium: a central regulator of plant growth and development. *The Plant Cell*, 17, 2142-2155. <https://doi.org/10.1105/tpc.105.032508>
- Hiru, R., & Kapoor, R. (2018). *Plant biotechnology. India: Woodhead Publishing in Agriculture.* <http://doi.org/isbn-9789385059339>

- Hussain, A., Qarshi, I.A., Nazir, H., & Ullah, I. (2012). Plant tissue culture: current status and opportunities. IntechOpen, <http://doi.org/10.5772/50568>
- Jan, A., Bhat, K.M., Bhat, S.J.A., Mir, M.A., Wani, I.A., & Rather, J.A. (2013). Surface sterilization method for reducing microbial contamination of field grown strawberry explants intended for *in vitro* culture. African Journal of Biotechnology, 12(39), 5749-5753. <http://doi.org/10.5897/AJB2013.12918>
- Jani, J., Jha, S., & Nagar, D. (2006). Root explant produces multiple shoot from pericycle in *Psoralea corylifolia*-a leprosy destroyer medicinal plant. Science Horticulturae, 332-336. <http://doi.org/10.1016/j.indcrop.2015.02.001>
- Jani, N., Qazi, H. A., Ramzan, S. (2018). Developing stress-tolerant plants through *in vitro* tissue culture: family Brassicacea. Biotechnologies of Crop Improvement, 1, 327-372. http://doi.org/10.1007/978-3-319-78283-6_10
- Jasińska, A. (2018). Spent mushroom compost (SMC) – retrieved added value product closing loop in agricultural production. Acta Agraria Debreceniensis, 185-202. <https://doi.org/10.34101/actaagrar/150/1715>
- Jha, T., & Ghosh, B. (2005). Plant Tissue Culture Basic and Applied. Orient Blackswan. <http://doi.org/isbn-8173714886>
- Jordan, S. N., Mullen, G. J., & Murphy, M. C. (2008). Composition variability of spent mushroom compost in Ireland. Bioresource technology, 411-418. <http://doi.org/10.1016/j.biortech.2006.12.012>
- Kamarudzaman, A. N., Tay, C. C., Ab Jalil, M. F., & Abdul Talib, S. (2013). Biosorption of iron(III) from aqueous solution using *Pleurotus ostreatus* spent mushroom compost as biosorbent. Advanced Materials Research, 781-784. <https://doi.org/10.4028/www.scientific.net/AMR.781-784.636>
- Karimian, R., Lahouti, M., & Davarpanah, S. (2014). Effects of different concentrations of 2-4-D and Kinetin on callogenesis of *Taxus Brevifolia* Nutt. Journal of Applied Biotechnology Reports, 1 (4), 167-170. http://www.biotechrep.ir/article_69158_81453a8132c66e99d0e8c6d6f96dd8cc.pdf
- Kavand, S., Kermani, M.J., Haghazari, A., Khosravi, P., & Azimi, M.P. (2011). Micropropagation and medium-term conservation of *Rosa pulverulenta*. Acta Scientiarum, Agronomy, 33(2), 297-301. <http://doi.org/10.4025/actasciagron.v33i2.10279>
- Kieber, J.J., & Schaller, G.E. (2014). Cytokinins. The Arabidopsis Book, 12. <http://doi.org/isbn-10.1199/tab.0168>.
- Krishna, H., Alizadeh, M., Singh, D., Chauhan, N., Eftekhari, M., & Sadh, R. K. (2016). Somaclonal variations and their applications in horticultural crops improvement. <http://www.connectjournals.com/bca/ issn 0972-5075>
- Kwagyan, M.W., & Odamtten, G.T. (2018). Use of *Pleurotus eous* Strain P-31 Spent Mushroom Compost (SMC) as Soil Conditioner on the Growth and Yield Performance of *Capsicum annuum* L. and *Solanum lycopersicon* L. Seedlings under Greenhouse Conditions in Ghana. Tropical Science Research, 29(1), 173-194.

<http://doi.org/10.21315/tlsr2018.29.1.12>

- Ling, W., Liew, F. C., Lim, W. Y., Subramaniam, S., & Chew, B. L. (2018). Shoot induction from axillary shoot tip explants of fig (*Ficus carica*) cv. Japanese BTM 6. *Tropical Life Sciences Research*, 165–174. <https://doi.org/10.21315/tlsr2018.29.2.11>
- Mahna, N., Vahed, S. Z., & Khani, S. (2013). Plant *In vitro* Culture goes Nano: Nanosilver-Mediated Decontamination of *Ex vitro* Explants. *J Nanomed Nanotechol*, 4,161. doi:10.4172/2157-7439.1000161
- Malhotra, H., Vandana, Sharma, S., & Pandey, R.M. (2018). Phosphorus nutrition: plant growth in response to deficiency and excess, *Plant Nutrients and Abiotic Stress Tolerance*, 171-190. http://doi.org/10.1007/978-981-10-9044-8_7
- Mardiyani, S., Murwani, I., & Lestari, M. W. (2019). The effect of spent mushroom compost and various composting starter combination on the growth and yield of kangkong (*Ipomoea reptans*). *International Conference on Green Agro-industry and Bioeconomy 2018 ,IOP Conference Series: Earth and Environmental Science*, 23(1) <http://doi.org/10.1088/1755-1315/230/1/012023>
- Martin, K.P. (2003). Rapid *in vitro* multiplication and *ex vitro* rooting of *Rotula aquatica* Lour., a rare rheophytic woody medicinal plant. *Plant Cell Rep*, 21, 415-420. <http://doi.org/10.1007/s00299-002-0547-8>
- Marschner, H. (1995). *Mineral Nutrition of Higher Plant: Second Edition*. United State of America: Academic Press. <http://doi.org/isbn-9780080571874>
- Mawa, S., Husain, K., & Jantan, I.(2013). *Ficus carica* L. (Moraceae): Phytochemistry, traditional uses and biological activities. *Evidence-Based Complementary and Alternative Medicine*, 1-8. <https://http://doi.org/10.1155/2013/974256>
- Melisa, I. B., Natalie, R. M., & Silvia, N. L. (2014). *Ficus carica* L.(Moracea): An ancient source of food and health. *Food Chemistry*, 119-127. <http://doi.org/10.1016/j.foodchem.2014.04.112>.
- Michalak, I., Saied, A., Chojnacka, K.W., & Gramza, M. (2018). Trace elements as fertilizer micronutrients, *Recent Advances in Trace Elements*, 299-138.<http://doi.org/isbn-978-1-119-13380-3>
- Miret, J.A., & Bosch, S.M. (2014). Plant amino acid-derived vitamins: biosynthesis and function, *Amino Acids*, 46 (4), 809-824. <http://doi.org/10.1007/s00726-013-1653-3>.
- Miri, S.M., & Roughani, A. (2018). Factors affecting tissue culture success in ornamental crops, 2nd international & 3rd National Congress on Flower and Ornamental Plant, 39,At Mahallat, Iran
- Mohamed, A.M.,Sekar, S., & Vincent, S. (2010). Plant growth substances in crop production: a review. *Asian Journal of Plant Science*, 9(4), 215-222. <http://dx.doi.org/10.3923/ajps.2010.215.222>
- Moses,P. (1987). Gene transfer method applicable to agricultural organism. *Agricultural biotechnology: strategies for national competitiveness*. National Academy Press, 149-192. <http://doi.org/isbn-0-309-0-3745-x>

- Motohiro, O. Haruhiko, O., Yuji, Y. (2019). Use of corticosteroids for remission induction therapy in patients with new-onset ulcerative colitis in real-world settings. *Journal of Market Access and Health Policy*, 7(1).
<http://doi.org/10.1080/20016689.2019.1565889>
- Munajad, A., Subroto, C., & Suwarno (2018). Fourier transform infrared (ftir) spectroscopy analysis of transformer paper in mineral oil-paper composite insulation under accelerated thermal aging. *Energies*, 11, 364. <http://doi.org/10.3390/en11020364>
- Nadia, A. O., Salim, O., Nassima, M., Yamina, I., Dolores, H., & Carla, M. S., (2017). Bioactive metabolites involved in antioxidant anticancer and anticalpain activities of *Ficus carica* L., *Ceratonia siliqua* L. and *Quercus ilex* L. extracts. *Industrial Crops and Production*, 6-17.
<https://doi.org/10.1016/j.indcrop.2016.10.007>
- Nakhooda, M., Watt, M. P., & Mycock, D. (2013). The choice of auxin analogue for *in vitro* root induction influences post-induction root development in *Eucalyptus grandis*. *Turkish Journal of Agriculture and Forestry*, 258-266. <http://doi.org/10.3906/tar-1302-110>
- Nasution, N.H., & Nasution, S.M. (2019). The effect of plant growth regulators on callus induction of mangosteen (*Garcinia mangostana* L.). The 4th International Conference of Biological Sciences and Biotechnology, Conference Series; Earth and Environmental Science, 305, 10p.<http://doi.org/10.1088/1755-1315/305/1/012049>
- Naqvi, S. S., Sultana, R. A., & Rasheed, H. A. (2005). Tissue culture studies in *Oryza sativa* L. cvs. Basmati 385 and Super Basmati. *Pak. J. Bot.* 823-828.
<https://pdfs.semanticscholar.org/70c8/04687178b43e49a8ca5fe377de9268c83df7.pdf>
- Nepali, K., Lee, H.Y., & Liou, J.P. (2018). Nitro group containing drugs. *Journal of Medicinal Chemistry*, 62(6). <http://doi.org/10.1021/acs.jmedchem.8b00147>
- Ncube, B., & Staden, J.V. (2015). Tilting plant metabolism for improved metabolite biosynthesis and enhance human benefits. *Molecules*, 20(7), 12698-12731.
<http://doi.org/10.3390/molecules200712698>
- Nhut, D.S., Silva, J. A. T. D. & Aswath, C.R. (2003). The importance of the explant of regeneration in thin cell layer technology. *In vitro Cell Development of Biological Plant*, 39, 266-276. <https://doi.org/10.1079/IVP2002408>
- Noureddine, B. (2018). Pharmacological activity of alkaloids:a review. *Asian Journal of Botany*, Volume 1. <http://doi.org/10.63019/ajb.v1i2.467>
- Orhan, I. (2012). Biotechnology production of plant secondary metabolites. Bentham e Books Publisher.<https://doi.org/10.2174/97816080511441120101>
- Ortuno, A., Diaz, L., Perez, I., Sanchez, F., & Rio, J. (2018). Biological active compounds from *Limonium insigne* and alternative methods for its micropropagation. Elsevier Publisher, 78-85. <http://doi.org/10.1016/j.scienta.2017.11.022>
- Oseni, O.M., Pande, V., & Nailwal, T.K. (2018). A review on plant tissue culture, a technique for propagation and conservation of endangered plant species. *International Journal of Current Microbiology and Applied Sciences*, 7(7), 3778-3786.

<https://doi.org/10.20546/ijcmas.2018.707.438>

- Ozlem, G., & Giuseppe. M. (2007). Saponins: properties, applications and processing. *Critical Reviews in Food Science and Nutrition*, 231-258.
<https://doi.org/10.1080/10408390600698197>
- Pagare,S., Bhatia, M., Tripathi, N., Pagare, S. & Bansal, Y.K. (2015). Secondary metabolites of plant and their role: overview. *Current Trends in Biotechnology and Pharmacy*, 9(3), 293-304. <http://doi.org/issn-0973-8916>
- Palacio, L., Cantero, J. J., Cusidó, R., & Goleniowski, M. (2011). Phenolic compound production by *Larrea divaricata* Cav. plant cell cultures and effect of precursor feeding. *Process Biochemistry*, 418-422. <http://doi.org/10.1016/j.procbio.2010.08.029>
- Panda, H. (2013). Handbook on ayurvedic medicines with formulae, processes, & their use, 2nd revised edition. Niir Project Consultancy Series, <http://doi.org/isbn-978-93-81039-25-0>
- Pakkirisamy, M., Kalakandan, S.K., & Ravichandran, K. (2017). Phytochemical screening, gc-ms, ft-ir analysis of methanolic extract of *Curcuma caesia* Roxb (black turmeric). *Pharmacogn J.*,9(6), 952-956. <http://doi.org/10.5530/pj.2017.6.149>
- Pessarakli, M., (2001). Handbook of Plant and Crop Physiology, 2nd ed. Revised and Expanded. Marcel Dekker, Inc., New York, 973.<http://doi.org/isbn-0-8247-0546-7>
- Puliamackal,A.J., Kareem, A. V. K., Durgaprasad,K., Trivedi, Z.B. & Prasad, K. (2014).Competence and regulatory interactions during regeneration in plants, *Plant Science*. <https://doi.org/10.3389/fpls.2014.00142>
- Rameshkumar, K.B. (2012). Phytochemistry- the fascinating chemistry of plants. *India Science*, 15, 10-18. https://www.researchgate.net/publication/299435971_Phytochemistry-_The_Fascinating_Chemistry_of_Plants
- Rashid, A., Khan, A.A., Dar, S.H., Ahmad, Y., Nabi, N.G., Ganaie, M.A., & Teli, A.R. (2017). Extraction, isolation, and spectral analysis of the psoralen compound from *Ficus carica* Linn. leaves. *The Pharma Innovation Journal*, 740-743.<http://ww.thepharmajournal.com/issn/2277-7695>
- Razdan, M.K. (2002). Introduction to Plant Tissue Culture: Second Edition. Science Publisher. <http://doi.org/812041571X>
- Sarkar, A. (2009).Plant stem cell, encyclopedia of stem cells. Discovery Publishing House. https://www.abebooks.com/isbn/9788183564076/30249023374&cm_sp=snippet_-_srp1_-_PLP1
- Sathyanarayana, B. N., & Varghese, D. B. (2007). Plant tissue culture. Practices and New Experimental Protocols, IK International Publishing Hse. Pvt. Ltd.,318pp. <http://doi.org/isbn-9788189866112>
- Sathyagowri, S., & Seran, T.H. (2011). *In vitro* plant regeneration of ginger (*Zingiber officinale* Rosc.) with emphasis on initial culture establishment. *International Journal Medical Aromatic Plants*, 1(3), 195-202. <http://www.openaccessscience.com/issn-2249-4340>

- Selvakumar, V., Anbudural, P. R., & Balakumar, T. (2000). *In vitro* propagation of the medicinal plant *Plumbago zeylanica* L. through nodal explants. *Society for In Vitro Biology*, 280-284. <https://doi.org/10.1007/s11627-001-0050-x>
- Sharma, M., Rashid, A., Ahmad, Y., Ganaie, M., Bashir, M., & Tripathi, J. (2017). Extraction, isolation and spectral analysis of the psoralen compound from *Ficus carica* Linn. leaves. *Der Pharma Chemical*, 9(6), 6-10. <http://www.derpharmachemica.com/archive.html>
- Shomali, I., Sadder, M., & Ateyyeha, A. (2017). Culture media comparative assessment of common fig (*Ficus carica* L.) and carryover effect. *Jordan Journal of Biological Sciences*, 10(1), 13-18. <http://doi.org/issn-1995-6673>
- Singh, A., Saha, S., & Singh, R. Electronic sources: Fresh figs market, Future Market Insight. <https://www.futuremarketinsights.com/reports/fresh-figs-market> (20/10/2020)
- Singh, V., Tyagi, A., Chauhan, P. K., Kumari, P., & Kaushal, S. (2011). Identification and prevention of bacterial contamination on explant used in plant tissue culture labs. *International Journal of Pharmacy & Pharmaceutical Sciences*, 3(4). <http://doi.org/issn-0975-1491>
- Sinha, K. K. (2003). *Encyclopedia of food science and nutrition* (second edition). Academic Press, 2394-2399. [isbn/9780080917917](https://doi.org/10.1016/B978-0-12-397179-1.00179-7)
- Silva, J.A., & Uchida, R.A. (2000). Plant nutrient management in Hawaii's soils, approaches for tropical and subtropical agriculture, *College of Tropical Agriculture and Human Resources*, 4-6. <http://doi.org/isbn-1-929235-08-8>
- Smith, A.C., Yardley, V., Rhodes, J., & Croft, S.L. (2000). Activity of the novel immunomodulatory compound tucaresol against experimental visceral Leishmaniasis. *Antimicrobial Agents and Chemotherapy*, 44(6), 1494–1498. <http://doi.org/10.1128/aac.44.6.1494-1498.2000>
- Soni, N., Mehta, S., Satpathy, G., & Gupta, R. K. (2014). Estimation of nutritional phytochemical, antioxidant and antibacterial activity of dried fig (*Ficus carica*). *Journal of Pharmacognosy and Phytochemistry*, 3(2), 158-165. <http://doi.org/issn-2278-4136>
- Stewart, D.P., Cameron, K. C., Cornforth, I. S., & Main, B.E. (1998). Release of sulphate, potassium, calcium and magnesium from spent mushroom compost under laboratory conditions. *Biology Fertile Soils*, 146-151. <https://doi.org/10.1007/s003740050358>
- Stigter, K.A., Plaxton, W.C. (2005). Molecular mechanisms of phosphorus metabolism and transport during leaf senescence, *Journal of National Institute of Health*, 4(4), 773-798. <http://doi.org/10.3390/plants4040773>
- Sugandh, S. (2017). Plant tissue culture: a promising tool of quality material production with special reference to micropropagation of banana, *Biochemical and Cellular Archives*, 17 (1):1-26. http://www.connectjournals.com/toc2.php?abstract=2685001H_1A.pdf&&bookmark=CJ-033216

- Tan, S.H., Maziah, M., & Ariff, A.B. (2013). Synergism effect between inoculum size and aggregatesize on flavonoid production in *Centella asiatica* (L.) urban (pegaga) cell suspension cultures. *International Journal of Research in Engineering and Technology*, 2(8), 244-253. <http://doi.org/issn-2319-1163>
- Taha, R. A., Mustafa, N. S., & Hassan, S. A. (2013). Protocol for Micropropagation of Two *Ficus carica* Cultivars. *World Journal of Agricultural Sciences*, 383-388. <http://doi.org/10.5829/idosi.wjas.2013.9.5.1802>
- Tay, C., Mohamad, S., Wan, N., & Saiyidah, N. (2006). *Pleurotus* spent mushroom compost as green supplementary nutrient in tissue culture. *MATEC Web of Conferences* 47, (pp. 45-46). <https://doi.org/10.1051/mateconf%2F20164705010>
- Tei, F., Nicola, S., & Benincasa, P. (2017). Advances in research on fertilization management of vegetable crops. *Advances in Olericulture*. <http://doi.org/10.1007/978-3-319-53626-2>
- Thirumurugan, D., Cholarajan, A., Raja, S.S.S., & Vijayakumar, R. (2018). An introductory chapter: secondary metabolites. *Intech Open*. <http://doi.org/10.5772/intechopen.79766>
- Thimann, K.V. (2008). Auxins and the inhibition of plant growth. *Biological Reviews*, 14 (3), 314-337. <http://doi.org/10.1111/j.1469-185X.1939.tb00937.x>
- Thorpe, T. A. (2007). History of plant tissue culture. *Molecular Biotechnology*, 169-180. <https://doi.org/10.1007/s12033-007-0031-3>.
- Thorpe, T.A., Stasolla, C., Yeung, E., & Klerk, M.A. (2008). The components of plant tissue culture media II organics addition, osmotic and pH effects and support systems. *Plant Propagation by Tissue Culture*, 3rd edition, 115-173. https://doi.org/10.1007/978-1-4020-5005-3_4
- Tripathi, D.K., Singh, V.P., Chauhan, D.K., Prasad, S.M., & Dubey, N.K. (2014). Role of macronutrients in plant growth and acclimatization: recent advances and future prospective, *Springer Sciences + Business Media New York*, 2 http://doi.org/10.1007/978-1-4614-8824-8_8
- Underwood, S.R., Anagnostopoulos, C., Cerqueira, M., Ell, P.J., Flint, E.J., Harbinson, M., Kelion, A.D., Al-Mohamad, A., Prvulovich, E.M., Shaw, L.J., & Tweddel, A.C. (2004). Myocardial perfusion scintigraphy: the evidence. *Eur J Nucl Med Mol Imaging*, 31(2), 261–291. <https://dx.doi.org/10.1007%2Fs00259-003-1344-5>
- Uzun, I. (2004). Use of spent mushroom compost in sustainable fruit production. *Journal of Fruit and Ornamental Plant Research*, 12, 157-165. http://www.inhort.pl/files/journal_pdf/journal_2004spec/full2004-18spec.pdf
- Vahid, A., Saadatmand, S., Riahi, H., & Khavari-Nejad, R. (2019). Influence of spent mushroom compost (smc) as an organic fertilizer on nutrient, growth, yield, and essential oil composition of german chamomile (*Matricaria recutita* L.). *Communication in Soil Science and plant analysis*, 50(1), 1-11. <http://doi.org/10.1080/00103624.2019.1568450>

- Vanneste, S., & Friml, J. (2013). Calcium: the missing link in auxin action. *Journal of National Institute of Health*, 2 (4), 650-675. <https://dx.doi.org/10.3390%2Fplants2040650>
- Vargas, V. M. L., & Alejo, N. O. (2018). *Plant Cell culture Protocols ,Methods in Molecular Biology*, Springer Science + Business Media,1815.http://doi.org/10.1007/978-1-4939-8594-4_1
- Wang, T. L., Fui, C. L., Wei, Y. M., Sreeramanan, S., & Bee, L. C. (2018). Shoot induction from axillary shoot tip explants of fig (*Ficus carica*) cv. Japanese BTM 6. *Tropical Life Science Research*, 165-174.<https://doi.org/10.21315/tlsr2018.29.2.11>
- Wang, Y., & Irving, H. (2011). Developing a model of plant hormone interactions. *Plant Signal Behaviour*, 494-500.<http://doi.org/10.4161/psb.6.4.14558>
- Weijers, D., Nemhauser, J., & Yang, Z. (2018). Auxin: small molecule, big impact. *Journal of Experimental Botany*, 61, 49-64. <http://doi.org/10.1093/jxb/erx463>
- Wenhao, D., Yuanjie, S., & Castillo, C. (2010). Plant regeneration from *in vitro* leaf tissue of *Viburnum dendatum* L. *Plant Cell Tissue Organ Culture*, 104(2),257-262. <http://doi.org/10.1007/s11240-010-9829-y>
- William, E. N., & Alan, D. K. (2014). Steroids:pharmacology, complications and practice delivery issues. *The Ochsner Journal*,14(2), 203-207. <https://www.ncbi.nlm.nih.gov/pubmed/24940130>
- Yaacob,J.S., Mahmad, N., Taha, R.M., Mohamed, N., Yussof, A.I.M, & Saleh, A. (2014). Optimization of culture conditions (sucrose, pH, and photoperiod) for *in vitro* regeneration and early detection of somaclonal variation in ginger lime (*Citrus assamensis*). *The Scientific World Journal*,1-9. <https://doi.org/10.1155/2014/262710>