



SUPERVISOR'S DECLARATION

We hereby declare that we have checked this thesis and in our opinion, this thesis is adequate in terms of scope and quality for the award of the degree of Master of Science.

A handwritten signature in black ink, appearing to be 'Jana', written over a horizontal line.

(Supervisor's Signature)

Full Name : DR NASRATUN BINTI MASNGUT

Position : SENIOR LECTURER

Date : 28 OCTOBER 2021

A handwritten signature in black ink, appearing to be 'Shalyda', written over a horizontal line.

(Co-supervisor's Signature)

Full Name : DR SHALYDA BINTI MD SHAARANI@MD NAWI

Position : SENIOR LECTURER

Date : 28 OCTOBER 2021



STUDENT'S DECLARATION

I hereby declare that the work in this thesis is based on my original work except for quotations and citations which have been duly acknowledged. I also declare that it has not been previously or concurrently submitted for any other degree at Universiti Malaysia Pahang or any other institutions.



(Student's Signature)

Full Name : YASHINI A/P K.SELVANATHAN

ID Number : MKC19005

Date : 28 OCTOBER 2021

MICROBIAL ACID PRODUCTION BY INDIGENOUS MICROORGANISM
FROM *Ananas comosus*

YASHINI A/P K.SELVANATHAN

Thesis submitted in fulfillment of the requirements
for the award of the degree of
Master of Science

College of Engineering
UNIVERSITI MALAYSIA PAHANG

NOVEMBER 2021

ACKNOWLEDGEMENTS

My gratitude specially dedicated to my supervisor, Dr. Nasratun Masngut and Dr. Shalyda Binti Md Shaarani@Md Nawi for their advice, insightful comments, and constant support within the making this research possible. Thanks for your guide and without it this research won't be ready to complete in a well-organized manner.

My special and sincere thanks to Pekan Pina Sdn Bhd for generously supplied of pineapple fruit (Royal Pepina) and the industrial grant of UIC190802. The Ministry of Education (MOE) for research grant FRGS/1/2018/TK02/UMP/02/10.

I am grateful for all of the encouragement and support from my beloved parents, Selvanathan Krishnan and Santhi Somalu, and my beloved siblings, Megalashini Selvanathan and Vikneswaran Selvanathan by giving advice and emotional support during my experiments and writing. And also, a huge thanks to Sri Mariamman amma for everything; without your presence, I would not be able to go through this.

I hereby acknowledge services and efforts of members and technical staffs of Faculty of Chemical and Process Engineering Technology (FTKKP), College of Engineering, and my colleague those involved during this research directly or indirectly, for his or her invaluable time, guidance, and advice. Without your cooperation and sacrifices, this research won't be able to complete and publish.

*Yashini Selvanathan
October 2021*

ABSTRAK

Akibat kekurangan sumber tenaga fosil, penghasilan asid mikrob daripada biojisim bagi menggantikan proses petrokimia semakin mendapat perhatian. Penggunaan sisa buangan dari industri buah, termasuklah industri nanas, boleh dilihat sebagai salah satu pendekatan yang boleh dilaksanakan. Kulit, isi, batang, dan daun dari tanaman nanas adalah biojisim bukan sumber makanan yang kompetitif, banyak dan murah yang boleh digunakan untuk penghasilan asid mikrob. Penghasilan asid mikrob pada masa kini menggunakan mikroorganisma tertentu, yang memerlukan keadaan spesifik untuk hidup. Kepelbagaian asid yang dihasilkan adalah terhad kepada metabolisme mikroorganisma tersebut. Sehingga kini, tidak ada kajian yang dilaporkan mengenai penggunaan mikroorganisma asli dari sisa nanas dan penggunaan sisa nanas tersebut sebagai sumber karbon untuk penghasilan asid mikrob. Kajian ini mengetengahkan pengoptimuman secara statistik untuk penghasilan asid mikrob dari mikroorganisma asli dalam kulit nanas. Ini turut merangkumi beberapa kajian mengenai fizikokimia, kualiti dan fungsi asid mikrob yang dihasilkan. Kajian ini melibatkan proses penyaringan faktor menggunakan reka bentuk faktorial (FFD) untuk menentukan faktor yang signifikan, diikuti dengan proses pengoptimuman statistik menggunakan reka bentuk komposit berpusat (CCD) daripada kaedah gerak balas permukaan (RSM). Prestasi proses penapaian dinilai berdasarkan keasidan (%). Keadaan optimum untuk fermentasi asid mikrob menggunakan mikroorganisma asli dicapai pada suhu 27 °C, dengan penambahan glukosa 7 % di dalam jus kulit nanas. Penapaian dilakukan selama lima hari secara aerobik tanpa menambahkan ragi. Keadaan fermentasi seperti penambahan glukosa dan mod penapaian sangat mempengaruhi pengeluaran asid yang mana ianya berkaitan dengan kesan *Crabtree*. Asid mikrob yang dihasilkan dicirikan berdasarkan fizikokimia, kualiti dan fungsi. Asid mikrob mengandungi 3.18 % gula pengurangan, 4.0 % sukrosa, 1.03 % etanol, dan 3.03 % asid, terutamanya asid asetik, malik dan sitrik dengan pH 3.16 dan 8.0 °Brix jumlah pepejal larut. Sementara itu, untuk kualiti dan kefungsiannya, asid mikrob yang dihasilkan mengandungi 1.43 mg equi. AA/100mL asid askorbik dan 82.06 % aktiviti radikal bebas antioksidan. Kesimpulannya, pengeluaran asid mikrob oleh mikroorganisma asli dari kulit nanas berjaya dioptimumkan menggunakan RSM. Walaupun dengan kandungan asid yang lebih rendah daripada asid mikrob komersial, kualitinya asid yang dihasilkan adalah setara.

ABSTRACT

Threatened with the gradual and inescapable exhaustion of the earth's fossil energy resources, fermentative acid production from renewable biomass to replace the conventional petrochemical process is receiving an increasing amount of attention. Microbial acids production from fruit waste to compensate for the wastage from fruit industries, including pineapple fruit, can be seen as one of the feasible approach. Peel, core, stem, and leaves from the pineapple plant are attractive, abundant, cheap, and non-food competitive biomass feedstock for microbial acid production. Current microbial acid production uses specific strains which require particular condition to grow. The diversity of the produced acid was limited to the strain metabolism. To date, there is no study reported on using indigenous microorganisms from the pineapple waste and using such waste as the carbon source for microbial acid fermentation. This study had work on a statistical optimisation for microbial acid production from indigenous microorganisms in the pineapple peel. The works involved were factors screening using Full Factorial Design (FFD) to determine the significant one, followed by a statistical optimisation process using Central Composite Design (CCD) of Response Surface Methodology (RSM). Study on the physicochemical, quality, and functionality of the produced microbial acid were also carried out. The performance of the fermentation process was evaluated based on the percentage of acidity (%). The optimised condition for the fermentation of pineapple peel using indigenous microorganisms was achieved at 27 °C, with 7 % glucose addition to the pineapple peel juice. The fermentation was carried out aerobically for five days without adding yeast. The condition of fermentation, such as sugar and mode of fermentation, was identified to affect the acid production in which showing the Crabtree effect. The produced microbial acid was characterised based on its physicochemical, quality, and functionality. The microbial acid contained 3.18 % reducing sugar, 4.0 % sucrose, 1.03 % ethanol, and 3.03 % acids, mainly acetic, malic, and citric acid with pH of 3.16 and 8.0 °Brix total soluble solid. Meanwhile, for the quality and functionality, the microbial acid produced contained 1.43 mg equi. AA/100mL ascorbic acid and 82.06 % free-radical scavenging activity of antioxidant. In conclusion, microbial acid production by indigenous microorganisms from pineapple peel was successfully optimised using RSM. Although with lower acid content than the commercial microbial acids, its quality was at par.

TABLE OF CONTENT

DECLARATION	
TITLE PAGE	
ACKNOWLEDGEMENTS	ii
ABSTRAK	iii
ABSTRACT	iv
TABLE OF CONTENT	v
LIST OF TABLES	viii
LIST OF FIGURES	ix
LIST OF SYMBOLS	x
LIST OF ABBREVIATIONS	xi
CHAPTER 1 INTRODUCTION	1
1.1 Background of Study	1
1.2 Problem Statement	2
1.3 Objectives	4
1.4 Scopes of the Study	4
CHAPTER 2 LITERATURE REVIEW	5
2.1 Pineapple Waste	5
2.1.1 The pineapple fruit	5
2.1.2 Pineapple waste disposal	5
2.1.3 Pineapple waste composition	6
2.1.4 Pineapple waste utilisation	9
2.2 Microbial Acid Production	13
2.2.1 Microbial acid production methods	13

2.3	Factors Affecting Microbial Acids Production	16
2.4	Optimisation of Microbial Acid Production	20
2.5	Characteristics of Microbial Acids	23
2.6	Conclusion	23
CHAPTER 3 METHODOLOGY		25
3.1	Experimental Workflow	25
3.2	Pineapple Peel Substrate Preparation	26
3.2.1	Source of pineapple peel	26
3.2.2	Preparation and characterisation of pineapple peel substrate	27
3.2.3	Microbial acid production	27
3.3	Screening of Microbial Acid Production	28
3.3.1	Preliminary studies	28
3.3.2	Screening the significant process factors on microbial acid production using full factorial design	28
3.3.3	Validation of the screening condition	29
3.4	Optimisation of Microbial Acid Production	30
3.4.1	Experimental design	30
3.4.2	Validation of optimisation condition	31
3.5	Characterisation of Acids and Analytical Procedures	31
3.5.1	Determination of pH	32
3.5.2	Determination of total soluble solids, sucrose and ethanol content	32
3.5.3	Estimation of total acidity using the titration method	32
3.5.4	Quantification of acid	33
3.5.5	Quantification of reducing sugar	33
3.5.6	DPPH radical scavenging activity and ascorbic acid content	33
3.5.7	Quantification of total plate count	34

CHAPTER 4 RESULTS AND DISCUSSION	35
4.1 Introduction	35
4.2 Characterisation of Pineapple Peel Substrate	35
4.3 Screening the Microbial Acid Production	36
4.3.1 Preliminary studies	36
4.3.2 Screening the significant process factors on microbial acid production using FFD	39
4.3.3 Main factors contribution and its interaction	43
4.3.4 Validation of the Full Factorial Design model	48
4.4 Optimisation of the Microbial Acid Production	48
4.4.1 Response surface plot	50
4.4.2 Validation of the CCD model	52
4.5 Characterisation of Acid	59
4.5.1 Physicochemical properties of microbial acid	59
4.5.2 Quality and functionality of acid	61
CHAPTER 5 CONCLUSIONS	64
5.1 Introduction	64
5.2 Conclusion	64
5.3 Recommendation	64
REFERENCES	66
APPENDICES	74
APPENDIX A: SOLUTION AND BUFFER PREPARATION	74
APPENDIX B: PREPARATION OF STANDARD CURVE	76

LIST OF TABLES

Table 2.1	Chemical composition of pineapple waste parts	8
Table 2.2	Phenolic compound from pineapple fruit and waste	10
Table 2.3	Microbial acid production from pineapple waste	12
Table 2.4	The process factors tested previously for microbial acid production	17
Table 2.5	Optimization studies of microbial acid fermentation using experimental design	22
Table 3.1	Factors and their coded and actual levels used in 2 ⁶ full factorial design experiments	29
Table 3.2	Factors condition for a validation run in the screening process	30
Table 3.3	The working range of factors in CCD	30
Table 3.4	Factors for validation of optimised condition	31
Table 4.1	Chemical composition of peel slurry and peel juice for Royal Pepina pineapple peel	36
Table 4.2	The result of screening experiments using full factorial design	40
Table 4.3	Test of significance for the regression coefficient	42
Table 4.4	Percentage contribution of main factor and their interaction	44
Table 4.5	Comparison between addition of yeast on pineapple peel without adding sugar	47
Table 4.6	Comparison between addition of sugar on pineapple peel without added yeast	47
Table 4.7	Validation result of full factorial screening	48
Table 4.8	The result of optimisation using a CCD	49
Table 4.9	ANOVA for optimisation of acid production	49
Table 4.10	Validation results of microbial acid optimisation	52
Table 4.11	Comparison between before and after optimisation	53
Table 4.12	Physicochemical properties of pineapple peel and commercial microbial acids	60

LIST OF FIGURES

Figure 2.1	Utilisation of pineapple in Malaysia.	6
Figure 2.2	Different parts of pineapple waste.	7
Figure 2.3	Microbial acid production route	14
Figure 2.4	Acetic acid fermentation pathway. EtOH., Ethanol; AcOH., Acetic acid; UQ., Ubiquinone; UGH ₂ ., Ubiquinol; PQQ., Pyrroloquinoline quinone; Acetyl CoA., Acetyl coenzyme A; NAD., Nicotinamide adenine dinucleotide; ADH., Alcohol dehydrogenase	15
Figure 3.1	Experimental flowchart	26
Figure 4.1	Effect of yeast concentration on acid production	37
Figure 4.2	Central metabolism of fermentation in yeasts	38
Figure 4.3	Interaction between factor C (temperature) and factor D (addition of glucose) with factor B (condition of fermentation) on acid production shown by the effect of individual factors; (a) Interaction between factor B and C and (b) Interaction between factor B and D	46
Figure 4.4	Interaction between temperature and addition of glucose on acid production shown by a 3D surface curve. One factor is varied while another one is kept constant at their centre points.	51
Figure 4.5	Model graphs for fermentation time (days)	54
Figure 4.6	Model graph for condition of fermentation (aerobic and anaerobic)	55
Figure 4.7	Sugar profile for microbial acid production	55
Figure 4.8	Acidity profile for microbial acid production	56
Figure 4.9	Pathways of microbial acid production without addition of yeast	58
Figure 4.10	Pathways of microbial acid production with added 0.3 % yeast	59
Figure 4.11	DPPH radical scavenging activity of pineapple peel and commercial microbial acids	62
Figure 4.12	Ascorbic acid concentration of pineapple peel and commercial microbial acid	63

REFERENCES

- Ahmed, J., Al-Jasass, F. M., & Siddiq, M. (2014). Date Fruit Composition and Nutrition. In L. John Wiley & Sons (Ed.), *Dates: Postharvest Science, Processing Technology and Health Benefits*. John Wiley & Sons, Ltd., 261-283.
- Amorima, J. C., Piccolib, R. H., & Duarte, W. F. (2018). Probiotic potential of yeasts isolated from pineapple and their use in the elaboration of potentially functional fermented beverages. *Food Research International* 107, 518–527.
- Arrizon, J., & Gschaedler, A. (2002). Increasing fermentation efficiency at high sugar concentrations by supplementing an additional source of nitrogen during the exponential phase of the tequila fermentation process. *Can. J. Microbiol.* 48, 965–970.
- Arroyo-López, F. N., Orlic, S., Querol, A., & Barrio, E. (2009). Effects of temperature, pH and sugar concentration on the growth parameters of *Saccharomyces cerevisiae*, *S. kudriavzevii* and their interspecific hybrid. *International Journal of Food Microbiology* 131, 120–127.
- Avissar, Y., Choi, J., & DeSaix, J. (2013). 2.2 Water. In *Biology* (1st edition ed., pp. 1482): OpenStax College.
- Ayub, M., Ullah, J., Muhammad, A., & Zeb, A. (2010). Evaluation of strawberry juice preserved with chemical preservatives at refrigeration temperature. *International Journal of Nutrition and Metabolism* 2, 027–032.
- Azizan, A., Lee, A. X., Hamid, N. A. A., Maulidiani, M., Mediani, A., Ghafar, S. Z. A., Zolkeflee, N. K. Z., & Abas, F. (2020). Potentially Bioactive Metabolites from Pineapple Waste Extracts and Their Antioxidant and α -Glucosidase Inhibitory Activities by ^1H NMR. *Foods* 9, 1–19.
- Barth, M., Hankinson, T. R., Zhuang, H., & Breidt, F. (2009). Microbiological Spoilage of Fruits and Vegetables. *Food Microbiology and Food Safety*, 135–183.
- Bezerraa, M. A., Santelli, R. E., Oliveiraa, E. P., Villar, L. S., & Escaleira, L. A. (2008). Response surface methodology (RSM) as a tool for optimization in analytical chemistry. *Talanta* 76, 965–977.
- Bhat, S. V., Akhtar, R., & Amin, T. (2014). An Overview on the Biological Production of Vinegar. *International Journal of Fermented Foods* 3, 139–155.
- Bulut, S., Elibol, M., & Ozer, D. (2004). Effect of different carbon sources on l(+) -lactic acid production by *Rhizopus oryzae*. *Biochemical Engineering Journal* 21, 33–37.
- Chai, L.-J., Qiu, T., Lu, Z.-M., Deng, Y.-J., Zhang, X.-J., Shi, J.-S., & Xu, Z.-H. (2020). Modulating microbiota metabolism via bioaugmentation with *Lactobacillus casei* and *Acetobacter pasteurianus* to enhance acetoin accumulation during cereal vinegar fermentation. *Food Research International* 138, 1–10.

- Chakraborty, K., Saha, S. K., Raychaudhuri, U., & Chakraborty, R. (2017). Vinegar from Bael (*Aegle marmelos*): A Mixed Culture Approach. *Indian Chemical Engineer* 60, 384–395.
- Chanprasartsuk, O.-o., Prakitchaiwattana, C., Sanguandeeikul, R., & Fleet, G. H. (2010). Autochthonous yeasts associated with mature pineapple fruits, freshly crushed juice and their ferments; and the chemical changes during natural fermentation. *Bioresource Technology* 101, 7500–7509.
- Chen, Y., Huang, Y., Bai, Y., Fu, C., Zhou, M., Gao, B., Wang, C., Li, D., Hu, Y., & Xu, N. (2017). Effects of mixed cultures of *Saccharomyces cerevisiae* and *Lactobacillus plantarum* in alcoholic fermentation on the physicochemical and sensory properties of citrus vinegar. *LWT—Food Science and Technology* 84, 753–763.
- Chukwuma, O. B., Rafatullah, M., Tajarudin, H. A., & Ismail, N. (2021). A Review on Bacterial Contribution to Lignocellulose Breakdown into Useful Bio-Products. *International Journal of Environmental Research and Public Health* 18, 1–27.
- Dacera, D. D. M., Babel, S., & Parkpian, P. (2009). Potential for land application of contaminated sewage sludge treated with fermented liquid from pineapple wastes. *Journal of Hazardous Materials* 167, 886–872.
- Dashko, S., Zhou, N., Compagno, C., & Piškur, J. 2014. Why, when, and how did yeast evolve alcoholic fermentation? *FEMS Yeast Res.* 14, 826–832.
- Fang, G.-Y., Chai, L.-J., Zhong, X.-Z., & Jiang, Y.-J. (2021). Deciphering the succession patterns of bacterial community and their correlations with environmental factors and flavor compounds during the fermentation of Zhejiang rosy vinegar. *International Journal of Food Microbiology* 341, 1–11.
- Fauzi, N. K. B. M. (2019). Screening of Process Parameter for Acetic Acid Production from Pineapple Waste Using Mixed Culture. (Bachelor of Chemical Engineering), University Malaysia Pahang,
- Ferreiraa, S., Duartea, A. P., Ribeiro, M. H. L., Queiroz, J. A., & Domingues, F. C. (2009). Response surface optimization of enzymatic hydrolysis of *Cistus ladanifer* and *Cytisus striatus* for bioethanol production. *Biochemical Engineering Journal* 45, 192–200.
- Fossi, B. T., & Tavea, F. d. r. (2013). Application of Amyolytic *Lactobacillus fermentum* 04BBA19 in Fermentation for Simultaneous Production of Thermostable Alpha-Amylase and Lactic Acid. In M. Kongo (Ed.), *R&D for Food, Health and Livestock Purposes*.
- García-Torres, R., Ponagandla, N. R., Rouseff, R. L., Goodrich-Schneider, R. M., & Reyes-De-Corcuera, J. I. (2009). Effects of Dissolved Oxygen in Fruit Juices and Methods of Removal. *Comprehensive Reviews in Food Science and Food Safety* 8, 409–423.
- Gardner, P. T., White, T. A. C., McPhail, D. B., & Duthie, G. G. (2000). The relative contributions of vitamin C, carotenoids and phenolics to the antioxidant potential

- of fruit juices. *Food Chemistry* 68, 471–474.
- Ghosh, S., Chakraborty, R., Chatterjee, A., & Raychaudhuri, U. (2014). Optimization of media components for the production of palm vinegar using response surface methodology. *Institute of Brewing & Distilling* 120, 550–558.
- Ghosh, S., Chakraborty, R., Chatterjee, G., & Raychaudhuri, U. (2012). Study on Fermentation Conditions of Palm Juice Vinegar by Response Surface Methodology and Development of a Kinetic Model. *Brazilian Journal of Chemical Engineering* 29, 461–472.
- Gomes, R. J., Borges, M. d. F., Rosa, M. d. F., Castro-Gómez, R. J. H., & Spinosa, W. A. (2018). Acetic Acid Bacteria in the Food Industry: Systematics, Characteristics and Applications. *Food Technology & Biotechnology* 56, 139–151.
- Grégrová, A., Čížková, H., Mazáč, J., & Voldřich, M. (2012). Authenticity and quality of spirit vinegar: Methods for detection of synthetic acetic acid addition. *Journal of Food and Nutrition Research* 51, 123–131.
- Gullo, M., Verzelloni, E., & Canonico, M. (2014). Aerobic submerged fermentation by acetic acid bacteria for vinegar production: Process and biotechnological aspects. *Process Biochemistry* 49, 1571–1579.
- Hamzaoui, A. H., Jamoussi, B., & M'nif, A. (2008). Lithium recovery from highly concentrated solutions: Responsesurface methodology (RSM) process parameters optimization. *Hydrometallurgy* 90, 1–7.
- Han, J. (2018). Malaysia masih kekurangan tanaman nanas premium: Nogeh. Utusan Borneo Online.
- Hewitson, J., & Hill, C. (2021). Science & Plants for Schools. How Does Sugar Affect Yeast Growth?
- Ho, C. W., Lazim, A. M., Fazry, S., Zaki, U. K. H. H., & Lim, S. J. (2017). Varieties, production, composition and health benefits of vinegars: A review. *Food Chemistry* 221, 1621–1630.
- Ho, C. W., Lazim, A., Fazry, S., Zaki, U. K. H. H., Massae, S., & Seng Joe Lima. (2019). Alcoholic fermentation of soursop (*Annona muricata*) juice via an alternative fermentation technique. *Journal of the Science of Food and Agriculture* 100, 1012–1021.
- Idris, A., & Wahidin, S. (2006). Effect of sodium alginate concentration, bead diameter, initial pH and temperature on lactic acid production from pineapple waste using immobilized *Lactobacillus delbrueckii*. *Process Biochemistry* 41, 1117–1123.
- Imandi, S. B., Bandaru, V. V., Somalanka, S. R., Bandaru, S. R., & Garapati, H. R. (2008). Application of statistical experimental designs for the optimization of medium constituents for the production of citric acid from pineapple waste. *Bioresource Technology* 99, 4445–4450.
- Islam, M. K., Khan, M. Z. H., Sarkar, M. A. R., Absar, N., & Sarkar, S. K. (2013).

- Changes in Acidity, TSS, and Sugar Content at Different Storage Periods of the Postharvest Mango (*Mangifera indica* L.) Influenced by Bavistin DF. *International Journal of Food Science* 13, 1–8.
- Ivanchenko, A., Yelatontsev, D., & Savenkov, A. (2021). Anaerobic co-digestion of agro-industrial waste with cheese whey: Impact of centrifuge comminution on biogas release and digestate agrochemical properties. *Biomass and Bioenergy* 147, 1–8.
- Jagannath, A., & K., S. (2020). Multi Target Preservation as an Effective Postharvest Processing Technology for the Chemical and Microbiological Stability of Pineapple (*Ananus Comosus*). *International Journal of Fruit Science*, 1–18.
- Johnston, C. S., & Gaas, C. A. (2006). Vinegar: Medicinal Uses and Antiglycemic Effect. *Medscape General Medicine* 8.
- Kang, J.-W., Kim, W.-J., & Kang, D.-H. (2020). Synergistic effect of 222-nm krypton-chlorine excilamp and mild heating combined treatment on inactivation of *Escherichia coli* O157:H7 and *Salmonella Typhimurium* in apple juice. *International Journal of Food Microbiology* 329, 1–13.
- Kim, S.-H., Cho, H.-K., & Shin, H.-S. (2012). Physicochemical Properties and Antioxidant Activities of Commercial Vinegar Drinks in Korea. *Food Sci. Biotechnol.* 21, 1729–1734.
- Kocher, G. S., Dhillon, H. K., & Joshi, N. (2012). Scale Up Of Sugarcane Vinegar Production By Recycling Of Successive Fermentation Batches And Its Organoleptic Evaluation. *Journal of Food Processing and Preservation* 38, 955–963. doi:10.1111/jfpp.12050
- Koffi, L. B., & Han, Y. W. (1990). Alcohol production from pineapple waste. *World journal of Microbiology and Biotechnology* 5, 281–284.
- Kong, C. T., Ho, C. W., Ling, J. W. A., Lazim, A., Fazry, S., & Lim, S. J. (2018). Chemical Changes and Optimisation of Acetous Fermentation Time and Mother of Vinegar Concentration in the Production of Vinegar-like Fermented Papaya Beverage. *Sains Malaysiana* 47, 2017–2026.
- Kumar, D., Jain, V. K., Shanker, G., & Srivastava, A. (2002). Utilisation of fruits waste for citric acid production by solid state fermentation. *Process Biochemistry* 38, 1725–1729.
- Leong, Y. K., Xui, O. C., & Ong, K. C. (2008). Survival of *SA11 Rotavirus* in Fresh Fruit Juices of Pineapple, Papaya, and Honeydew Melon. *Journal of Food Protection* 71, 1035–1037.
- Leonel, M., Suman, P. A., & Garcia, E. L. (2015). Production of Ginger Vinegar. *Ciência e Agrotecnologia* 39, 183–190.
- Leeuwen, J. H. v., Rasmussen, M. L., Sankaran, S., Koza, C. R., Erickson, D. T., Mitra, D., & Jin, B. (2005). Fungal Treatment of Crop Processing Wastewaters with Value-Added Co-Products. *Journal of Industrial Microbiology and*

Biotechnology 7, 13-44.

- Li, S., Li, P., Feng, F., & Luo, L.-X. (2015). Microbial diversity and their roles in the vinegar fermentation process. *Appl Microbiol Biotechnol* 99, 4997–5024.
- Liu, D., Zhu, Y., Beftink, R., Ooijkaas, L., Rinzema, A., Chen, J., & Tramper, J. (2004). Chinese Vinegar and its Solid-State Fermentation Process. *Food Reviews International* 20, 407–424.
- Liu, Q., Li, X., Sun, C., Wang, Q., Yao, H., Yang, W., Zheng, Z., Jiang, S., & Wu, X. (2019). Effects of mixed cultures of *Candida tropicalis* and aromatizing yeast in alcoholic fermentation on the quality of apple vinegar. *3 Biotech* 9, 1–10.
- Liu, Q., Tang, G.-Y., Zhao, C.-N., Gan, R.-Y., & Li, H.-B. (2019). Antioxidant Activities, Phenolic Profiles, and Organic Acid Contents of Fruit Vinegars. *Antioxidants*, 1–12.
- Lun, O. K., Wai, T. B., & Ling, L. S. (2014). Pineapple cannery waste as a potential substrate for microbial biotransformation to produce vanillic acid and vanillin. *International Food Research Journal* 21, 953–958
- Lunelli, B. H., Andrade, R. R., Atala, D. I. P., Maciel, M. R. W., Filho, F. M., & Filho, R. M. (2010). Production of lactic acid from sucrose: strain selection, fermentation, and kinetic modeling. *Appl Biochem Biotechnol*. 161, 227–237.
- Luzón-Quintana, L. M., Castro, R., & Durán-Guerrero, E. (2021). Biotechnological Processes in Fruit Vinegar Production. *Foods* 10, 1–23.
- Mamlouk, D., & Gullo, M. (2013). Acetic Acid Bacteria: Physiology and Carbon Sources Oxidation. *Indian J Microbial* 53, 377–384.
- Maicas, S. (2020). The Role of Yeasts in Fermentation Processes. *Microorganism* 8.
- Martins, S., Mussatto, S. I., Martínez-Avila, G., Montañez-Saenz, J., Aguilar, C. N., & Teixeira, J. A. (2011). Bioactive phenolic compounds: Production and extraction by solid-state fermentation. A review. *Biotechnology Advances* 29, 365–373.
- Marques, W. L., Raghavendran, V., Stambuk, B. U., & Gombert, A. K. (2015). Sucrose and *Saccharomyces cerevisiae*: a relationship most sweet. *FEMS Yeast Research* 16, 1–16.
- Masoumi, H. R. F., Kassim, A., Basri, M., & Abdullah, D. K. (2011). Determining Optimum Conditions for Lipase-Catalyzed Synthesis of Triethanolamine (TEA)-Based Esterquat Cationic Surfactant by a Taguchi Robust Design Method. *Molecules* 16, 4672–4680.
- Mukherjee, K., Paul, P., & Banerjee, E. R. (2014). Free Radical Scavenging Activities of Date Palm (*Phoenix sylvestris*) Fruit Extracts. *Natural Products Chemistry & Research* 2, 1–6.
- Mutalib, S. R. A., Samicho, Z., Abdullah, N., Zaman, N. K., & Hajar, N. (2012). *Effect of Maturity of Pineapple Variety N36 on Its Waste Physico-Chemical Properties*.

Paper presented at the IEEE Colloquium on Humanities, Science & Engineering Research.

- Orozco, F. G., Valadez-González, A., Domínguez-Maldonado, J. A., Zuluaga, F., Figueroa-Oyosa, L. E., & Alzate-Gaviria, L. M. (2014). Lactic Acid Yield Using Different Bacterial Strains, Its Purification, and Polymerization through Ring-Opening Reactions. *International Journal of Polymer Science*.
- OTI, & O, W. J. (2016). Using Refractometer to Determine the Sugar Content in Soft Drinks Commonly Consumed In Abakaliki, Nigeria. *IOSR Journal of Applied Chemistry* 9, 89–91.
- Owuama, C. I., & Ododo, J. C. (1993). Refractometric determination of ethanol concentration. *Food Chemistry* 48, 415–417.
- Pandey, A., Soccol, C. R., & Mitchell, D. (2000). New developments in solid state fermentation: I-bioprocesses and products. *Process Biochemistry* 35, 1153–1169.
- Patel, R., & Pandya, H. N. (2015). Production Of Acetic Acid From Molasses By Fermentation Process. *IJARIE* 1, 58–60.
- Pazuch, C. M., Kalschne, D. L., Siepmann, F. B., Marx, I. M. G., Oliveira, T. C. G. D., Spinosa, W. A., Canan, C., & Colla, E. (2020). Optimization and characterization of vinegar produced from rice bran. *Food Science and Technology* 40, 608–613.
- Pepin, C., & Marzacco, C. (2015). The fermentation of sugars using yeast: A discovery experiment. *CHEM 3 News Magazine*.
- Pfeiffer, T., & Morley, A. (2014). An evolutionary perspective on the Crabtree effect. *Frontiers in Molecular Biosciences* 17, 1–6.
- Prakitchaiwattana, C., Boonin, K., & Kaewklin, P. (2017). De-acidification of fresh whole pineapple juice wine by secondary malolactic fermentation with lactic acid bacteria. *International Food Research Journal* 24, 223–231.
- Raji, Y. O., Jibril, M., Misau, I. M., & Danjuma, B. Y. (2012). Production of Vinegar from Pineapple Peel International. *Journal of Advanced Scientific Research and Technology* 3, 656–666.
- Rasheed, A., Cobham, E., Zeighami, M., & Ong, S. (2012). *Extraction of Phenolic Compounds from Pineapple fruit* Paper presented at the The 2nd International Symposium on Processing & Drying of Foods, Vegetables and Fruits (ISPDFVF 2012) University of Nottingham, Malaysia Campus.
- Ray, R. C., & Sivakumar, P. S. (2009). Traditional and novel fermented foods and beverages from tropical root and tuber crops: review. *International Journal of Food Science and Technology* 44, 1073–1087.
- Roda, A., Lucini, L., Torchio, F., Dordoni, R., Faveri, D. M. D., & Lambri, M. (2017). Metabolite profiling and volatiles of pineapple wine and vinegar obtained from pineapple waste. *Food Chemistry* 229, 734–742.

- Roha, A. M. S., Zainal, S., Noriham, A., & Nadzirah, K. Z. (2013). Determination of sugar content in pineapple waste variety N36. *International Food Research Journal* 20, 1941–1943.
- Rongtong, B., Suwonsichon, T., Ritthiruangdej, P., & Kasemsumran, S. (2018). Determination of water activity, total soluble solids and moisture, sucrose, glucose and fructose contents in osmotically dehydrated papaya using near-infrared spectroscopy. *Agriculture and Natural Resources* 52, 557–564.
- Sakanaka, S., & Ishihara, Y. (2008). Comparison of antioxidant properties of persimmon vinegar and some other commercial vinegars in radical-scavenging assays and on lipid oxidation in tuna homogenates. *Food Chemistry* 107, 739–744.
- Salim, S. (2016). Transforming The Malaysian Pineapple Industry. *5th International Plantation Industry Conference & Exhibition (IPiCEx2016)*, 24.
- Sharma, R., Garg, P., Kumar, P., Bhatia, S. K., & Kulshrestha, S. (2020). Microbial Fermentation and Its Role in Quality Improvement of Fermented Foods. *Fermentation* 6, 1–20.
- Sjöblom, M., Matsakas, L., Christakopoulos, P., & Rova, U. (2015). Production of butyric acid by *Clostridium tyrobutyricum* (ATCC25755) using sweet sorghum stalks and beet molasses. *Industrial Crops and Products* 74, 535–544.
- Song, Y., Annous, B. A., & Fan, X. (2020). Cold plasma-activated hydrogen peroxide aerosol on populations of Salmonella Typhimurium and Listeria innocua and quality changes of apple, tomato and cantaloupe during storage - A pilot scale study. *Food Control* 117, 1–8.
- Sossou, S. K., Ameyapoh, Y., Karou, S. D., & Souza, C. d. (2009). Study of Pineapple Peelings Processing into Vinegar by Biotechnology. *Pakistan Journal of Biological Sciences* 12, 859–865.
- Spinosa, W. A., Júnior, V. d. S., Galvan, D., Fiorio, J. L., & Gomez, R. J. H. C. (2015). Vinegar rice (*Oryza sativa* L.) produced by a submerged fermentation process from alcoholic fermented rice. *Food Science and Technology* 35, 196–201.
- Srivastava, A. K., & Gupta, S. (2011). Comprehensive Biotechnology (Second Edition). In M. Moo-Young (Ed.), *Comprehensive Biotechnology* 2, 515–526.
- Stevenson, A., Buchanan, C. J., Abia, R., & Eastwood, M. A. (1997). A Simple In Vitro Fermentation System for Polysaccharides—The Effects of Fermenter Fluid Surface Area/Fluid Volume Ratio and Amount of Substrate. *Journal of the Science of Food and Agriculture* 73, 101–105.
- Sun, J., Chu, Y. F., & Liu, R. H. (2002). Antioxidant and antiproliferative activities of common fruits. *Journal of Agriculture and Food Chemistry* 50, 7449–7454.
- Swain, M. R., Anandharaj, M., Ray, R. C., & Rani, R. P. (2014). Fermented Fruits and Vegetables of Asia: A Potential Source of Probiotics. *Biotechnology Research Internationa* 1–32.

- Taamalli, A., Contreras, M. a. d. M., Abu-Reidah, I. M., Trabelsi, N., & Youssef, N. B. (2019). Quality of Phenolic Compounds: Occurrence, Health Benefits, and Applications in Food Industry. *Journal of Food Quality* 19, 1–2.
- Teixeira, R. S. S., Silva, A. S. A. d., Ferreira-Leitão, V. S., & Bon, E. P. d. S. (2012). Amino acids interference on the quantification of reducing sugars by the 3,5-dinitrosalicylic acid assay mislead carbohydase activity measurements. *Carbohydrate Research* 363, 33–37.
- Tesfaye, W., Garcia-Parrilla, M. C., & Troncoso, A. M. (2000). Set Up and Optimization of a Laboratory Scale Fermentor for the Production of Wine Vinegar. Set up and Optimization of a Laboratory Scale Fenueittor 106, 215–219.
- Thaipong, K., Boonprakob, U., Crosby, K., Cisneros-Zevallos, L., & Byrne, D. H. (2006). Comparison of ABTS, DPPH, FRAP, and ORAC assays for estimating antioxidant activity from guava fruit extracts. *Journal of Food Composition and Analysis* 19, 669–675.
- Ueno, T., Ozawa, Y., Ishikawa, M., Nakanishi, K., & Kimura, T. (2003). Lactic acid production using two food processing wastes, canned pineapple syrup and grape invertase, as substrate and enzyme. *Biotechnology Letters* 25, 573–577.
- Upadhyaya, S., Chetia, J., Bora, D. K., & Saikia, L. R. (2015). Total Phenolic Content, Antioxidant and Antimicrobial Activity of *Talinum cuneifolium* (Vahl.) from Dibrugarh, North East India. *Vedic Research International Phytomedicine* 3, 1–4.
- Wang, Q., Yang, L., Feng, K., Li, H., Deng, Z., & Liua, J. (2021). Promote lactic acid production from food waste fermentation using biogas slurry recirculation. *Bioresource Technology* 337.
- Xu, W., Huang, Z., Zhang, X., Li, Q., Lu, Z., Shi, J., Xu, Z., & Ma, Y. (2011). Monitoring the microbial community during solid-state acetic acid fermentation of Zhenjiang aromatic vinegar. *Food Microbiology* 28, 1175–1181.
- Yun, T. Y., Feng, R. J., Zhou, D. B., Pan, Y. Y., Chen, Y. F., Wang, F., Yin, L. Y., Zhang, Y. D., & Xie, J. H. (2018). Optimization of fermentation conditions through response surface methodology for enhanced antibacterial metabolite production by *Streptomyces* sp. 1-14 from cassava rhizosphere. *PLoS ONE* 13.
- Zhang, L., Huang, J., Zhou, R., & Wua, C. (2017). Evaluating the feasibility of fermentation starter inoculated with *Bacillus amyloliquefaciens* for improving acetoin and tetramethylpyrazine in Baoning bran vinegar. *International Journal of Food Microbiology* 255, 42–50.
- Zhang, B., Xia, T., Duan, W., Zhang, Z., Li, Y., Fang, B., . . . Wang, M. (2019). Effects of Organic Acids, Amino Acids and Phenolic Compounds on Antioxidant Characteristic of Zhenjiang Aromatic Vinegar. *Molecules* 24, 1–12.