

**DESIGN AND ANALYSIS OF INTEGRATED
PAPER BASED BIOCHIP WITH
MICROHEATER FOR LAMP USING VIRAL
MIMICKING NANO PARTICLES**

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ABSTRAK

Virus ialah organisma berjangkit sub-mikroskopik yang menyebabkan penyakit kepada manusia, haiwan dan tumbuhan yang mengakibatkan morbiditi dan boleh menyebabkan kematian. Beberapa teknik, termasuk antibodi, antigen, dan RNA, boleh digunakan untuk mengenal pasti jangkitan virus. Walau bagaimanapun, modaliti pengesanan RNA dan antigen lebih disukai untuk mendiagnosis jangkitan aktif. Oleh itu, keperluan ujian titik penjagaan (POCT) dalam mengesan RNA dan antigen adalah luas, manakala teknik yang lebih sedikit tersedia disebabkan oleh keperluan struktur dan metodologi yang kompleks. Selain itu, penapisan sampel virus daripada darah adalah proses yang membosankan. Pengkomersilan cip berasaskan kertas dalam bidang penahanan pengesanan RNA disebabkan oleh penyelidikan tidak mencukupi dalam penyepaduan cip berasaskan kertas dengan teknik yang disesuaikan untuk pengesanan RNA virus. Memandangkan penguatan berlaku pada suhu tinggi, pemanas mikro tertentu digunakan, Tetapi pemanas mikro sama ada lebih mahal untuk digunakan semula atau murah untuk dilupuskan. Untuk aplikasi pakai buang berskala besar, teknik fabrikasi sputtering atau PVD memakan masa dan meningkatkan tenaga kerja. Penyepaduan cip kertas dengan pemanas mikro boleh guna semula untuk teknologi penguatan isoterma (LAMP) ubat gelung adalah penting untuk membangunkan diagnosis titik penjagaan (POC) mudah alih yang pantas dan mudah digunakan. Dengan mempertimbangkan semua kebimbangan ini, kami telah membangunkan cip kertas untuk penapisan berkesan zarah virus daripada sampel dan menyepadukannya dengan pemanas mikro boleh guna semula untuk menyediakan suhu yang diperlukan untuk memulakan ujian LAMP. Dalam penyelidikan ini, cip kertas aliran sisi dan menegak telah dibuat menggunakan pencetak laser dan pemotongan manual, dan kecekapan penapisan dianalisis menggunakan mikrosfera. Halangan hidrofobik dijana dengan mengapit cip kertas di antara polietilena tereftalat (PET) dan slaid kaca. Sifat berliang kertas penapis menapis zarah dalam sampel. Begitu juga, pemanas mikro berasaskan papan litar bercetak (PCB) telah direka menggunakan teknik goresan basah dan penganalisaan ciri terma. Keputusan cip kertas menunjukkan bahawa cip kertas aliran sisi tidak cekap disebabkan oleh pengkapsulan PET. Namun, cip kertas aliran menegak dengan salur masuk dan keluar gred 4 menapis 98.57% zarah yang tidak diperlukan daripada sampel dalam masa 5 saat. Selain itu, pembaziran cip boleh dikurangkan dengan menggunakan semula slaid kaca bawah dengan pensterilan yang betul. Analisis reka bentuk pemanas mikro mendedahkan bahawa konfigurasi meander mengatasi reka bentuk lain dengan perbezaan haba hanya $\sim 8^{\circ}\text{C}$ dan penggunaan kuasa 1.79 W. Selain itu, lebar jalur pemanas mikro 1.75mm meningkatkan produktiviti. Di samping itu, pemanas mikro yang direka dengan penyebar haba mempunyai perbezaan terma hanya $<5^{\circ}\text{C}$ jika dibandingkan dengan $\sim 10^{\circ}\text{C}$ dalam pemanas mikro tanpa penyebar haba. Pemanas mikro yang dibangunkan adalah stabil selama lebih 6 bulan pada suhu bilik dan 10 hari di dalam air apabila dikapsulkan dengan PET. Akibatnya, ia mempunyai jangka hayat yang panjang dan boleh mengendalikan sampel biologi basah. Selain itu, pemanas mikro yang disepadukan dengan cip kertas berkesan memindahkan haba dengan perbezaan suhu 0.5°C . Oleh itu, pemanas mikro bersepada cip kertas boleh membuka jalan untuk beberapa aplikasi seperti peranti makmal pada cip, ujian POC, ujian penguatan asid nukleik pantas, kultur sel dan penyelidikan biomolekul. Pada masa hadapan, biocip berasaskan kertas yang dibangunkan akan diuji untuk sampel biologi yang sebenar bagi mengesan jangkitan virus yang berbeza.

ABSTRACT

A virus is a sub-microscopic infectious organism that causes diseases to humans, animals, and plants resulting in morbidity and may cause mortality. Several techniques, including antibodies, antigens, and RNA, can be used to identify viral infections. However, RNA and antigen detection modalities are preferred to diagnose an active infection. Hence, the requirement of point-of-care tests (POCT) in detecting RNA and antigen is vast, whereas fewer techniques are available due to the requirement of complex structures and methodologies. Moreover, the filtration of viral samples from the blood is a tedious process. The commercialization of paper-based chips in the field of RNA detection holdups due to the inadequate research in the integration of paper-based chips with the techniques adapted for viral RNA detection. Since the amplification occurs at elevated temperatures, certain microheaters are employed, But the microheaters are either costlier for reuse or inexpensive for disposal. For large-scale disposable applications, sputtering or PVD fabrication techniques is time-consuming and increases the workforce. The integration of paper chips with the reusable microheater for loop-mediated isothermal amplification (LAMP) technology is crucial for developing rapid and easy-to-use hand-held point-of-care (POC) diagnosis. By considering all these concerns, we have developed a paper chip for effective filtration of viral particles from the sample and integrated it with a reusable microheater to provide the necessary temperature to initiate the LAMP assay. In this research, lateral and vertical flow paper chips were fabricated using a laser printer and manual cutting, and the filtration efficiency was analyzed using microspheres. The hydrophobic barriers are generated by sandwiching the paper chip between the polyethylene terephthalate (PET) and the glass slide. The porous nature of the filter paper filters the particles in the sample. Similarly, the printed circuit board (PCB) based microheater was fabricated using the wet etching technique and analyzed the thermal characteristics. The results of the paper chip convey that the lateral flow paper chip was inefficient due to PET encapsulation. Still, the vertical flow paper chip with grade 4 inlet and outlet filters 98.57% of unnecessary particles from the sample within 5 sec. Moreover, the wastage of chips can be reduced by reusing the bottom glass slide with proper sterilization. The design analysis of the microheater reveals that the meander configuration outperforms other designs with the thermal difference of only $\sim 8^{\circ}\text{C}$ and the power consumption of 1.79 W. Also, the microheater strip width of 1.75mm enhances productivity. In addition, the fabricated microheater with a heat spreader has a thermal difference of only $<5^{\circ}\text{C}$ when compared with $\sim 10^{\circ}\text{C}$ in a microheater without a heat spreader. The developed microheater was stable for over 6 months at room temperature and 10 days in water when encapsulated with PET. As a result, it has a long shelf life and can handle wet biological samples. Moreover, the microheater integrated with paper chip effectively transfers the heat with a temperature difference of 0.5°C . Thus paper chip integrated microheater can pave the way for several applications like lab-on-chip devices, POC assays, rapid nucleic acid amplification tests, cell cultures, and biomolecular research. In the future, the developed paper-based biochip will be tested for real-time biological samples to detect different viral infections.

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REFERENCES

- Abdeslam. A. A, Fouad, K., & A, K. (2020). Design and Optimization of Platinum Heaters for Gas Sensor Applications. *Digest Journal of Nanomaterials and Biostructures*, 5(1), 133 - 141.
- Advanced Diagnostics Laboratory. (2021).
- Albertoni, G., Castelo Girao, M. J., & Schor, N. (2014). Mini review: current molecular methods for the detection and quantification of hepatitis B virus, hepatitis C virus, and human immunodeficiency virus type 1. *International Journal of Infectious Disease*, 25, 145-149. <https://doi.org/10.1016/j.ijid.2014.04.007>
- Amasia, M., Cozzens, M., & Madou, M. J. (2012). Centrifugal microfluidic platform for rapid PCR amplification using integrated thermoelectric heating and ice-valving. *Sensors and Actuators B: Chemical*, 161(1), 1191-1197. <https://doi.org/10.1016/j.snb.2011.11.080>
- Asiello, P. J., & Baeumner, A. J. (2011). Miniaturized isothermal nucleic acid amplification, a review. *Lab Chip*, 11(8), 1420-1430. <https://doi.org/10.1039/c0lc00666a>
- Ast, V., Costina, V., Eichner, R., Bode, A., Aida, S., Gerhards, C., Thiaucourt, M., Dobler, G., Geilenkeuser, W. J., Wolfel, R., Neumaier, M., & Haselmann, V. (2021). Assessing the Quality of Serological Testing in the COVID-19 Pandemic: Results of a European External Quality Assessment (EQA) Scheme for Anti-SARS-CoV-2 Antibody Detection. *Journal Clin Microbiol*, 59(9), e0055921. <https://doi.org/10.1128/JCM.00559-21>
- Bartlett, J. M., & Stirling, D. (2003). A short history of the polymerase chain reaction. *Methods Mol Biol*, 226, 3-6. <https://doi.org/10.1385/1-59259-384-4:3>
- Basiri, A., Heidari, A., Nadi, M. F., Fallahy, M. T. P., Nezamabadi, S. S., Sedighi, M., Saghazadeh, A., & Rezaei, N. (2021). Microfluidic devices for detection of RNA viruses. *Rev Med Virol*, 31(1), 1-11. <https://doi.org/10.1002/rmv.2154>
- Blow, N. (2007). Microfluidics: in search of a killer application. *Nature Methods*, 4(8), 665-670. <https://doi.org/10.1038/nmeth0807-665>
- Bodulev, O. L., & Sakharov, I. Y. (2020). Isothermal Nucleic Acid Amplification Techniques and Their Use in Bioanalysis. *Biochemistry (Mosc)*, 85(2), 147-166. <https://doi.org/10.1134/S0006297920020030>
- Botau, A., Bonfert, D., Negrea, C., Svasta, P., & Ionescu, C. (2015). Electro-thermal analysis of flexible micro-heater. Paper presented at the 2015 38th International Spring Seminar on Electronics Technology (ISSE), Eger, Hungary.
- Byers, K. M., Lin, L. K., Moehling, T. J., Stanciu, L., & Linnes, J. C. (2019). Versatile

- printed microheaters to enable low-power thermal control in paper diagnostics. *Analyst*, 145(1), 184-196. <https://doi.org/10.1039/c9an01546a>
- Byrnes, S., Thiessen, G., & Fu, E. (2013). Progress in the development of paper-based diagnostics for low-resource point-of-care settings. *Bioanalysis*, 5(22), 2821-2836. <https://doi.org/10.4155/bio.13.243>
- Cao, J. T., Cui, F., Chen, W., Guo, Z. X., Chen, W. Y., Wu, X. S., Liu, W., & Zhang, W. P. (2015). A PCR Chip with Reusable Electrodes and its Temperature Analysis. *Applied Mechanics and Materials*, 742, 54-57. <https://doi.org/10.4028/www.scientific.net/AMM.742.54>
- Cao, Y., & Dong, J. (2019). High-performance low-voltage soft electrothermal actuator with directly printed micro-heater. *Sensors and Actuators A: Physical*, 297. <https://doi.org/10.1016/j.sna.2019.111546>
- Cate, D. M., Adkins, J. A., Mettakoonpitak, J., & Henry, C. S. (2015). Recent developments in paper-based microfluidic devices. *Anal Chem*, 87(1), 19-41. <https://doi.org/10.1021/ac503968p>
- Chander, Y., Koelbl, J., Puckett, J., Moser, M. J., Klingele, A. J., Liles, M. R., Carrias, A., Mead, D. A., & Schoenfeld, T. W. (2014). A novel thermostable polymerase for RNA and DNA loop-mediated isothermal amplification (LAMP). *Front Microbiol*, 5, 395. <https://doi.org/10.3389/fmicb.2014.00395>
- Chang, C. M., Chang, W. H., Wang, C. H., Wang, J. H., Mai, J. D., & Lee, G. B. (2013). Nucleic acid amplification using microfluidic systems. *Lab Chip*, 13(7), 1225-1242. <https://doi.org/10.1039/c3lc41097h>
- Chang, H.-C., Chao, Y.-T., Yen, J.-Y., Yu, Y.-L., Lee, C.-N., Ho, B.-C., Liu, K.-C., Fang, J., Lin, C.-W., & Lee, J.-H. (2015). A Turbidity Test Based Centrifugal Microfluidics Diagnostic System for Simultaneous Detection of HBV, HCV, and CMV. *Advances in Materials Science and Engineering*, 2015, 1-8. <https://doi.org/10.1155/2015/306708>
- Chen, C., Liu, P., Zhao, X., Du, W., Feng, X., & Liu, B.-F. (2017). A self-contained microfluidic in-gel loop-mediated isothermal amplification for multiplexed pathogen detection. *Sensors and Actuators B: Chemical*, 239, 1-8. <https://doi.org/10.1016/j.snb.2016.07.164>
- Chen, J., Xu, Y., Yan, H., Zhu, Y., Wang, L., Zhang, Y., Lu, Y., & Xing, W. (2018). Sensitive and rapid detection of pathogenic bacteria from urine samples using multiplex recombinase polymerase amplification. *Lab Chip*, 18(16), 2441-2452. <https://doi.org/10.1039/c8lc00399h>
- Cheng, C. M., Martinez, A. W., Gong, J., Mace, C. R., Phillips, S. T., Carrilho, E., Mirica, K. A., & Whitesides, G. M. (2010). Paper-based ELISA. *Angew Chem Int Ed Engl*, 49(28), 4771-4774. <https://doi.org/10.1002/anie.201001005>

- Chevaliez, S., & Pawlotsky, J. M. (2007). Hepatitis C virus: virology, diagnosis and management of antiviral therapy. *World Journal Gastroenterol*, 13(17), 2461-2466. <https://doi.org/10.3748/wjg.v13.i17.2461>
- Chevaliez, S., & Pawlotsky, J. M. (2009). Virological techniques for the diagnosis and monitoring of hepatitis B and C. *Ann Hepatol*, 8(1), 7-12.
- Ching-Hsing Luo, Tsung-Min Hsieh, Chia-Sheng Liao, Gwo-Bin Lee, & Fu-Chun Huang. (2005). A Micromachined Low-power-consumption Portable PCR System. *Journal of Medical and Biological Engineering*, 26(1), 43-49.
- Chon, C. H., & Li, D. (2014). Temperature Control in Microfluidic Systems. In *Encyclopedia of Microfluidics and Nanofluidics* (pp. 1-6).
- Coarsey, C., Coleman, B., Kabir, M. A., Sher, M., & Asghar, W. (2019). Development of a Flow-Free Magnetic Actuation Platform for an Automated Microfluidic ELISA. *RSC Adv*, 9(15), 8159-8168. <https://doi.org/10.1039/C8RA07607C>
- Courbat, J., Canonica, M., Teyssieux, D., Briand, D., & de Rooij, N. F. (2011). Design and fabrication of micro-hotplates made on a polyimide foil: electrothermal simulation and characterization to achieve power consumption in the low mW range. *Journal of Micromechanics and Microengineering*, 21(1). <https://doi.org/10.1088/0960-1317/21/1/015014>
- Cui, F., Chen, W., Wu, X., Guo, Z., Liu, W., Zhang, W., & Chen, W. (2016). Design and experiment of a PDMS-based PCR chip with reusable heater of optimized electrode. *Microsystem Technologies*, 23(8), 3069-3079. <https://doi.org/10.1007/s00542-016-3064-3>
- Dao, T. N. T., Lee, E. Y., Koo, B., Jin, C. E., Lee, T. Y., & Shin, Y. (2018). A microfluidic enrichment platform with a recombinase polymerase amplification sensor for pathogen diagnosis. *Anal Biochem*, 544, 87-92. <https://doi.org/10.1016/j.ab.2017.12.030>
- Das, K., & Kakoty, P. (2015). Design, Simulation, and Performance Evaluation of a High Temperature and Low Power Consumption Microheater Structure for MOS Gas Sensors. In *Advances in Communication and Computing* (pp. 221-229).
- DeShields, J. B., Moroz, N., Braley, L. E., Mora-Romero, G. A., & Tanaka, K. (2019). Recombinase Polymerase Amplification (RPA) for the Rapid Isothermal Detection of Spongosporea subterranea f. sp. subterranea and Potato Mop-Top Virus. *American Journal of Potato Research*, 96(6), 617-624. <https://doi.org/10.1007/s12230-019-09750-7>
- Dungchai, W., Chailapakul, O., & Henry, C. S. (2009). Electrochemical detection for paper-based microfluidics. *Anal Chem*, 81(14), 5821-5826. <https://doi.org/10.1021/ac9007573>
- Farci, P., Alter, H. J., Govindarajan, S., Wong, D. C., Engle, R., Lesniewski, R. R.,

- Mushahwar, I. K., Desai, S. M., Miller, R. H., & Ogata, N. (1992). Lack of protective immunity against reinfection with hepatitis C virus. *Science*, 258(5079), 135-140. <https://doi.org/10.1126/science.1279801>
- Firdaus, R., Saha, K., Biswas, A., & Sadhukhan, P. C. (2015). Current molecular methods for the detection of hepatitis C virus in high risk group population: A systematic review. *World Journal Virol*, 4(1), 25-32. <https://doi.org/10.5501/wjv.v4.i1.25>
- Fire, A., & Xu, S. Q. (1995). Rolling replication of short DNA circles. *Proc Natl Acad Sci U S A*, 92(10), 4641-4645. <https://doi.org/10.1073/pnas.92.10.4641>
- Fronczek, C. F., Park, T. S., Harshman, D. K., Nicolini, A. M., & Yoon, J.-Y. (2014). Paper microfluidic extraction and direct smartphone-based identification of pathogenic nucleic acids from field and clinical samples. *RSC Advances*, 4(22). <https://doi.org/10.1039/c3ra47688j>
- Gayake. M, Bodas. D, & Gangal. S. (2011). Simulations of Polymer based Microheater Operated at Low Voltage. Paper presented at *the 2011 COMSOL Conference*, Bangalore.
- GeneCraft Labs. (2021).
- Gill, P., & Ghaemi, A. (2008). Nucleic acid isothermal amplification technologies: a review. *Nucleosides Nucleotides Nucleic Acids*, 27(3), 224-243. <https://doi.org/10.1080/15257770701845204>
- Goo, N. I., & Kim, D. E. (2016). Rolling circle amplification as isothermal gene amplification in molecular diagnostics. *Biochip Journal*, 10(4), 262-271. <https://doi.org/10.1007/s13206-016-0402-6>
- Guan, T., & Puers, R. (2010). Thermal analysis of a Ag/Ti based microheater. *Procedia Engineering*, 5, 1356-1359. <https://doi.org/10.1016/j.proeng.2010.09.366>
- Ha, B. H., Lee, K. S., Destgeer, G., Park, J., Choung, J. S., Jung, J. H., Shin, J. H., & Sung, H. J. (2015). Acoustothermal heating of polydimethylsiloxane microfluidic system. *Sci Rep*, 5, 11851. <https://doi.org/10.1038/srep11851>
- Han, J.-W., & Meyyappan, M. (2016). A Built-In Temperature Sensor in an Integrated Microheater. *IEEE Sensors Journal*, 16(14), 5543-5547. <https://doi.org/10.1109/jsen.2016.2569445>
- Hasan, M. N., Acharjee, D., Kumar, D., Kumar, A., & Maity, S. (2016). Simulation of Low Power Heater for Gas Sensing Application. *Procedia Computer Science*, 92, 213-221. <https://doi.org/10.1016/j.procs.2016.07.348>
- Hilton, J. P., Nguyen, T., Barbu, M., Pei, R., Stojanovic, M., & Lin, Q. (2012). Bead-based polymerase chain reaction on a microchip. *Microfluidics and Nanofluidics*, 13(5), 749-760. <https://doi.org/10.1007/s10404-012-0993-8>

- Holt, N., Marques, L. G., Van Horn, A., Montazeri, M., & Zhou, W. (2017). Fabrication and control of a microheater array for Microheater Array Powder Sintering. *The International Journal of Advanced Manufacturing Technology*, 95(1-4), 1369-1376. <https://doi.org/10.1007/s00170-017-1316-8>
- Holt, N., Van Horn, A., Montazeri, M., & Zhou, W. (2018). Microheater array powder sintering: A novel additive manufacturing process. *Journal of Manufacturing Processes*, 31, 536-551. <https://doi.org/10.1016/j.jmapro.2017.12.009>
- Horade, M., Kojima, M., Kamiyama, K., Mae, Y., & Arai, T. (2016). Development of a Novel 2-Dimensional Micro-Heater Array Device with Regional Selective Heating. *Mechanical Engineering Research*, 6(1). <https://doi.org/10.5539/mer.v6n1p66>
- Hwang, I.-S., Lee, E.-B., Kim, S.-J., Choi, J.-K., Cha, J.-H., Lee, H.-J., Ju, B.-K., & Lee, J.-H. (2011). Gas sensing properties of SnO₂ nanowires on micro-heater. *Sensors and Actuators B: Chemical*, 154(2), 295-300. <https://doi.org/10.1016/j.snb.2009.11.012>
- Hwang, W. J., Shin, K. S., Roh, J. H., Lee, D. S., & Choa, S. H. (2011). Development of micro-heaters with optimized temperature compensation design for gas sensors. *Sensors (Basel)*, 11(3), 2580-2591. <https://doi.org/10.3390/s110302580>
- James, J., Sushmitha, M., Premkumar, R., Narayananamurthy, V., & Kalpana, R. (2017). Microfluidic micro-well (size and shape) by numerical optimization for single cell applications: Vertical trapping approach. Paper presented at the 2017 International conference on Microelectronic Devices, Circuits and Systems (ICMDCS).
- Javed, A., Iqbal, S. M., & Jain, A. (2012). Microheater platform for selective detachment of DNA. *Applied Physics Letters*, 101(9). <https://doi.org/10.1063/1.4748308>
- Jeong, S., Lim, J., Kim, M. Y., Yeom, J., Cho, H., Lee, H., Shin, Y. B., & Lee, J. H. (2018). Portable low-power thermal cycler with dual thin-film Pt heaters for a polymeric PCR chip. *Biomed Microdevices*, 20(1), 14. <https://doi.org/10.1007/s10544-018-0257-9>
- Jha, S. K., Joo, G.-S., Ra, G.-S., Lee, H. H., & Kim, Y.-S. (2011). Development of PCR Microchip for Early Cancer Risk Prediction. *IEEE Sensors Journal*, 11(9), 2065-2070. <https://doi.org/10.1109/jsen.2011.2105262>
- Jiang, Y., Li, S., Qiu, Z., Le, T., Zou, S., & Cao, X. (2019). Rolling circle amplification and its application in microfluidic systems for Escherichia coli O157:H7 detections. *Journal of Food Safety*, 39(5). <https://doi.org/10.1111/jfs.12671>
- Jinsol, J., & Jungchul, L. (2014). Design, Fabrication, and Characterization of Liquid Metal Microheaters. *Journal of Microelectromechanical Systems*, 23(5), 1156-1163. <https://doi.org/10.1109/jmems.2014.2307358>

- Joy, S., & Antony, J. K. (2015). Design and Simulation of a Micro Hotplate Using COMSOL Multiphysics for MEMS Based Gas Sensor. Paper presented at the *2015 Fifth International Conference on Advances in Computing and Communications (ICACC)*.
- Kaarj, K., Akarapipad, P., & Yoon, J. Y. (2018). Simpler, Faster, and Sensitive Zika Virus Assay Using Smartphone Detection of Loop-mediated Isothermal Amplification on Paper Microfluidic Chips. *Sci Rep*, 8(1), 12438. <https://doi.org/10.1038/s41598-018-30797-9>
- Karazi, S. M., Ahad, I. U., & Benyounis, K. Y. (2017). Laser Micromachining for Transparent Materials. In Reference Module in Materials Science and Materials Engineering.
- Kessararat Ugsornrat, Nitin V. Afzulpurkar, Anurat Wisitsoraat, & Adisorn Tuantranont. (2010). Design, Simulation, and Experimental Study of a Droplet-Based PCR by EWOD. *Sensors and Materials*, 22(6), 271–284.
- Kim, J. A., Lee, S. H., Park, H., Kim, J. H., & Park, T. H. (2010). Microheater based on magnetic nanoparticle embedded PDMS. *Nanotechnology*, 21(16), 165102. <https://doi.org/10.1088/0957-4484/21/16/165102>
- Koonin, E. V., Senkevich, T. G., & Dolja, V. V. (2006). The ancient Virus World and evolution of cells. *Biol Direct*, 1, 29. <https://doi.org/10.1186/1745-6150-1-29>
- Kubista, M., Andrade, J. M., Bengtsson, M., Forootan, A., Jonak, J., Lind, K., Sindelka, R., Sjöback, R., Sjogreen, B., Strombom, L., Stahlberg, A., & Zoric, N. (2006). The real-time polymerase chain reaction. *Mol Aspects Med*, 27(2-3), 95-125. <https://doi.org/10.1016/j.mam.2005.12.007>
- Kweon, O. J., Lim, Y. K., Kim, H. R., Kim, T. H., & Lee, M. K. (2019). Analytical performance of newly developed rapid point-of-care test for the simultaneous detection of hepatitis A, B, and C viruses in serum samples. *J Med Virol*, 91(6), 1056-1062. <https://doi.org/10.1002/jmv.25405>
- Lee, C. Y., Degani, I., Cheong, J., Lee, J. H., Choi, H. J., Cheon, J., & Lee, H. (2021). Fluorescence polarization system for rapid COVID-19 diagnosis. *Biosens Bioelectron*, 178, 113049. <https://doi.org/10.1016/j.bios.2021.113049>
- Lee, D.-S., Choi, O. R., & Seo, Y. (2019). A microheater on polyimide substrate for hand-held realtime microfluidic polymerase chain reaction amplification. *Micro and Nano Systems Letters*, 7(1). <https://doi.org/10.1186/s40486-019-0098-1>
- Lee, D.-S., Park, S. H., Chung, K. H., & Pyo, H.-B. (2008). A Disposable Plastic-Silicon Micro PCR Chip Using Flexible Printed Circuit Board Protocols and Its Application to Genomic DNA Amplification. *IEEE Sensors Journal*, 8(5), 558-564. <https://doi.org/10.1109/jsen.2008.918923>
- Lee, S., Kang, J., Ren, S., Laurell, T., Kim, S., & Jeong, O. C. (2013). A cross-

contamination-free SELEX platform for a multi-target selection strategy. *BioChip Journal*, 7(1), 38-45. <https://doi.org/10.1007/s13206-013-7106-y>

Lee, S. R., Kardos, K. W., Schiff, E., Berne, C. A., Mounzer, K., Banks, A. T., Tatum, H. A., Friel, T. J., Demicco, M. P., Lee, W. M., Eder, S. E., Monto, A., Yearwood, G. D., Guillou, G. B., Kurtz, L. A., Fischl, M., Unangst, J. L., Kriebel, L., Feiss, G., & Roehler, M. (2011). Evaluation of a new, rapid test for detecting HCV infection, suitable for use with blood or oral fluid. *J Virol Methods*, 172(1-2), 27-31. <https://doi.org/10.1016/j.jviromet.2010.12.009>

Lee, W. C., Ng, H. Y., Hou, C. Y., Lee, C. T., & Fu, L. M. (2021). Recent advances in lab-on-paper diagnostic devices using blood samples. *Lab Chip*, 21(8), 1433-1453. <https://doi.org/10.1039/d0lc01304h>

Lekshmi, M. S., Pamula, R., Kartik, A., & Suja, K. J. (2018). Performance analysis of micro hotplate based metal oxide nanowire gas sensor. Paper presented at the 2018 7th International Symposium on Next Generation Electronics (ISNE), Taipei, Taiwan.

Li, J., & Macdonald, J. (2015). Advances in isothermal amplification: novel strategies inspired by biological processes. *Biosens Bioelectron*, 64, 196-211. <https://doi.org/10.1016/j.bios.2014.08.069>

Li T., Xu L., & Y., W. (2018). Micro-Heater-Based Gas Sensors. Singapore: Springer.

Li, Z. a., Yang, J., Zhu, L., & Tang, W. (2016). Fabrication of paper micro-devices with wax jetting. *RSC Advances*, 6(22), 17921-17928. <https://doi.org/10.1039/c5ra26255k>

Lin, L., Jr., Wang, S. S., Wu, M. H., & Oh-Yang, C. C. (2011). Development of an integrated microfluidic perfusion cell culture system for real-time microscopic observation of biological cells. *Sensors (Basel)*, 11(9), 8395-8411. <https://doi.org/10.3390/s110908395>

Lin, Y.-C., Yamanishi, Y., & Arai, F. (2007). On-chip Temperature Sensing and Control for Cell Immobilization. Paper presented at the 2007 2nd IEEE International Conference on Nano/Micro Engineered and Molecular Systems.

Liu, H., & Crooks, R. M. (2011). Three-dimensional paper microfluidic devices assembled using the principles of origami. *J Am Chem Soc*, 133(44), 17564-17566. <https://doi.org/10.1021/ja2071779>

Liu, L., Cao, W., Wu, J., Wen, W., Chang, D. C., & Sheng, P. (2008). Design and integration of an all-in-one biomicrofluidic chip. *Biomicrofluidics*, 2(3), 34103. <https://doi.org/10.1063/1.2966453>

Liu, M., Hui, C. Y., Zhang, Q., Gu, J., Kannan, B., Jahanshahi-Anbuhi, S., Filipe, C. D., Brennan, J. D., & Li, Y. (2016). Target-Induced and Equipment-Free DNA Amplification with a Simple Paper Device. *Angew Chem Int Ed Engl*, 55(8),

2709-2713. <https://doi.org/10.1002/anie.201509389>

Lopes, C. M. A., & Felisberti, M. I. (2004). Thermal conductivity of PET/(LDPE/AI) composites determined by MDSC. *Polymer Testing*, 23(6), 637-643. <https://doi.org/10.1016/j.polymertesting.2004.01.013>

Martinez, A. W., Phillips, S. T., & Whitesides, G. M. (2008). Three-dimensional microfluidic devices fabricated in layered paper and tape. *Proc Natl Acad Sci U S A*, 105(50), 19606-19611. <https://doi.org/10.1073/pnas.0810903105>

Martinez, A. W., Phillips, S. T., Whitesides, G. M., & Carrilho, E. (2010). Diagnostics for the developing world: microfluidic paper-based analytical devices. *Anal Chem*, 82(1), 3-10. <https://doi.org/10.1021/ac9013989>

Mavraki, E., Moschou, D., Kokkoris, G., Vourdas, N., Chatzandroulis, S., & Tserepi, A. (2011). A continuous flow μPCR device with integrated microheaters on a flexible polyimide substrate. *Procedia Engineering*, 25, 1245-1248. <https://doi.org/10.1016/j.proeng.2011.12.307>

Mesforush, S., Jahanshahi, A., & Zadeh, M. K. (2019). Finite element simulation of isothermal regions in serpentine shaped PCB electrodes of a micro-PCR device. Paper presented at the *2019 27th Iranian Conference on Electrical Engineering (ICEE)*, Yazd, Iran, Iran.

Miao, G., Zhang, L., Zhang, J., Ge, S., Xia, N., Qian, S., Yu, D., & Qiu, X. (2020). Free convective PCR: From principle study to commercial applications-A critical review. *Anal Chim Acta*, 1108, 177-197. <https://doi.org/10.1016/j.aca.2020.01.069>

Miralles, V., Huerre, A., Malloggi, F., & Jullien, M. C. (2013). A Review of Heating and Temperature Control in Microfluidic Systems: Techniques and Applications. *Diagnostics (Basel)*, 3(1), 33-67. <https://doi.org/10.3390/diagnostics3010033>

Mirasoli, M., Bonvicini, F., Lovecchio, N., Petrucci, G., Zangheri, M., Calabria, D., Costantini, F., Roda, A., Gallinella, G., Caputo, D., de Cesare, G., & Nascetti, A. (2018). On-chip LAMP-BART reaction for viral DNA real-time bioluminescence detection. *Sensors and Actuators B: Chemical*, 262, 1024-1033. <https://doi.org/10.1016/j.snb.2018.02.086>

Montenegro, L., De Michina, A., Misciagna, G., Guerra, V., & Di Leo, A. (2013). Virus C hepatitis and type 2 diabetes: a cohort study in southern Italy. *Am J Gastroenterol*, 108(7), 1108-1111. <https://doi.org/10.1038/ajg.2013.90>

Moschou, D., Vourdas, N., Kokkoris, G., Papadakis, G., Parthenios, J., Chatzandroulis, S., & Tserepi, A. (2014). All-plastic, low-power, disposable, continuous-flow PCR chip with integrated microheaters for rapid DNA amplification. *Sensors and Actuators B: Chemical*, 199, 470-478. <https://doi.org/10.1016/j.snb.2014.04.007>

Motamedi, M. K., Saghafinia, M., Karami, A., & Gill, P. (2011). A review of the current

isothermal amplification techniques: Applications, advantages and disadvantages. *Journal of Global Infectious Diseases*, 3(3). <https://doi.org/10.4103/0974-777x.83538>

Na, W., Nam, D., Lee, H., & Shin, S. (2018). Rapid molecular diagnosis of infectious viruses in microfluidics using DNA hydrogel formation. *Biosens Bioelectron*, 108, 9-13. <https://doi.org/10.1016/j.bios.2018.02.040>

Narayananamurthy, V., Jeroish, Z. E., Bhuvaneshwari, K. S., Bayat, P., Premkumar, R., Samsuri, F., & Yusoff, M. M. (2020). Advances in passively driven microfluidics and lab-on-chip devices: a comprehensive literature review and patent analysis. *RSC Advances*, 10(20), 11652-11680. <https://doi.org/10.1039/d0ra00263a>

Narayananamurthy, V., Jeroish, Z. E., Bhuvaneshwari, K. S., & Samsuri, F. (2021). Hepatitis C virus (HCV) diagnosis via microfluidics. *Anal Methods*, 13(6), 740-763. <https://doi.org/10.1039/d0ay02045a>

Narayananamurthy, V., Lee, T., Khan, A. a., Samsuri, F., Mohamed, K., Hamzah, H., & Baharom, M. (2018). Pipette Petri Dish Single-Cell Trapping (PP-SCT) in Microfluidic Platforms: A Passive Hydrodynamic Technique. *Fluids*, 3(3). <https://doi.org/10.3390/fluids3030051>

Nge, P. N., Rogers, C. I., & Woolley, A. T. (2013). Advances in microfluidic materials, functions, integration, and applications. *Chem Rev*, 113(4), 2550-2583. <https://doi.org/10.1021/cr300337x>

Nicolau, D. V., Zhu, Y., Bui, A., Jin, H., Nahavandi, S., Harvey, E. C., & Sutalo, I. D. (2005). Thermal modeling of a microheater in a microchannel chip. Paper presented at the *BioMEMS and Nanotechnology II*, Brisbane, Australia.

Nie, J., Zhao, Y., & Peng, N. (2014). Multichannel oscillatory-flow PCR micro-fluidic chip with controllable temperature gradient. *Microsystem Technologies*, 21(1), 41-48. <https://doi.org/10.1007/s00542-014-2077-z>

Nieto, D., McGlynn, P., de la Fuente, M., Lopez-Lopez, R., & O'Connor G, M. (2017). Laser microfabrication of a microheater chip for cell culture outside a cell incubator. *Colloids Surf B Biointerfaces*, 154, 263-269. <https://doi.org/10.1016/j.colsurfb.2017.03.043>

Niu, X. Z., Peng, S. L., Liu, L. Y., Wen, W. J., & Sheng, P. (2007). Characterizing and Patterning of PDMS-Based Conducting Composites. *Advanced Materials*, 19(18), 2682-2686. <https://doi.org/10.1002/adma.200602515>

Noh, J. Y., Yoon, S. W., Kim, Y., Lo, T. V., Ahn, M. J., Jung, M. C., Le, T. B., Na, W., Song, D., Le, V. P., Haam, S., Jeong, D. G., & Kim, H. K. (2019). Pipetting-based immunoassay for point-of-care testing: Application for detection of the influenza A virus. *Sci Rep*, 9(1), 16661. <https://doi.org/10.1038/s41598-019-53083-8>

Notomi, T., Okayama, H., Masubuchi, H., Yonekawa, T., Watanabe, K., Amino, N., &

- Hase, T. (2000). Loop-mediated isothermal amplification of DNA. *Nucleic Acids Res*, 28(12), E63. <https://doi.org/10.1093/nar/28.12.e63>
- Oh, S. J., Park, B. H., Choi, G., Seo, J. H., Jung, J. H., Choi, J. S., Kim do, H., & Seo, T. S. (2016). Fully automated and colorimetric foodborne pathogen detection on an integrated centrifugal microfluidic device. *Lab Chip*, 16(10), 1917-1926. <https://doi.org/10.1039/c6lc00326e>
- P.Wilkins, P. (2013). Immunodiagnosis of CNS parasitic infections. In *Handbook of Clinical Neurology* (Vol. 114, pp. 13-36): Elsevier.
- Payne, D., Zitterkopf, N. L., & Muthukumar, A. (2008). Molecular Tools for the Detection and Characterization of Bacterial Infections: A Review. *Laboratory Medicine*, 39(7), 430-436. <https://doi.org/10.1309/m6mbu1kgp0ff1c00>
- Petrucci, G., Caputo, D., Nascetti, A., Lovecchio, N., Parisi, E., Alameddine, S., de Cesare, G., & Zahra, A. (2015). Thermal characterization of thin film heater for lab-on-chip application. Paper presented at the *2015 XVIII AISEM Annual Conference*, Trento, Italy.
- Ponde, R. A. (2011). Hidden hazards of HCV transmission. *Med Microbiol Immunol*, 200(1), 7-11. <https://doi.org/10.1007/s00430-010-0159-9>
- Prabowo, M. H., Chatthen, S., Rijiravanich, P., Limkittikul, K., & Surareungchai, W. (2020). Dengue NS1 detection in pediatric serum using microfluidic paper-based analytical devices. *Anal Bioanal Chem*, 412(12), 2915-2925. <https://doi.org/10.1007/s00216-020-02527-6>
- Rajput G.Y., Gofane M.S., & Dhobale S. (2018). Design of Micro-heater on 3D-SnO₂ Gas Sensor (Vol. 810). Singapore: Springer.
- Reboud, J., Xu, G., Garrett, A., Adriko, M., Yang, Z., Tukahebwa, E. M., Rowell, C., & Cooper, J. M. (2019). Paper-based microfluidics for DNA diagnostics of malaria in low resource underserved rural communities. *Proc Natl Acad Sci U S A*, 116(11), 4834-4842. <https://doi.org/10.1073/pnas.1812296116>
- Ren, K., Chen, Y., & Wu, H. (2014). New materials for microfluidics in biology. *Curr Opin Biotechnol*, 25, 78-85. <https://doi.org/10.1016/j.copbio.2013.09.004>
- Ren, K., Zhou, J., & Wu, H. (2013). Materials for microfluidic chip fabrication. *Acc Chem Res*, 46(11), 2396-2406. <https://doi.org/10.1021/ar300314s>
- Reverter, F., Prodromakis, T., Liu, Y., Georgiou, P., Nikolic, K., & Constandinou, T. (2014). Design considerations for a CMOS Lab-on-Chip microheater array to facilitate the in vitro thermal stimulation of neurons. Paper presented at the *2014 IEEE International Symposium on Circuits and Systems (ISCAS)*.
- Rivas, L., Reutersward, P., Rasti, R., Herrmann, B., Martensson, A., Alfven, T., Gantelius, J., & Andersson-Svahn, H. (2018). A vertical flow paper-microarray

assay with isothermal DNA amplification for detection of *Neisseria meningitidis*. *Talanta*, 183, 192-200. <https://doi.org/10.1016/j.talanta.2018.02.070>

Robin, L., Mboumba Bouassa, R. S., Nodjikouambaye, Z. A., Charmant, L., Matta, M., Simon, S., Filali, M., Mboup, S., & Belec, L. (2018). Analytical performances of simultaneous detection of HIV-1, HIV-2 and hepatitis C- specific antibodies and hepatitis B surface antigen (HBsAg) by multiplex immunochromatographic rapid test with serum samples: A cross-sectional study. *J Virol Methods*, 253, 1-4. <https://doi.org/10.1016/j.jviromet.2017.12.001>

Roy, S., Majhi, T., Sinha, S., Sarkar, C. K., & Saha, H. (2010). Electro thermal analysis and fabrication of low cost microheater using a nickel alloy for low temperature MEMS based gas sensor application. Paper presented at the *2010 International Conference on Industrial Electronics, Control and Robotics*.

Roy, S., Sarkar, C. K., & Bhattacharyya, P. (2012). A highly sensitive methane sensor with nickel alloy microheater on micromachined Si substrate. *Solid-State Electronics*, 76, 84-90. <https://doi.org/10.1016/j.sse.2012.05.040>

Ruiqi, L., Chandrappan, J., Vaidyanathan, K., & Win, S. S. (2011). Silicon micro heater based tagging module and the biocompatible packaging for capsule endoscope. Paper presented at the *2011 IEEE 61st Electronic Components and Technology Conference (ECTC)*.

Ryu, J. H., Kwon, M., Moon, J. D., Hwang, M. W., Lee, J. M., Park, K. H., Yun, S. J., Bae, H. J., Choi, A., Lee, H., Jung, B., Jeong, J., Han, K., Kim, Y., & Oh, E. J. (2018). Development of a Rapid Automated Fluorescent Lateral Flow Immunoassay to Detect Hepatitis B Surface Antigen (HBsAg), Antibody to HBsAg, and Antibody to Hepatitis C. *Ann Lab Med*, 38(6), 578-584. <https://doi.org/10.3343/alm.2018.38.6.578>

S. E. Moon, H. K. Lee, N. J. Choi, J. Lee, W. S. Yang, J. Kim, J. J. Jong, & D. J. Yoo. (2012). Low-power-Consumption metal oxide NO₂ gas sensor based on micro-heater and screen printing technology. *J Nanosci Nanotechnol*, 12(7), 5543-5546. <https://doi.org/10.1166/jnn.2012.6364>

Sabalza, M., Yasmin, R., Barber, C. A., Castro, T., Malamud, D., Kim, B. J., Zhu, H., Montagna, R. A., & Abrams, W. R. (2018). Detection of Zika virus using reverse-transcription LAMP coupled with reverse dot blot analysis in saliva. *PLoS One*, 13(2), e0192398. <https://doi.org/10.1371/journal.pone.0192398>

Sackmann, E. K., Fulton, A. L., & Beebe, D. J. (2014). The present and future role of microfluidics in biomedical research. *Nature*, 507(7491), 181-189. <https://doi.org/10.1038/nature13118>

Sagnelli, E., Coppola, N., Marrocco, C., Coviello, G., Rossi, G., Battaglia, M., Sagnelli, C., Messina, V., Tonziello, A., Scolastico, C., & Filippini, P. (2003). Diagnosis of HCV related acute hepatitis by serial determination of IgM to HCV: a preliminary observation. *J Biol Regul Homeost Agents*, 17(2), 207-210.

- Sam, S. S., Steinmetz, H. B., Tsongalis, G. J., Tafe, L. J., & Lefferts, J. A. (2013). Validation of a solid-phase electrochemical array for genotyping hepatitis C virus. *Exp Mol Pathol*, 95(1), 18-22. <https://doi.org/10.1016/j.yexmp.2013.04.001>
- Sanjay, S. T., Li, M., Zhou, W., Li, X., & Li, X. (2020). A reusable PMMA/paper hybrid plug-and-play microfluidic device for an ultrasensitive immunoassay with a wide dynamic range. *Microsystems & Nanoengineering*, 6(1). <https://doi.org/10.1038/s41378-020-0143-5>
- Scull, J. C. (2014). Nucleic Acid Extraction Techniques. In *Pathobiology of Human Disease* (pp. 4059-4063).
- Siemens Healthineers. (2021). Retrieved from <https://www.siemens-healthineers.com/molecular-diagnostics>
- Shen, F., Sun, B., Kreutz, J. E., Davydova, E. K., Du, W., Reddy, P. L., Joseph, L. J., & Ismagilov, R. F. (2011). Multiplexed quantification of nucleic acids with large dynamic range using multivolume digital RT-PCR on a rotational SlipChip tested with HIV and hepatitis C viral load. *J Am Chem Soc*, 133(44), 17705-17712. <https://doi.org/10.1021/ja2060116>
- Shen, K., Chen, X., Guo, M., & Cheng, J. (2005). A microchip-based PCR device using flexible printed circuit technology. *Sensors and Actuators B: Chemical*, 105(2), 251-258. [https://doi.org/10.1016/s0925-4005\(04\)00432-0](https://doi.org/10.1016/s0925-4005(04)00432-0)
- Shi, X., Xu, W., Shen, W., Wang, G., Wang, R., Li, X., & Song, W. (2018). Improving the stability of silver nanowire/polyimide composite films for transparent film heaters. *Journal of Materials Science: Materials in Electronics*, 30(3), 2089-2095. <https://doi.org/10.1007/s10854-018-0480-4>
- Son, J. M., Lee, J. H., Kim, J., & Cho, Y. H. (2015). Temperature distribution measurement of Au micro-heater in microfluidic channel using IR microscope. *International Journal of Precision Engineering and Manufacturing*, 16(2), 367-372. <https://doi.org/10.1007/s12541-015-0048-7>
- Sooknanan, R., Malek, L. T., & van Gemen, B. (1995). Nucleic Acid Sequence-Based Amplification. In *Molecular Methods for Virus Detection* (pp. 261-285).
- Spruit, R. G., van Omme, J. T., Ghatkesar, M. K., & Garza, H. H. P. (2017). A Review on Development and Optimization of Microheaters for High-Temperature In Situ Studies. *Journal of Microelectromechanical Systems*, 26(6), 1165-1182. <https://doi.org/10.1109/jmems.2017.2757402>
- Sri Surya Srikanth. S, Rajesh Kumar. B, Suresh. V, & Jyothi. V. (2019). Design and Simulation of Platinum Micro Heater for VOC sensing Applications. *International Journal of Innovative Technology and Exploring Engineering*, 9(2S3). <https://doi.org/10.35940/ijitee>
- Streets, A. M., & Huang, Y. (2013). Chip in a lab: Microfluidics for next generation life

science research. *Biomicrofluidics*, 7(1), 11302.
<https://doi.org/10.1063/1.4789751>

Strickland, G. T., & El-Kamary, S. S. (2013). Viral Hepatitis. In Hunter's Tropical Medicine and Emerging Infectious Disease (pp. 290-305).

Suresh, V., Qunya, O., Kanta, B. L., Yuh, L. Y., & Chong, K. S. L. (2018). Non-invasive paper-based microfluidic device for ultra-low detection of urea through enzyme catalysis. *R Soc Open Sci*, 5(3), 171980. <https://doi.org/10.1098/rsos.171980>

Thursz, M., & Fontanet, A. (2014). HCV transmission in industrialized countries and resource-constrained areas. *Nat Rev Gastroenterol Hepatol*, 11(1), 28-35. <https://doi.org/10.1038/nrgastro.2013.179>

Timurdogan, E., Alaca, B. E., Kavakli, I. H., & Urey, H. (2011). MEMS biosensor for detection of Hepatitis A and C viruses in serum. *Biosens Bioelectron*, 28(1), 189-194. <https://doi.org/10.1016/j.bios.2011.07.014>

Tiwari, S. K., Bhat, S., & Mahato, K. K. (2018). Design and fabrication of screen printed microheater. *Microsystem Technologies*, 24(8), 3273-3281. <https://doi.org/10.1007/s00542-018-3821-6>

Tropea, C., Yarin, A. L., & Foss, J. F. (2007). *Springer Handbook of Experimental Fluid Mechanics*.

Tucker, J. D., Bien, C. H., & Peeling, R. W. (2013). Point-of-care testing for sexually transmitted infections: recent advances and implications for disease control. *Curr Opin Infect Dis*, 26(1), 73-79. <https://doi.org/10.1097/QCO.0b013e32835c21b0>

Tudos, A. J., Besselink, G. J., & Schasfoort, R. B. (2001). Trends in miniaturized total analysis systems for point-of-care testing in clinical chemistry. *Lab Chip*, 1(2), 83-95. <https://doi.org/10.1039/b106958f>

Utomo, M. S., Whulanza, Y., & Kiswanto, G. (2019). Maskless visible-light photolithography of copper microheater for dynamic microbioreactor. Paper presented at the THE 4TH BIOMEDICAL ENGINEERING'S RECENT PROGRESS IN BIOMATERIALS, DRUGS DEVELOPMENT, HEALTH, AND MEDICAL DEVICES: *Proceedings of the International Symposium of Biomedical Engineering (ISBE) 2019*.

van der Helm, J., Geskus, R., Sabin, C., Meyer, L., Del Amo, J., Chene, G., Dorrucci, M., Muga, R., Porter, K., & Prins, M. (2013). Effect of HCV infection on cause-specific mortality after HIV seroconversion, before and after 1997. *Gastroenterology*, 144(4), 751-760 e752. <https://doi.org/10.1053/j.gastro.2012.12.026>

VanHorn, A., & Zhou, W. (2016). Design and optimization of a high temperature microheater for inkjet deposition. *The International Journal of Advanced Manufacturing Technology*, 86(9-12), 3101-3111.

<https://doi.org/10.1007/s00170-016-8440-8>

- Vasileva Wand, N. I., Bonney, L. C., Watson, R. J., Graham, V., & Hewson, R. (2018). Point-of-care diagnostic assay for the detection of Zika virus using the recombinase polymerase amplification method. *J Gen Virol*, 99(8), 1012-1026. <https://doi.org/10.1099/jgv.0.001083>
- Velmathi, G., Ramshanker, N., & Mohan, S. (2010). Design, Electro-Thermal simulation and geometrical optimization of double spiral shaped microheater on a suspended membrane for gas sensing. Paper presented at the *IECON 2010 - 36th Annual Conference on IEEE Industrial Electronics Society*.
- Vigneswaran, N., Samsuri, F., Ranganathan, B., & Padmapriya. (2014). Recent Advances in Nano Patterning and Nano Imprint Lithography for Biological Applications. *Procedia Engineering*, 97, 1387-1398. <https://doi.org/10.1016/j.proeng.2014.12.420>
- Virus strains. (2007). Retrieved from <https://www.sciencelearn.org.nz/resources/184-virus-strains>
- Visseaux, B., Larrouy, L., Calin, R., Katlama, C., Poynard, T., Raziu, V., & Thibault, V. (2013). Anti-hepatitis C virus antibody detection in oral fluid: influence of human immunodeficiency virus co-infection. *J Clin Virol*, 58(2), 385-390. <https://doi.org/10.1016/j.jcv.2013.07.015>
- Waheed, S., Cabot, J. M., Macdonald, N. P., Lewis, T., Guijt, R. M., Paull, B., & Breadmore, M. C. (2016). 3D printed microfluidic devices: enablers and barriers. *Lab Chip*, 16(11), 1993-2013. <https://doi.org/10.1039/c6lc00284f>
- Wang, C.-P., Hsiao, M.-H., Lee, G.-H., Chang, T.-L., & Lee, Y.-W. (2020). The investigation of electrothermal response and reliability of flexible graphene micro-heaters. *Microelectronic Engineering*, 228. <https://doi.org/10.1016/j.mee.2020.111334>
- Wang, P., Ge, L., Yan, M., Song, X., Ge, S., & Yu, J. (2012). Paper-based three-dimensional electrochemical immunodevice based on multi-walled carbon nanotubes functionalized paper for sensitive point-of-care testing. *Biosens Bioelectron*, 32(1), 238-243. <https://doi.org/10.1016/j.bios.2011.12.021>
- Warkad, S. D., Song, K. S., Pal, D., & Nimse, S. B. (2019). Developments in the HCV Screening Technologies Based on the Detection of Antigens and Antibodies. *Sensors (Basel)*, 19(19). <https://doi.org/10.3390/s19194257>
- Warren M. Rohsenow, James R Hartnett , & Young I. Cho. (1998). *Handbook of heat transfer (Third Edition ed.)*. New York: MCGRAW-HILL2.
- Weibel, D. B., & Whitesides, G. M. (2006). Applications of microfluidics in chemical biology. *Curr Opin Chem Biol*, 10(6), 584-591. <https://doi.org/10.1016/j.cbpa.2006.10.016>

- Weil, A. A., & Harris, J. B. (2015). *Vibrio cholerae*. In *Molecular Medical Microbiology* (pp. 1079-1098).
- Wong, Y. P., Othman, S., Lau, Y. L., Radu, S., & Chee, H. Y. (2018). Loop-mediated isothermal amplification (LAMP): a versatile technique for detection of micro-organisms. *J Appl Microbiol*, 124(3), 626-643. <https://doi.org/10.1111/jam.13647>
- Wu, J., Cao, W., Wen, W., Chang, D. C., & Sheng, P. (2009). Polydimethylsiloxane microfluidic chip with integrated microheater and thermal sensor. *Biomicrofluidics*, 3(1), 12005. <https://doi.org/10.1063/1.3058587>
- Yeh, C.-H., Chen, K.-R., & Lin, Y.-C. (2013). Developing heatable microfluidic chip to generate gelatin emulsions and microcapsules. *Microfluidics and Nanofluidics*, 15(6), 775-784. <https://doi.org/10.1007/s10404-013-1193-x>
- Yin, M., Xiao, L., Liu, Q., Kwon, S. Y., Zhang, Y., Sharma, P. R., Jin, L., Li, X., & Xu, B. (2019). 3D Printed Microheater Sensor-Integrated, Drug-Encapsulated Microneedle Patch System for Pain Management. *Adv Healthc Mater*, 8(23), e1901170. <https://doi.org/10.1002/adhm.201901170>
- Yu, S., Wang, S., Lu, M., & Zuo, L. (2015). A Novel Micro Heater Integrated on Flexible Polyimide Substrate With Fast Response and Uniform Temperature Distribution. Paper presented at the Volume 4: 20th Design for Manufacturing and the Life Cycle Conference; *9th International Conference on Micro- and Nanosystems*, Boston, Massachusetts, USA.
- Yu, S., Wang, S., Lu, M., & Zuo, L. (2017). A novel polyimide based micro heater with high temperature uniformity. *Sensors and Actuators A: Physical*, 257, 58-64. <https://doi.org/10.1016/j.sna.2017.02.006>
- Yuki, N., Hayashi, N., Ohkawa, K., Hagiwara, H., Oshita, M., Katayama, K., Sasaki, Y., Kasahara, A., Fusamoto, H., & Kamada, T. (1995). The significance of immunoglobulin M antibody response to hepatitis C virus core protein in patients with chronic hepatitis C. *Hepatology*, 22(2), 402-406.
- Z. E. Jeroish, K. S. Bhuvaneshwari, Vigneswaran Narayananmurthy, R. Premkumar, & Fahmi Samsuri. (2020). Tilt Based Passive Optimizations for Microfluidics And Lab-on-Chip Devices - A Simulation Study. *Journal of Engineering Science and Technology*, 15(3), 1840-1854.
- Zaghoul, H., & El-Shahat, M. (2014). Recombinase polymerase amplification as a promising tool in hepatitis C virus diagnosis. *World J Hepatol*, 6(12), 916-922. <https://doi.org/10.4254/wjh.v6.i12.916>
- Zhang, L., Zhang, P., Wang, R., Zhang, R., Li, Z., Liu, W., Wang, Q., Gao, M., & Gui, L. (2020). A Performance-Enhanced Liquid Metal-Based Microheater with Parallel Ventilating Side-Channels. *Micromachines (Basel)*, 11(2). <https://doi.org/10.3390/mi11020133>

- Zhao, W., Ali, M. M., Brook, M. A., & Li, Y. (2008). Rolling circle amplification: applications in nanotechnology and biodetection with functional nucleic acids. *Angew Chem Int Ed Engl*, 47(34), 6330-6337. <https://doi.org/10.1002/anie.200705982>
- Zhao, Y., Chen, F., Li, Q., Wang, L., & Fan, C. (2015). Isothermal Amplification of Nucleic Acids. *Chem Rev*, 115(22), 12491-12545. <https://doi.org/10.1021/acs.chemrev.5b00428>
- Zheng, C., Balasubramanian, G. P. S., Tan, Y., Maniatty, A. M., Hull, R., & Wen, J. T. (2017). Simulation, Microfabrication, and Control of a Microheater Array. *IEEE/ASME Transactions on Mechatronics*, 22(4), 1914-1919. <https://doi.org/10.1109/tmech.2017.2650682>
- Zhong, R., Pan, X., Jiang, L., Dai, Z., Qin, J., & Lin, B. (2009). Simply and reliably integrating micro heaters/sensors in a monolithic PCR-CE microfluidic genetic analysis system. *Electrophoresis*, 30(8), 1297-1305. <https://doi.org/10.1002/elps.200800491>
- Zhu, H., Fohlerova, Z., Pekarek, J., Basova, E., & Neuzil, P. (2020). Recent advances in lab-on-a-chip technologies for viral diagnosis. *Biosens Bioelectron*, 153, 112041. <https://doi.org/10.1016/j.bios.2020.112041>
- Zhu, L., Jiang, G., Wang, S., Wang, C., Li, Q., Yu, H., Zhou, Y., Zhao, B., Huang, H., Xing, W., Mitchelson, K., Cheng, J., Zhao, Y., & Guo, Y. (2010). Biochip system for rapid and accurate identification of mycobacterial species from isolates and sputum. *J Clin Microbiol*, 48(10), 3654-3660. <https://doi.org/10.1128/JCM.00158-10>