

CULTIVATION OF MICROALGAE
NANNOCHLOROPSIS SP. IN PALM OIL MILL
EFFLUENT FOR BIODIESEL PRODUCTION

KARTHIANI KANAGESAN

MASTER OF SCIENCE

UNIVERSITI MALAYSIA PAHANG

SUPERVISOR'S DECLARATION

We hereby declare that we have checked this thesis and in our opinion, this thesis is adequate in terms of scope and quality for the award of the degree of Master of Science.



(Supervisor's Signature)

Full Name : DR. NATANAMURUGARAJ GOVINDAN
Position : SENIOR LECTURER
Date : 23/05/2022



(Co-supervisor's Signature)

Full Name : PROF. DR. GAANTY PRAGAS MANIAM
Position : PROFESSOR
Date : 23/05/2022



STUDENT'S DECLARATION

I hereby declare that the work in this thesis is based on my original work except for quotations and citations which have been duly acknowledged. I also declare that it has not been previously or concurrently submitted for any other degree at Universiti Malaysia Pahang or any other institutions.

A handwritten signature in black ink, appearing to read 'Kartiani', is written above a horizontal line.

(Student's Signature)

Full Name : KARTHIANI KANAGESAN

ID Number : MKB20002

Date : 23/05/2022

CULTIVATION OF MICROALGAE *NANNOCHLOROPSIS* SP. IN PALM OIL
MILL EFFLUENT FOR BIODIESEL PRODUCTION

KARTHIANI KANAGESAN

Thesis submitted in fulfillment of the requirements
for the award of the degree of
Master of Science

Faculty of Industrial Sciences and Technology
UNIVERSITI MALAYSIA PAHANG

MAY 2022

ACKNOWLEDGEMENTS

First and foremost, praises and thanks to God, the Almighty, for His showers of blessings throughout my research work to complete the research successfully with minimal obstacles. I would like to express my deepest and most sincere gratitude to my research supervisor, Dr. Natanamurugaraj Govindan for his patience, motivation, enthusiasm, and immense knowledge. His guidance helped me in all the time of research and writing of this thesis. I could not have imagined having a better advisor and mentor for my research study.

I also express my appreciation to Prof. Dr. Gaanty Pragas Maniam, my co-supervisor, for his encouragement, insightful comments, and valuable inputs during the whole study. It was a great privilege and honor to work and study under his guidance. I am extremely grateful for what he has offered me.

I am forever thankful to my colleagues, Mr. Karthick Murugan Palanisamy and Ms. Primilla Paramasivam at the Faculty of Industrial Sciences and Technology, Universiti Malaysia Pahang for their friendship and support, and for creating a cordial working environment. Besides, I am heartily grateful to all academic, administration, and technical staffs of the Faculty of Industrial Sciences and Technology, Universiti Malaysia Pahang for their contributions during the study period. I am highly indebted to Universiti Malaysia Pahang for the financial support for this research work via research grants (RDU 190337).

It is a pleasure to thank my beloved supporting friends Ms. Pushparani Chadayam and Ms. Kaabilaswary Ganapathi for numerous discussions on related topics that helped me improve my knowledge in the area of study. Besides, I would like to thank all my friends in Universiti Malaysia Pahang particularly Ms. Sarmilaah Dewi Subramaniam who gave me helpful advice, guidance, effective encouragement throughout this research.

Words cannot adequately express my heartfelt gratitude towards my beloved Father, Mr. Kanagesan Vasu, and Mother, Mrs. Mageswari Kanagesan for being a constant source of love, concern, support, and strength these years. I am also grateful to Mrs. Thibasiinee Ganesan, Ms. Puguneswary, Ms. Thivya, and Mr. Hari who give me all the support, useful bits of advice, the prayer, and always be patient with me, thank you so much.

Last but not least, the completion of this undertaking could not have been possible without the participation and assistance of so many people whose names may not all be enumerated. Their contributions are sincerely appreciated and gratefully acknowledged. Only God will reciprocate your goodness.

ABSTRAK

Penggunaan sumber fosil secara meluas mengakibatkan kekurangan bahan bakar fosil dengan cepat. Biodiesel merupakan sumber tenaga alternatif yang boleh diperbaharui dan dapat mengurangkan pergantungan pada bahan bakar fosil. Penjanaaan biodiesel generasi terdahulu tidak ideal sebagai pengganti bahan bakar fosil kerana persaingan tanah untuk penanaman sumber makanan manusia. Untuk mengatasi persaingan tanah ini, biodiesel dihasilkan daripada sumber mikroalga. Walau bagaimanapun, penanaman mikroalga adalah mahal kerana memerlukan sumber nutrien dan air dalam jumlah besar. Akibatnya, efluen kilang kelapa sawit (POME), yang mengandungi kandungan nutrien yang banyak digunakan sebagai medium permakanan alternatif untuk mengurangkan kos pertumbuhan mikroalga. Dalam kajian ini, mikroalga dari Teluk Cempedak, Kuantan yang mempunyai kandungan lipid yang tinggi diasing dan dikultur untuk penghasilan biodiesel. Enam jenis mikroalga disaring dan dikenal pasti dalam pemeriksaan awal dengan kajian morfologi. Namun, mikroalga hijau *Nannochloropsis* sp. telah dipilih kerana memiliki kadar pertumbuhan yang tinggi dan kandungan lipid yang banyak. Faktor yang mempengaruhi penghasilan lipid intraselular seperti kesan kepekatan POME, jangkamasa pendedahan kepada cahaya dan panjang gelombang cahaya LED dikaji untuk menentukan faktor optimum bagi pertumbuhan dan penghasilan lipid *Nannochloropsis* sp. Pengkulturan *Nannochloropsis* sp. secara skala besar diikuti dengan kaedah pengekstrakan Soxhlet telah menghasilkan kandungan lipid sebanyak 61.5%. Lipid yang diekstrak kemudian ditransesterifikasikan dengan metanol dan kalium hidroksida menghasilkan metil ester (biodiesel) dalam 1.5 jam. Analisis kromatografi lapisan nipis (TLC) dilakukan untuk memastikan penukaran lipid *Nannochloropsis* sp. ke biodiesel. Pembentukan metil ester asid lemak yang diperoleh adalah sebanyak 80.24%. Kromatogram gas kromatografi spektrometri jisim (GS-MS) menggambarkan metil ester dari *Nannochloropsis* sp. mengandungi kandungan asid oleik (C_{18:1}) dan asid palmitik (C_{16:0}) yang tinggi, masing-masing sebanyak 66.13% dan 12.38%. Lipid *Nannochloropsis* sp. berpotensi untuk digunakan dalam industri biodiesel kerana komposisi asid lemak dan lipid yang tinggi.

ABSTRACT

Extensive usage of fossil reserves resulted in rapid fossil fuel depletion. Biodiesel is one of the renewable energy alternatives designed to reduce reliance on fossil fuels. The generation of biodiesel from edible and non-edible crops is not identified as an ideal substitute to fossil fuels due to the competition for limited cultivable land proposed to cultivate crops for human consumption. To overcome the drawback, biodiesel is derived from the cultivation of microalgae. However, the cultivation of microalgae is costly as they require nutrients and water in a large amount. As a result, palm oil mill effluent (POME), which contains a large number of nutrients required for microalgae growth, is used as an alternative nutrition medium for microalgae cultivation while treating wastewater. In this study, attempts have been made to isolate and mass cultivate high lipid content microalgae from Teluk Cempedak, Kuantan coast, for biodiesel production. Among the collected samples, six microalgae were screened upon preliminary screening for morphological studies however green microalgae *Nannochloropsis* sp. was identified to be the most suitable microalgae with a high growth rate and abundant lipid content. Culture factors influencing the intracellular lipid body were investigated. The effect of different POME concentrations, photoperiod regimes and light-emitting diode (LED) light wavelengths were examined to determine the optimum factor for *Nannochloropsis* sp. growth and lipid enhancement. The mass cultivation under combined optimized culture factors of *Nannochloropsis* sp. followed by the Soxhlet extraction method yielded a lipid content of 61.5%. The extracted lipid was then transesterified with methanol to produce methyl esters (biodiesel) in 1.5 h, where potassium hydroxide (KOH) was used as a homogenous catalyst. Thin-layer chromatography (TLC) was done to ensure the conversion of *Nannochloropsis* sp. oil to biodiesel. The highest fatty acid methyl ester (FAME) formation from *Nannochloropsis* sp. was 80.24%. The output of gas chromatography- mass spectrometry (GC-MS) analysis proves that FAME comprises of high amount of oleic acid (C_{18:1}) 66.13% and palmitic acid (C_{16:0}) 12.38% respectively. *Nannochloropsis* sp. is a promising candidate for biodiesel production due to its composition of fatty acids and higher lipid content.

TABLE OF CONTENT

DECLARATION	
TITLE PAGE	
ACKNOWLEDGEMENTS	ii
ABSTRAK	iii
ABSTRACT	iv
TABLE OF CONTENT	v
LIST OF TABLES	ix
LIST OF FIGURES	x
LIST OF SYMBOLS	xii
LIST OF ABBREVIATIONS	xiii
LIST OF APPENDICES	xv
CHAPTER 1 INTRODUCTION	1
1.1 Background of the study	1
1.2 Problem statement	2
1.3 Objectives of the study	3
1.4 Scope of the study	4
1.5 Significance of the study	5
1.6 Thesis outline	5
CHAPTER 2 LITERATURE REVIEW	6
2.1 Palm oil industry	6
2.1.1 Palm oil trees	7
2.1.2 Brief history of palm oil in Malaysia	8
2.2 Oil palm biomass in Malaysia	10
2.3 Palm oil mill effluent (POME)	11

2.3.1	Properties of POME	12
2.3.2	Treatment of POME	13
2.3.3	POME as a source of nutrient for microalgae cultivation	16
2.4	Algae	17
2.4.1	Macroalgae	18
2.4.2	Microalgae	19
	2.4.2.1 <i>Nannochloropsis</i> sp.	21
2.5	Bioactive compounds	22
2.5.1	Lipids	24
	2.5.1.1 Fatty acids	26
2.6	Biofuel (Biodiesel) generations	27
2.7	Microalgae biodiesel production process	30
	2.7.1 Selection and isolation of strain	30
	2.7.2 Cultivation	30
	2.7.3 Harvesting	32
	2.7.4 Drying	33
	2.7.5 Lipid extraction	33
	2.7.6 Transesterification	35
CHAPTER 3 METHODOLOGY		36
3.1	Overall work plan of the study	36
3.2	Description of the study area	38
3.3	Collection of microalgae sample	38
3.4	Preparation of Conway media	39
3.5	Isolation and identification of microalgae	40
3.6	Pre-cultivation of <i>Nannochloropsis</i> sp.	41

3.7	Collection of wastewater (POME)	42
3.8	Preparation of POME medium	43
3.9	Cultivation of <i>Nannochloropsis</i> sp. under growth parameters	43
3.9.1	Cultivation of <i>Nannochloropsis</i> sp. under different POME concentration	43
3.9.2	Cultivation of <i>Nannochloropsis</i> sp. under different photoperiod regimes	44
3.9.3	Cultivation of <i>Nannochloropsis</i> sp. under different LED light wavelengths	44
3.10	Determination of <i>Nannochloropsis</i> sp. growth	45
3.10.1	UV-Visible spectrometry analysis	45
3.10.2	Dry weight (DW) measurements	45
3.11	Mass cultivation of <i>Nannochloropsis</i> sp. under optimized conditions	46
3.12	Harvesting of <i>Nannochloropsis</i> sp. biomass	46
3.13	Analysis of POME	47
3.14	Microalgae lipid extraction methods	47
3.14.1	Solvent extraction method	47
3.14.2	Soxhlet extraction method	48
3.15	Transesterification of <i>Nannochloropsis</i> sp. lipid	49
3.16	Analysis of fatty acid methyl ester (FAME)	49
3.16.1	Qualitative analysis of FAME	49
3.16.2	Quantitative analysis of FAME	50
3.17	Statistical analysis	51
CHAPTER 4 RESULTS AND DISCUSSION		52
4.1	Microalgal sample collection and isolation	52
4.2	Identification and morphological characteristics of <i>Nannochloropsis</i> sp.	53

4.3	Evaluation of <i>Nannochloropsis</i> sp. growth under different growth parameters	56
4.3.1	Evaluation of <i>Nannochloropsis</i> sp. growth in different POME concentrations	56
4.3.2	Evaluation of <i>Nannochloropsis</i> sp. growth in different photoperiod regimes	66
4.3.3	Evaluation of <i>Nannochloropsis</i> sp. growth in different LED light wavelengths	76
4.4	Mass cultivation of <i>Nannochloropsis</i> sp. under optimized parameters	84
4.5	Removal of nutrient	87
4.6	Extraction using Soxhlet extraction method	89
4.7	Transesterification of <i>Nannochloropsis</i> sp. oil	92
4.8	Qualitative analysis of FAME using TLC technique	94
4.9	Quantitative analysis of FAME using GC-MS	96
	CHAPTER 5 CONCLUSION	99
5.1	Conclusion	99
5.2	Recommendations	100
	REFERENCES	101
	LIST OF PUBLICATIONS	117
	APPENDICES	118

LIST OF TABLES

Table 2.1	Palm oil mill effluent (POME) characteristics	13
Table 2.2	The comparison of microalgae with other feedstocks of biodiesel	29
Table 3.1	Composition of Conway media	39
Table 4.1	Reading of growth rate (OD 680nm) and dry cell weight g L ⁻¹ under optimized conditions	85
Table 4.2	Statistical analysis of optimized parameters using ANOVA	86
Table 4.3	Characteristics of POME before and after cultivation of <i>Nannochloropsis</i> sp.	89
Table 4.4	Lipid composition of <i>Nannochloropsis</i> sp.	98

LIST OF FIGURES

Figure 2.1	Palm oil's top five producers in the world (2016/2017)	6
Figure 2.2	Cross-section of an oil palm fruit	7
Figure 2.3	Oil palm production efficiency compared to other major oil crops	8
Figure 2.4	Types of oil palm biomass	10
Figure 2.5	A common ponding treatment system for palm oil mill effluent (POME)	15
Figure 2.6	Primary modes of algae reproduction: (A) asexual reproduction and (B) sexual reproduction	18
Figure 2.7	Bio-refinery of constituent derived from microalgae biomass	23
Figure 3.1	The overall work plan of the study	37
Figure 3.2	The map of Pahang state and location of Pantai Teluk Cempedak	38
Figure 3.3	The 1000 mL of Erlenmeyer flask with a 1:1 ratio of culture and fresh Conway media.	42
Figure 3.4	The location of POME sample collected: (A) Map of LCSB Palm Oil Mill Lepar and (B) Logo of LCSB Palm Oil Mill Lepar	42
Figure 3.5	Cultivation of <i>Nannochloropsis</i> sp. under different POME concentrations	44
Figure 3.6	Cultivation of <i>Nannochloropsis</i> sp. under different LED light wavelengths: (A) Control, (B) Blue, (C) Red, and (D) Green	45
Figure 3.7	Ground biomass of <i>Nannochloropsis</i> sp.	47
Figure 3.8	Lipid extraction of <i>Nannochloropsis</i> sp. using Soxhlet method	49
Figure 4.1	The fluorescence microscope image at 100X of microalgae species isolated: (A) <i>Chlorella</i> sp., (B) <i>Amphora</i> sp., (C) <i>Gyrosigma</i> sp., (D) <i>Tetraselmis</i> sp., (E) <i>Spirulina</i> sp., and (F) <i>Nannochloropsis</i> sp.	53
Figure 4.2	Fluorescence micrograph of <i>Nannochloropsis</i> sp. under 100X	54
Figure 4.3	FESEM micrograph of <i>Nannochloropsis</i> sp. under (A) 1,000X, (B) 5,500X, (C) 11,000X, and (D) 20,000X.	54
Figure 4.4	The growth curve of <i>Nannochloropsis</i> sp. cultivated under control, 5, 10, 15, and 20% POME media.	58
Figure 4.5	The growth rate (OD680 nm) and dry weight (g L ⁻¹) of <i>Nannochloropsis</i> sp. cultivated under control media.	60
Figure 4.6	The growth rate (OD680 nm) and dry weight (g L ⁻¹) of <i>Nannochloropsis</i> sp. cultivated under 5% POME media.	62
Figure 4.7	The growth rate (OD680 nm) and dry weight (g L ⁻¹) of <i>Nannochloropsis</i> sp. cultivated under 10% POME media.	63
Figure 4.8	The growth rate (OD680 nm) and dry weight (g L ⁻¹) of <i>Nannochloropsis</i> sp. cultivated under 15% POME media.	65

Figure 4.9	The growth rate (OD680 nm) and dry weight (g L ⁻¹) of <i>Nannochloropsis</i> sp. cultivated under 20% POME media.	66
Figure 4.10	The growth curve of <i>Nannochloropsis</i> sp. cultivated under 24:00, 18:06, 12:12, and 06:18 h L/D.	69
Figure 4.11	The growth rate (OD680 nm) and dry weight (g L ⁻¹) of <i>Nannochloropsis</i> sp. cultivated at 24:00 h L/D cycle	71
Figure 4.12	The growth rate (OD680 nm) and dry weight (g L ⁻¹) of <i>Nannochloropsis</i> sp. cultivated at 18:06 h L/D cycle	73
Figure 4.13	The growth rate (OD680 nm) and dry weight (g L ⁻¹) of <i>Nannochloropsis</i> sp. cultivated at 12:12 h L/D cycle	74
Figure 4.14	The growth rate (OD680 nm) and dry weight (g L ⁻¹) of <i>Nannochloropsis</i> sp. cultivated at 06:18 h L/D cycle	75
Figure 4.15	The growth curve of <i>Nannochloropsis</i> sp. cultivated in 10% POME under 18:06 h L/D with fluorescent, blue, red, and green LED lights	78
Figure 4.16	The growth rate (OD680 nm) and dry weight (g L ⁻¹) of <i>Nannochloropsis</i> sp. in 10% POME under 18:06 h L/D with fluorescent light	79
Figure 4.17	The growth rate (OD680 nm) and dry weight (g L ⁻¹) of <i>Nannochloropsis</i> sp. cultivated in 10% POME under 18:06 h L/D with blue light	81
Figure 4.18	The growth rate (OD680 nm) and dry weight (g L ⁻¹) of <i>Nannochloropsis</i> sp. cultivated in 10% POME under 18:06 h L/D with red light	83
Figure 4.19	The growth rate (OD680 nm) and dry weight (g L ⁻¹) of <i>Nannochloropsis</i> sp. cultivated in 10% POME under 18:06 h L/D with green light	84
Figure 4.20	The growth curve of <i>Nannochloropsis</i> sp. cultivated under optimized conditions	86
Figure 4.21	TLC plate illustrating the separation and presence of TAG and methyl ester based on different reaction times	96
Figure 4.22	Chromatogram of methyl ester from <i>Nannochloropsis</i> sp. using GC-MS	98

LIST OF APPENDICES

Appendix A:	Absorbance reading of <i>Nannochloropsis</i> sp. cultivated under different POME concentrations	119
Appendix B:	Appendix B: Dry weight of <i>Nannochloropsis</i> sp. cultivated under different POME concentrations	120
Appendix C:	Absorbance reading of <i>Nannochloropsis</i> sp. cultivated under different photoperiod regimes	121
Appendix D:	Dry weight of <i>Nannochloropsis</i> sp. cultivated under different photoperiod regimes	122
Appendix E:	Absorbance reading of <i>Nannochloropsis</i> sp. cultivated under different LED light wavelengths	123
Appendix F:	Dry weight of <i>Nannochloropsis</i> sp. cultivated under different LED light wavelengths	124
Appendix G:	Absorbance reading of <i>Nannochloropsis</i> sp. cultivated under optimized condition	125
Appendix H:	Dry weight of <i>Nannochloropsis</i> sp. cultivated under optimized condition	126
Appendix I:	Characteristics of POME before and after cultivation	127

REFERENCES

- Abdullah, M. A., Ahmad, A., Shah, S. M. U., Shanab, S. M. M., Ali, H. E. A., Abo-State, M. A. M., & Othman, M. F. (2016). Integrated algal engineering for bioenergy generation, effluent remediation, and production of high-value bioactive compounds. *Biotechnology and Bioprocess Engineering*, *21*(2), 236–249.
- Abomohra, A. E. F., & Almutairi, A. W. (2020). A close-loop integrated approach for microalgae cultivation and efficient utilization of agar-free seaweed residues for enhanced biofuel recovery. *Bioresource Technology*, *317*, 1–11.
- Abomohra, A. E. F., Shang, H., El-Sheekh, M., Eladel, H., Ebaid, R., Wang, S., & Wang, Q. (2019). Night illumination using monochromatic light-emitting diodes for enhanced microalgal growth and biodiesel production. *Bioresource Technology*, *288*, 1-8.
- Abou-shanab, R. A. I., El-dalatony, M. M., El-sheekh, M. M., Ji, M., & Salama, E. (2014). Cultivation of a new microalga, *Micractinium reisseri*, in municipal wastewater for nutrient removal, biomass, lipid, and fatty acid production. *Biotechnology and Bioprocess Engineering*, *518*, 510–518.
- Akhbari, A., Kutty, P. K., Chuen, O. C., & Ibrahim, S. (2020). A study of palm oil mill processing and environmental assessment of palm oil mill effluent treatment. *Environmental Engineering Research*, *25*(2), 212–221.
- Alqadi, M. A., Taib, S. M., Din, M. F. M., & Kamyab, H. (2017). Effect of photoperiod on the growth of *Chlamydomonas*. *Malaysian Journal of Civil Engineering*, *1*, 69–78.
- Al-Qasbi, M., Raut, N., Talebi, S., Al-Rajhi, S., & Al-Barwani, T. (2012). A review of effect of light on microalgae growth. *Proceedings of the World Congress on Engineering 2012*, *I*(2), 1–7.
- Ananthi, V., Raja, R., Carvalho, I. S., Brindhadevi, K., Pugazhendhi, A., & Arun, A. (2021). A realistic scenario on microalgae-based biodiesel production: Third generation biofuel. *Fuel*, *284*, 1–10.
- Aratboni, H. A., Rafiei, N., Garcia-Granados, R., Alemzadeh, A., & Morones-Ramírez, J. R. (2019). Biomass and lipid induction strategies in microalgae for biofuel production and other applications. *Microbial Cell Factories*, *18*(1), 1–17.
- Archambault, S., Starbuck Downes, C. M., Van Voorhies, W., Erickson, C. A., & Lammers, P. (2014). *Nannochloropsis* sp. algae for use as biofuel: Analyzing a translog production function using data from multiple sites in the southwestern United States. *Algal Research*, *6*, 124–131.
- Atta, M., Idris, A., Bukhari, A., & Wahidin, S. (2013). Intensity of blue LED light: A potential stimulus for biomass and lipid content in freshwater microalgae

Chlorella vulgaris. *Bioresource Technology*, 148, 373–378.

- Babuskin, S., Radhakrishnan, K., Babu, P. A. S., Sivarajan, M., & Sukumar, M. (2014). Effect of photoperiod, light intensity, and carbon sources on biomass and lipid productivities of *Isochrysis galbana*. *Biotechnology Letters*, 36(8), 1653–1660.
- Badar, S. N., Yaakob, Z., & Timmiati, S. N. (2017). Growth evaluation of microalgae isolated from palm oil mill effluent in synthetic media. *Malaysian Journal of Analytical Science*, 21(1), 82–94.
- Benzidane, D., Hamed Mohamed, B., & Abi-Ayad Sidi-Mohammed, E.-A. (2017). Biodiesel production from marine microalgae *Nannochloropsis gaditana* by in situ transesterification process. *African Journal of Biotechnology*, 16(22), 1270–1277.
- Bhalamurugan, G. L., Valerie, O., & Mark, L. (2018). Valuable bioproducts obtained from microalgal biomass and their commercial applications: A review. *Environmental Engineering Research*, 23(3), 229–241.
- Bhuyar, P., Farez, F., Ab. Rahim, M. H., Maniam, G. P., & Govindan, N. (2021). Removal of nitrogen and phosphorus from agro-industrial wastewater by using microalgae collected from coastal region of peninsular Malaysia. *African Journal of Biological Sciences*, 3(1), 58–66.
- Bhuyar, P., Yusoff, M. M., Rahim, M. H. A., Sundararaju, S., Maniam, G. P., & Govindan, N. (2020). Effect of plant hormones on the production of biomass and lipid extraction for biodiesel production from microalgae *Chlorella* sp. *Journal of Microbiology, Biotechnology, and Food Sciences*, 9(4), 671–674.
- Bombo, K., Lekgoba, T., Azeez, O., & Muzenda, E. (2021). The Sustainability of Biodiesel Synthesis from Different Feedstocks: A Review. *Petroleum and Coal*, 63(2), 284–291.
- Budhwani, A. A. A., Maqbool, A., Hussain, T., & Syed, M. N. (2019). Production of biodiesel by enzymatic transesterification of non-edible *Salvadora persica* (Pilu) oil and crude coconut oil in a solvent-free system. *Bioresources and Bioprocessing*, 6(1), 1-9.
- Buschmann, A. H., & Camus, C. (2019). An introduction to farming and biomass utilisation of marine macroalgae. *Phycologia*, 58(5), 443–445.
- Cagliari, A., Margis, R., Dos Santos Maraschin, F., Turchetto-Zolet, A. C., Loss, G., & Margis-Pinheiro, M. (2011). Biosynthesis of triacylglycerols (TAGs) in plants and algae. *International Journal of Plant Biology*, 2(1), 40–52.
- Cardoso, L. G., Duarte, J. H., Costa, J. A. V., de Jesus Assis, D., Lemos, P. V. F., Druzian, J. I., de Souza, C. O., Nunes, I. L., & Chinalia, F. A. (2020). *Spirulina* sp. as a bioremediation agent for aquaculture wastewater: Production of high added value compounds and estimation of theoretical biodiesel. *Bioenergy Research*. 1,1-11.

- Carmichael, W. W. (1992). Cyanobacteria secondary metabolites—the cyanotoxins. *Journal of Applied Bacteriology*, 72(6), 445–459.
- Chattopadhyay, S., Das, S., & Sen, R. (2011). Rapid and precise estimation of biodiesel by high-performance thin-layer chromatography. *Applied Energy*, 88(12), 5188–5192.
- Cheah, W. Y., Show, P. L., Juan, J. C., Chang, J. S., & Ling, T. C. (2018). Microalgae cultivation in palm oil mill effluent (POME) for lipid production and pollutants removal. *Energy Conversion and Management*, 174, 430–438.
- Chen, C. Y., Chen, Y. C., Huang, H. C., Huang, C. C., Lee, W. L., & Chang, J. S. (2013). Engineering strategies for enhancing the production of eicosapentaenoic acid (EPA) from an isolated microalga *Nannochloropsis oceanica* CY2. *Bioresource Technology*, 147, 160–167.
- Colusse, G. A., Mendes, C. R. B., Duarte, M. E. R., Carvalho, J. C. de, & Noseda, M. D. (2020). Effects of different culture media on physiological features and laboratory scale production cost of *Dunaliella salina*. *Biotechnology Reports*, 27, 1-9.
- Das, P., Lei, W., Aziz, S. S., & Obbard, J. P. (2011). Enhanced algae growth in both phototrophic and mixotrophic culture under blue light. *Bioresource Technology*, 102(4), 3883–3887.
- Dianursanti, Sistiafi, A. G., & Putri, D. N. (2018). Biodiesel synthesis from *Nannochloropsis oculata* and *Chlorella vulgaris* through transesterification process using NaOH/zeolite heterogeneous catalyst. *IOP Conference Series: Earth and Environmental Science*, 105(1), 1-6.
- Diaz-Pulido, G., & McCook, L. J. (2008). Environmental Status: Macroalgae (Seaweeds). *Great Barrier Reef Marine Park Authority*, 1, 1–44.
- Ding, G. T., Yaakob, Z., Takriff, M. S., Salihon, J., & Abd Rahaman, M. S. (2016). Biomass production and nutrients removal by a newly-isolated microalgal strain *Chlamydomonas* sp. in palm oil mill effluent (POME). *International Journal of Hydrogen Energy*, 41(8), 4888–4895.
- Dolganyuk, V., Andreeva, A., Budenkova, E., Sukhikh, S., Babich, O., Ivanova, S., Prosekov, A., & Ulrikh, E. (2020). Study of morphological features and determination of the fatty acid composition of the microalgae lipid complex. *Biomolecules*, 10(11), 1–15.
- Duarte, J. H., Cardoso, L. G., de Souza, C. O., Nunes, I. L., Druzian, J. I., de Moraes, M. G., & Costa, J. A. V. (2020). Brackish groundwater from Brazilian backlands in *Spirulina* cultures: Potential of carbohydrate and polyunsaturated fatty acid production. *Applied Biochemistry and Biotechnology*, 190(3), 907–917.
- Efroymsen, R. A., Jager, H. I., Mandal, S., Parish, E. S., & Mathews, T. J. (2020). Better management practices for environmentally sustainable production of microalgae

and algal biofuels. *Journal of Cleaner Production*, 1, 1-14.

- Elystia, S., Muria, S. R., & Erlangga, H. F. (2020). Cultivation of *Chlorella pyrenoidosa* as a raw material for the production of biofuels in palm oil mill effluent medium with the addition of urea and triple super phosphate. *Environmental Health Engineering and Management*, 7(1), 1–6.
- Emparan, Q., Harun, R., & Jye, Y. S. (2019). Phycoremediation of treated palm oil mill effluent (TPOME) using *Nannochloropsis* sp. cells immobilized in the biological sodium alginate beads: Effect of POME concentration. *BioResources*, 14(4), 9429–9443.
- Emparan, Q., Jye, Y. S., Danquah, M. K., & Harun, R. (2020). Cultivation of *Nannochloropsis* sp. microalgae in palm oil mill effluent (POME) media for phycoremediation and biomass production: Effect of microalgae cells with and without beads. *Journal of Water Process Engineering*, 33, 1-6.
- Fakhri, M., Sanudi, Arifin, N. B., Ekawati, A. W., Yuniarti, A., & Hariati, A. M. (2017). Effect of photoperiod regimes on growth, biomass, and pigment content of *Nannochloropsis* sp. BJ17. *Asian Journal of Microbiology, Biotechnology, and Environmental Sciences*, 19(2), 263–267.
- Fernando, J. S. R., Premaratne, M., Dinalankara, D. M. S. D., Perera, G. L. N. J., & Ariyadasa, T. U. (2021). Cultivation of microalgae in palm oil mill effluent (POME) for astaxanthin production and simultaneous phycoremediation. *Journal of Environmental Chemical Engineering*, 9(4), 1-13.
- Ferreira, G. F., Ríos Pinto, L. F., Maciel Filho, R., & Fregolente, L. V. (2019). A review on lipid production from microalgae: Association between cultivation using waste streams and fatty acid profiles. *Renewable and Sustainable Energy Reviews*, 109, 448–466.
- Fithriani, D., Ambarwaty, D., & Nurhayati. (2019). Identification of bioactive compounds from *Nannochloropsis* sp. *IOP Conference Series: Earth and Environmental Science*, 404(1), 1–5.
- Galasso, C., Gentile, A., Orefice, I., Ianora, A., Bruno, A., Noonan, D. M., Sansone, C., Albin, A., & Brunet, C. (2019). Microalgal derivatives as potential nutraceutical and food supplements for human health: A focus on cancer prevention and interception. *Nutrients*, 11(6), 1–22
- Gamal, A. A. El. (2010). Biological importance of marine algae. *Saudi Pharmaceutical Journal*, 18(1), 1–25.
- Ghasemi Naghdi, F., González González, L. M., Chan, W., & Schenk, P. M. (2016). Progress on lipid extraction from wet algal biomass for biodiesel production. *Microbial Biotechnology*, 9(6), 718–726.

- Ghosh, U. K. (2018). An approach for phycoremediation of different wastewaters and biodiesel production using microalgae. *Environmental Science and Pollution Research*, 25, 18673–18681.
- Gim, G. H., & Kim, S. W. (2018). Optimization of cell disruption and transesterification of lipids from *Botryococcus braunii* LB572. *Biotechnology and Bioprocess Engineering*, 23(5), 550–556.
- Glemser, M., Heining, M., Schmidt, J., Becker, A., Garbe, D., Buchholz, R., & Brück, T. (2016). Application of light-emitting diodes (LEDs) in cultivation of phototrophic microalgae: current state and perspectives. *Applied Microbiology and Biotechnology*, 100(3), 1077–1088.
- Goncalves, A. L., Pires, J. C. M., & Simoes, M. (2013). *Biodiesel from microalgal oil extraction*. Springer Dordrecht Heidelberg.
- Govindan, N., Maniam, G. P., Yusoff, M. M., Mohd, M. H., Chatsungnoen, T., Ramaraj, R., & Chisti, Y. (2020). Statistical optimization of lipid production by the diatom *Gyrosigma* sp. grown in industrial wastewater. *Journal of Applied Phycology*, 32(1), 375–387.
- Griffiths, M. J., Dicks, R. G., Richardson, C., & Harrison, S. T. (2011). Advantages and challenges of microalgae as a source of oil for biodiesel. In *Biodiesel - Feedstocks and Processing Technologies*, 177–200.
- Gunawan, T. J., Ikhwan, Y., Restuhadi, F., & Pato, U. (2018). Effect of light Intensity and photoperiod on growth of *Chlorella pyrenoidosa* and CO₂ biofixation. *E3S Web of Conferences*, 31, 1–7.
- Guldhe, A., Ansari, F. A., Singh, P., & Bux, F. (2017). Heterotrophic cultivation of microalgae using aquaculture wastewater: A biorefinery concept for biomass production and nutrient remediation. *Ecological Engineering*, 99, 47–53.
- Guo, H., & Fang, Z. (2020). Effect of light quality on the cultivation of *Chlorella pyrenoidosa*. *E3S Web of Conferences*, 143, 1–6.
- Gupta, S., Pawar, S. B., & Pandey, R. A. (2019). Current practices and challenges in using microalgae for treatment of nutrient rich wastewater from agro-based industries. *Science of the Total Environment*, 687, 1107–1126.
- Hadiyanto, H., Soetrinanto, D., Silviana, S., Mahdi, M. Z., & Titisari, Y. N. (2017). Evaluation of growth and biomass productivity of marine microalga *Nannochloropsis* sp. cultured in palm oil mill effluent (POME). *Philippine Journal of Science*, 146(4), 355–360.
- Hariz, H. B., & Takriff, M. S. (2017). Palm oil mill effluent treatment and CO₂ sequestration by using microalgae—sustainable strategies for environmental protection. *Environmental Science and Pollution Research*, 24(25), 20209–20240.

- Hassan, M. A., Yacob, S., Hung, Y.-T., & Shirai, Y. (2005). Treatment of palm oil wastewaters. *Waste Treatment in the Food Processing Industry, 1*, 101–117.
- He, Y., Wang, X., Wei, H., Zhang, J., Chen, B., & Chen, F. (2019). Direct enzymatic ethanolysis of potential *Nannochloropsis* biomass for co-production of sustainable biodiesel and nutraceutical eicosapentaenoic acid. *Biotechnology for Biofuels, 12*(1), 1–15.
- Hidalgo, P., Toro, C., Ciudad, G., & Navia, R. (2013). Advances in direct transesterification of microalgal biomass for biodiesel production. *Reviews in Environmental Science and Biotechnology, 12*(2), 179–199.
- Hulatt, C. J., Wijffels, R. H., Bolla, S., & Kiron, V. (2017). Production of fatty acids and protein by *Nannochloropsis* in flat-plate photobioreactors. *PLoS ONE, 12*(1), 1–17.
- Jafari, A., Esmailzadeh, F., Mowla, D., Sadatshojaei, E., Heidari, S., & Wood, D. A. (2021). New insights to direct conversion of wet microalgae impregnated with ethanol to biodiesel exploiting extraction with supercritical carbon dioxide. *Fuel, 285*, 1–15.
- Jankowska, E., Sahu, A. K., & Oleskiewicz-Popiel, P. (2017). Biogas from microalgae: Review on microalgae's cultivation, harvesting, and pretreatment for anaerobic digestion. *Renewable and Sustainable Energy Reviews, 75*, 692–709.
- Jayakumar, S., Bhuyar, P., Pugazhendhi, A., Rahim, M. H. A., Maniam, G. P., & Govindan, N. (2021). Effects of light intensity and nutrients on the lipid content of marine microalga (diatom) *Amphiprora* sp. for promising biodiesel production. *Science of the Total Environment, 768*, 1–11.
- Junior, W. G. M., Gorgich, M., Corrêa, P. S., Martins, A. A., Mata, T. M., & Caetano, N. S. (2020). Microalgae for biotechnological applications: Cultivation, harvesting and biomass processing. *Aquaculture, 528*, 1–14.
- Kalil Rahiman, M., Balakrishnan, K., Venkatachalam, R., Murugappan, S., & Santhoshkumar, S. (2018). Study of diesel with oxygenated fuel blends for its prominence using high performance thin layer chromatography. *International Journal of Engineering and Advanced Technology, 8*(2), 45–53.
- Kamyab, H., Chelliapan, S., Md, M. F., Rezanian, S., Din, M. F. M., Rezanian, S., Khademi, T., & Kumar, A. (2018). Palm oil mill effluent as an environmental pollutant. *Intech, 32*, 137–144.
- Kamyab, H., Md Din, M. F., Ponraj, M., Keyvanfar, A., Rezanian, S., Taib, S. M., & Abd Majid, M. Z. (2016). Isolation and screening of microalgae from agro-industrial wastewater (POME) for biomass and biodiesel sources. *Desalination and Water Treatment, 57*(60), 29118–29125.

- Kanagesan, K., Palanisamy, K. M., Maniam, G. P., Rahim, A., & Govindan, N. (2021). Biodiesel production by microalgae *Nannochloropsis* sp. grown in palm oil mill effluent. *Maejo International Journal of Energy and Environmental Communication*, 2(3), 24–31.
- Kanda, H., Hoshino, R., Murakami, K., Wahyudiono, Zheng, Q., & Goto, M. (2020). Lipid extraction from microalgae covered with biomineralized cell walls using liquefied dimethyl ether. *Fuel*, 262, 1–8.
- Kendirlioglu, G., Agirman, N., & Cetin, A. K. (2015). The effects of photoperiod on the growth, protein amount, and pigment content of *Chlorella vulgaris*. *Turkish Journal of Science & Technology*, 10(2), 7–10.
- Khoeyi, Z. A., Seyfabadi, J., & Ramezanpour, Z. (2012). Effect of light intensity and photoperiod on biomass and fatty acid composition of the microalgae, *Chlorella vulgaris*. *Aquaculture International*, 20(1), 41–49.
- Kim, S. W., Xiao, M., & Shin, H. J. (2016). Fractionation and lipase-catalyzed conversion of microalgal lipids to biodiesel. *Biotechnology and Bioprocess Engineering*, 21(6), 743–750.
- Kim, C. W., Sung, M. G., Nam, K., Moon, M., Kwon, J. H., & Yang, J. W. (2014). Effect of monochromatic illumination on lipid accumulation of *Nannochloropsis gaditana* under continuous cultivation. *Bioresource Technology*, 159, 30–35.
- Kim, T. H., Lee, Y., Han, S. H., & Hwang, S. J. (2013). The effects of wavelength and wavelength mixing ratios on microalgae growth and nitrogen, phosphorus removal using *Scenedesmus* sp. for wastewater treatment. *Bioresource Technology*, 130, 75–80.
- Koay, A. (2014, March 24). Green wealth in oil palm. *The Star*. <https://www.thestar.com.my/news/environment/2014/03/24/green-wealth-in-oil-palm/>
- Koeh, A. K., Kumar, A., & Siagi, Z. O. (2020). In situ transesterification of *Spirulina* microalgae to produce biodiesel using microwave irradiation. *Journal of Energy*, 2020, 1–10.
- Koyande, A. K., Chew, K. W., Rambabu, K., Tao, Y., Chu, D. T., & Show, P. L. (2019). Microalgae: A potential alternative to health supplementation for humans. *Food Science and Human Wellness*, 8(1), 16–24.
- Krzemińska, I., Nosalewicz, A., Reszczyńska, E., & Pawlik-Skowrońska, B. (2020). Enhanced light-induced biosynthesis of fatty acids suitable for biodiesel production by the yellow-green alga *Eustigmatos magnus*. *Energies*, 13(22), 1–14.
- Kumar, R., Ghosh, A. K., & Pal, P. (2020). Synergy of biofuel production with waste remediation along with value-added co-products recovery through microalgae

cultivation: A review of membrane-integrated green approach. *Science of the Total Environment*, 698, 1–23.

- Lage, S., Gojkovic, Z., Funk, C., & Gentili, F. G. (2018). Algal biomass from wastewater and flue gases as a source of bioenergy. *Energies*, 11(3), 1–30.
- Li, Yajun, Fei, X., & Deng, X. (2012). Novel molecular insights into nitrogen starvation-induced triacylglycerols accumulation revealed by differential gene expression analysis in green algae *Micractinium pusillum*. *Biomass and Bioenergy*, 42, 199–211.
- Li, Yan, Ghasemi Naghdi, F., Garg, S., Adarme-Vega, T. C., Thurecht, K. J., Ghafor, W. A., Tannock, S., & Schenk, P. M. (2014). A comparative study: The impact of different lipid extraction methods on current microalgal lipid research. *Microbial Cell Factories*, 13(1), 1–9.
- Lu, W., Liu, S., Lin, Z., & Lin, M. (2020). Enhanced microalgae growth for biodiesel production and nutrients removal in raw swine wastewater by carbon sources supplementation. *Waste and Biomass Valorization*, 1, 1–9.
- Ma, X. N., Chen, T. P., Yang, B., Liu, J., & Chen, F. (2016). Lipid production from *Nannochloropsis*. *Marine Drugs*, 14(4), 1–18.
- Madaki, Y. S., & Seng, L. (2013). Palm oil mill effluent (POME) from Malaysia palm oil mills : Waste or Resource. *International Journal of Science, Environment and Technology*, 2(6), 1138–1155.
- Malakootian, M., Hatami, B., Dowlatshahi, S., & Rajabizadeh, A. (2015). Optimization of culture media for lipid production by *Nannochloropsis oculata* for Biodiesel production. *Environmental Health Engineering and Management Journal*, 2(3), 141–147.
- Maluin, F. N., Hussein, M. Z., & Idris, A. S. (2020). An overview of the oil palm industry: Challenges and some emerging opportunities for nanotechnology development. *Agronomy*, 10(3), 1-20.
- Markou, G., Chatzipavlidis, I., & Georgakakis, D. (2012). Effects of phosphorus concentration and light intensity on the biomass composition of *Arthrospira (Spirulina) platensis*. *World Journal of Microbiology and Biotechnology*, 28(8), 2661–2670.
- Marwan, Suhendrayatna, & Indarti, E. (2015). Preparation of biodiesel from microalgae and palm oil by direct transesterification in a batch microwave reactor. *Journal of Physics: Conference Series*, 622(1), 1-8.
- Matos, Â. P., Cavanholi, M. G., Moecke, E. H. S., & Sant'Anna, E. S. (2017). Effects of different photoperiod and trophic conditions on biomass, protein, and lipid production by the marine alga *Nannochloropsis gaditana* at optimal concentration of desalination concentrate. *Bioresource Technology*, 224, 490–497.

- Melsasail, K., Awan, A., Papilaya, P. M., & Rumahlatu, D. (2018). The ecological structure of macroalgae community (Seagrass) on various zones in the coastal waters of Nusalaut Island, Central Maluku District, Indonesia. *AACL Bioflux*, *11*(4), 957–966.
- Morais, M. G. De, Vaz, B. D. S., Morais, E. G. De, & Costa, J. A. V. (2015). Biologically Active Metabolites Synthesized by Microalgae. *BioMed Research International*, *15*, 1-15.
- Mostafa, S. S. M., & El-Gendy, N. S. (2017). Evaluation of fuel properties for microalgae *Spirulina platensis* bio-diesel and its blends with Egyptian petro-diesel. *Arabian Journal of Chemistry*, *10*, 2040–S2050.
- Mosunmola, A. G., & Olatunde, S. K. (2020). Palm oil mill effluents (POME) and its pollution potentials : A biodegradable prevalence. *Journal of Pollution Effects & Control*, *8*(5), 1–10.
- Murphy, D. J. (2014). The future of oil palm as a major global crop: Opportunities and challenges. *Journal of Oil Palm Research*, *26*, 1–24.
- Musharraf, S. G., Ahmed, M. A., Zehra, N., Kabir, N., Choudhary, M. I., & Rahman, A. ur. (2012). Biodiesel production from microalgal isolates of southern Pakistan and quantification of FAMES by GC-MS/MS analysis. *Chemistry Central Journal*, *6*(1), 1–10.
- Myers, J. A., Curtis, B. S., & Curtis, W. R. (2013). Converter OD g biomass. *BMC Biophysics*, *6*(4), 1-15.
- Naghdi, F. G., Thomas-Hall, S. R., Durairatnam, R., Pratt, S., & Schenk, P. M. (2014). Comparative effects of biomass pre-treatments for direct and indirect transesterification to enhance microalgal lipid recovery. *Frontiers in Energy Research*, *2*, 1–10.
- Nomanbhay, S., Salman, B., Hussain, R., & Ong, M. Y. (2017). Microwave pyrolysis of lignocellulosic biomass—a contribution to power Africa. *Energy, Sustainability and Society*, *7*(1), 1-24.
- Norhidayu, A., Nur-Syazwani, M., Radzil, R., Amin, I., & Balu, N. (2017). The production of crude palm oil in Malaysia. *International Journal of Economics and Management*, *11*(3), 591–606.
- Nur, M. M. Azimatun, Garcia, G. M., Boelen, P., & Buma, A. G. J. (2020). Influence of photodegradation on the removal of color and phenolic compounds from palm oil mill effluent by *Arthrospira platensis*. *Journal of Applied Phycology*, *1*, 1–15.
- Nur, Muhamad Maulana Azimatun, & Buma, A. G. J. (2019). Opportunities and challenges of microalgal cultivation on wastewater, with special focus on palm oil mill effluent and the production of high value compounds. *Waste and Biomass Valorization*, *10*(8), 2079–2097.

- Nur, M. M. A., Kristanto, D., Setyoningrum, T. M., & Budiaman, I. G. (2017). Utilization of microalgae cultivated in palm oil mill wastewater to produce lipid and carbohydrate by employing microwave-assisted irradiation. *Recent Innovations in Chemical Engineering*, 9(2), 107–116.
- Omar, S. A. S. (2019). Proposal for palm oil mill effluent (POME) treatment at source to reclaim water. *American Based Research Journal*, 8(5), 2304–7151.
- Onoja, E., Chandren, S., Abdul Razak, F. I., Mahat, N. A., & Wahab, R. A. (2019). Oil palm (*Elaeis guineensis*) biomass in Malaysia: The present and future prospects. *Waste and Biomass Valorization*, 10(8), 2099–2117.
- Osman, N. A., Ujang, F. A., Roslan, A. M., Ibrahim, M. F., & Hassan, M. A. (2020). The effect of palm oil mill effluent final discharge on the characteristics of *Pennisetum purpureum*. *Scientific Reports*, 10(1), 1–10.
- Palanisamy, K. M., Paramasivam, P., Maniam, G. P., Rahim, M. H. A., Govindan, N., & Chisti, Y. (2021). Production of lipids by *Chaetoceros affinis* in media based on palm oil mill effluent. *Journal of Biotechnology*, 327, 86–96.
- Palanisamy, K. M., Paramasivam, P., Maniam, G. P., Rahim, M. H. A., Ramaraj, R., & Govindan, N. (2019). Palm oil mill effluent as a potential medium for microalgae *Chlorella* sp. cultivation for lipid production. *Maejo International Journal of Energy and Environmental Communication*, 1, 1–7.
- Pangestuti, R., & Kim, S. K. (2011). Biological activities and health benefit effects of natural pigments derived from marine algae. *Journal of Functional Foods*, 3(4), 255–266.
- Panis, G., & Carreon, J. R. (2016). Commercial astaxanthin production derived by green alga *Haematococcus pluvialis*: A microalgae process model and a techno-economic assessment all through production line. *Algal Research*, 18, 175–190.
- Paramasivam, P., Palanisamy, K. M., Jayakumar, S., Govindan, N., Rahim, M. H. A., & Maniam, G. P. (2021). Lipid production from *Nannochloropsis* sp. grown in palm oil mill effluent. *International Conference of Sustainable Earth Resources Engineering 2020*, 641, 2–9.
- Pattanaik, A., Sukla, L. B., & Pradhan, D. (2018). Effect of LED lights on the growth of microalgae. *Inglomayor*, 14, 14–24.
- Peng, X., Meng, F., Wang, Y., Yi, X., & Cui, H. (2020). Effect of pH, temperature, and CO₂ concentration on growth and lipid accumulation of *Nannochloropsis* sp. MASCC 11. *Journal of Ocean University of China*, 19(5), 1183–1192.
- Ponraj, M., & Din, M. F. M. (2013). Effect of light/dark cycle on biomass and lipid productivity by *Chlorella pyrenoidosa* using palm oil mill effluent (POME). *Journal of Scientific and Industrial Research*, 72(11), 703–706.

- Poyadji, K., Stylianou, M., Agapiou, A., Kallis, C., & Kokkinos, N. (2018). Determination of quality properties of low-grade biodiesel and its heating oil blends. *Environments - MDPI*, 5(9), 1–16.
- Pradana, Y. S., Azmi, F. A., Masruri, W., & Hartono, M. (2018). biodiesel production from wet spirulina sp . by one-step. *MATEC Web of Conferences*, 156, 1–4.
- Pugliese, A., Biondi, L., Bartocci, P., & Fantozzi, F. (2020). Selenastrum capricornutum a new strain of algae for biodiesel production. *Fermentation*, 6(2), 1–12.
- Purkan, P., Nidianti, E., Abdulloh, A., Safa, A., Retnowati, W., Soemarjati, W., Nurlaila, H., & Wook Kim, S. (2019). Biodiesel production by lipids from Indonesian strain of microalgae *Chlorella vulgaris*. *Open Chemistry*, 17(1), 919–926.
- Putri, D. S., Astuti, S. P., & Alaa, S. (2019). The growth of microalgae *Chlorococcum* sp. isolated from Ampenan estuary of Lombok Island in Walne's medium. *AIP Conference Proceedings*, 2199, 1–6.
- Rahman, M. A., Aziz, M. A., Ruhul, A. M., & Rashid, M. M. (2017). Biodiesel production process optimization from *Spirulina maxima* microalgae and performance investigation in a diesel engine. *Journal of Mechanical Science and Technology*, 31(6), 3025–3033.
- Rajkumar, R., Yaakob, Z., & Takriff, M. S. (2014). Potential of the micro and macro algae for biofuel production: A brief review. *BioResources*, 9(1), 1606–1633.
- Ramluckan, K., Moodley, K. G., & Bux, F. (2014). An evaluation of the efficacy of using selected solvents for the extraction of lipids from algal biomass by the soxhlet extraction method. *Fuel*, 116, 103–108.
- Rashid, N., Ur Rehman, M. S., Sadiq, M., Mahmood, T., & Han, J. I. (2014). Current status, issues, and developments in microalgae derived biodiesel production. *Renewable and Sustainable Energy Reviews*, 40, 760–778.
- Ratomski, P., & Hawrot-Paw, M. (2021). Production of *Chlorella vulgaris* biomass in tubular photobioreactors during different culture conditions. *Applied Sciences*, 11(7), 1–13.
- Raven, J. A., & Giordano, M. (2014). Algae. *Current Biology*, 24(13), 590–595.
- Ren, H., Tuo, J., Addy, M. M., Zhang, R., Lu, Q., Anderson, E., Chen, P., & Ruan, R. (2017). Cultivation of *Chlorella vulgaris* in a pilot-scale photobioreactor using real centrate wastewater with waste glycerol for improving microalgae biomass production and wastewater nutrients removal. *Bioresource Technology*, 245, 1130–1138.
- Resdi, R., Lim, J. S., Kamyab, H., Lee, C. T., Hashim, H., Mohamad, N., & Ho, W. S. (2016). Review of microalgae growth in palm oil mill effluent for lipid production. *Clean Technologies and Environmental Policy*, 18(8), 2347–2361.

- Rocha, J. M. S., Garcia, J. E. C., & Henriques, M. H. F. (2003). Growth aspects of the marine microalga *Nannochloropsis gaditana*. *Biomolecular Engineering*, 20(4), 237–242.
- Rupani, P. F., Singh, R. P., Ibrahim, M. H., & Esa, N. (2010). Review of current palm oil mill effluent (POME) treatment methods: Vermicomposting as a sustainable practice. *World Applied Science* 11(1), 70–81.
- Sánchez-Bayo, A., Morales, V., Rodríguez, R., Vicente, G., & Bautista, L. F. (2020). Cultivation of microalgae and cyanobacteria: effect of operating conditions on growth and biomass composition. *Molecules*, 25(12), 1–17.
- Sandani, W. P., Nishshanka, G. K. S. H., Premaratne, R. G. M. M., Nanayakkara Wijayasekera, S. C., Ariyadasa, T. U., & Premachandra, J. K. (2020). Comparative assessment of pretreatment strategies for production of microalgae-based biodiesel from locally isolated *Chlorella homosphaera*. *Journal of Bioscience and Bioengineering*, 130(3), 295–305.
- Santhosh, S., Manivannan, N., Ragavendran, C., Mathivanan, N., Natarajan, D., Hemalatha, N., & Dhandapani, R. (2019). Growth optimization, free radical scavenging, and antibacterial potential of *Chlorella* sp. SRD3 extracts against clinical isolates. *Journal of Applied Microbiology*, 127(2), 481–494.
- Santos, M. G. B., Duarte, R. L., Maciel, A. M., Abreu, M., Reis, A., & de Mendonça, H. V. (2020). Microalgae Biomass Production for Biofuels in Brazilian Scenario: A Critical Review. *Bioenergy Research*, 1, 1–20.
- Sayuti, S. C., & Azoddein, A. A. M. (2015). Treatment of palm oil mill effluent (POME) by using electrocoagulation as an alternative method. *Malaysian Journal of Analytical Sciences*, 19(4), 663–668.
- Setthamongkol, P., Tunkijjanukij, S., Satapornvanit, K., & Salaenoi, J. (2015). Growth and nutrients analysis in marine macroalgae. *Kasetsart Journal - Natural Science*, 49(2), 211–218.
- Severes, A., Hegde, S., D'Souza, L., & Hegde, S. (2017). Use of light emitting diodes (LEDs) for enhanced lipid production in micro-algae based biofuels. *Journal of Photochemistry and Photobiology Biology*, 170, 235–240.
- Shah, S. M.U., Ahmad, A., Othman, M. F., & Abdullah, M. A. (2016). Effects of palm oil mill effluent media on cell growth and lipid content of *Nannochloropsis oculata* and *Tetraselmis suecica*. *International Journal of Green Energy*, 13(2), 200–207.
- Shah, Syed M.U., Ahmad, A., Othman, M. F., & Abdullah, M. A. (2014). Enhancement of lipid content in *Isochrysis galbana* and *Pavlova lutheri* using palm oil mill effluent as an alternative medium. *Chemical Engineering Transactions*, 37, 733–738.

- Shin, H. Y., Shim, S. H., Ryu, Y. J., Yang, J. H., Lim, S. M., & Lee, C. G. (2018). Lipid extraction from *Tetraselmis* sp. microalgae for biodiesel production using hexane-based solvent mixtures. *Biotechnology and Bioprocess Engineering*, 23(1), 16–22.
- Silambarasan, S., Logeswari, P., Sivaramakrishnan, R., Incharoensakdi, A., Cornejo, P., Kamaraj, B., & Chi, N. T. L. (2021). Removal of nutrients from domestic wastewater by microalgae coupled to lipid augmentation for biodiesel production and influence of deoiled algal biomass as biofertilizer for *Solanum lycopersicum* cultivation. *Chemosphere*, 268, 1–10.
- Silas, K., Kwaji, H. B., & Gutti, B. (2015). Lipid extraction and transesterification techniques of microalgae – a Review. *International Journal of Recent Research in Physics and Chemical Sciences (IJRRPCS)*, 2(1), 26–37.
- Singh, S. P., & Singh, P. (2015). Effect of temperature and light on the growth of algae species: A review. *Renewable and Sustainable Energy Reviews*, 50, 431–444.
- Sirisuk, P., Ra, C. H., Jeong, G. T., & Kim, S. K. (2018). Effects of wavelength mixing ratio and photoperiod on microalgal biomass and lipid production in a two-phase culture system using LED illumination. *Bioresource Technology*, 253, 175–181.
- Soares, A. T., da Costa, D. C., Silva, B. F., Lopes, R. G., Derner, R. B., & Antoniosi Filho, N. R. (2014). Comparative analysis of the fatty acid composition of microalgae obtained by different oil extraction methods and direct biomass transesterification. *Bioenergy Research*, 7(3), 1035–1044.
- Stracke, M. P., Antonio, V. dos S., Marchetti, M. C., Da, C. B. C., Brondani, M., Cabral Wancura, J. H., Mayer, F. D., & Hoffmann, R. (2019). Thin layer chromatography as innovative technique for qualitative characterization of biodiesel produced by esterification. *Revista Geintec*, 9, 5219–5233.
- Suastes-Rivas, J. K., Hernández-Altamirano, R., Mena-Cervantes, V. Y., Barrios Gómez, E. J., & Chairez, I. (2020). Biodiesel production, through intensification and profitable distribution of fatty acid methyl esters by a microalgae-yeast co-culture, isolated from wastewater as a function of the nutrients' composition of the culture media. *Fuel*, 280, 1–10.
- Suda, S., Atsumi, M., & Miyashita, H. (2002). Taxonomic characterization of a marine *Nannochloropsis* species, *N. oceanica* sp. (Eustigmatophyceae). *Phycologia*, 41(3), 273–279.
- Sun, X. M., Ren, L. J., Zhao, Q. Y., Ji, X. J., & Huang, H. (2018). Microalgae for the production of lipid and carotenoids: A review with focus on stress regulation and adaptation. *Biotechnology for Biofuels*, 11(1), 1–16.
- Suparmaniam, U., Lam, M. K., Uemura, Y., Lim, J. W., Lee, K. T., & Shuit, S. H. (2019). Insights into the microalgae cultivation technology and harvesting process for biofuel production: A review. *Renewable and Sustainable Energy Reviews*, 115,

- Taisir, M., Teo, C. L., Idris, A., & Yusuf, A. M. (2016). Cultivation of *Nannochloropsis* sp. using narrow beam angle light emitting diode in an internally illuminated photobioreactor. *Bioresources and Bioprocessing*, 3(1), 1-8.
- Tan, X. B., Lam, M. K., Uemura, Y., Lim, J. W., Wong, C. Y., & Lee, K. T. (2018). Cultivation of microalgae for biodiesel production: A review on upstream and downstream processing. *Chinese Journal of Chemical Engineering*, 26(1), 17–30.
- Tang, D. Y. Y., Khoo, K. S., Chew, K. W., Tao, Y., Ho, S. H., & Show, P. L. (2020). Potential utilization of bioproducts from microalgae for the quality enhancement of natural products. *Bioresource Technology*, 304, 1–11.
- Teo, C. L., Idris, A., Wahidin, S., & Lai, L. W. (2014). Effect of different light wavelength on the growth of marine microalgae. *Jurnal Teknologi (Sciences and Engineering)*, 67(3), 97–100.
- Umdu, E. S., Tuncer, M., & Seker, E. (2009). Transesterification of *Nannochloropsis oculata* microalga's lipid to biodiesel on Al₂O₃ supported CaO and MgO catalysts. *Bioresource Technology*, 100(11), 2828–2831.
- Uprety, B. K., Venkatesagowda, B., & Rakshit, S. K. (2017). Current prospects on production of microbial lipid and other value-added products using crude glycerol obtained from biodiesel industries. *Bioenergy Research*, 10(4), 1117–1137.
- Vairappan, C. S., & Yen, A. M. (2008). Palm oil mill effluent (POME) cultured marine microalgae as supplementary diet for rotifer culture. *Journal of Applied Phycology*, 20(5), 603–608.
- Vasudevan, S., Arulmoorthy, M. P., & Suresh, R. (2020). Isolation, purification, and structural elucidation of secondary metabolites from *Microcystis aeruginosa* bloom from Muttukadu estuary and its in vitro antibacterial, antioxidant and anticancer potency. *South African Journal of Botany*, 132, 59–67.
- Wahidin, S., Idris, A., & Shaleh, S. R. M. (2013). The influence of light intensity and photoperiod on the growth and lipid content of microalgae *Nannochloropsis* sp. *Bioresource Technology*, 129, 7–11.
- Walne PR. (1979). *Culture of Bivalve Molluscs: 50 years experience at Conwy*. 2th Edition. Londres: The Whitefriars Press Ltd.
- Ward, G. M., Faisan, J. P., Cottier-Cook, E. J., Gachon, C., Hurtado, A. Q., Lim, P. E., Matoju, I., Msuya, F. E., Bass, D., & Brodie, J. (2020). A review of reported seaweed diseases and pests in aquaculture in Asia. *Journal of the World Aquaculture Society*, 51(4), 815–828.

- Wong, Y.-K., Ho, Y.-H., Ho, K. C., & Man, L. H. (2016). Effect of different light sources on algal biomass and lipid production in internal leds-illuminated photobioreactor. *Journal of Marine Biology and Aquaculture*, 2(2), 1–8.
- Wu, J., Alam, M. A., Pan, Y., Huang, D., Wang, Z., & Wang, T. (2017). Enhanced extraction of lipids from microalgae with eco-friendly mixture of methanol and ethyl acetate for biodiesel production. *Journal of the Taiwan Institute of Chemical Engineers*, 71, 323–329.
- Xi, T., Kim, D. G., Roh, S. W., Choi, J. S., & Choi, Y. E. (2016). Enhancement of astaxanthin production using *Haematococcus pluvialis* with novel LED wavelength shift strategy. *Applied Microbiology and Biotechnology*, 100(14), 6231–6238.
- Xiao, M., Shin, H. J., & Dong, Q. (2013). Advances in cultivation and processing techniques for microalgal biodiesel: A review. *Korean Journal of Chemical Engineering*, 30(12), 2119–2126.
- Xu, H., Liu, X., Mei, Z., Lin, J., Aaron, S., & Du, H. (2019). Effects of various light-emitting diode (LED) wavelengths on the growth of *Scenedesmus obliquus* fachb-12 and accumulation of astaxanthin. *Phyton*, 88(3), 335–348.
- Yew, G. Y., Khoo, K. S., Chia, W. Y., Ho, Y. C., Law, C. L., Leong, H. Y., & Show, P. L. (2020). A novel lipids recovery strategy for biofuels generation on microalgae *Chlorella* cultivation with waste molasses. *Journal of Water Process Engineering*, 38, 1-11.
- Yu, Y., Chen, B., & You, W. (2007). Identification of the alga known as *Nannochloropsis* Z-1 isolated from a prawn farm in Hainan, China as *Chlorella*. *World Journal of Microbiology and Biotechnology*, 23(2), 207–210.
- Zahan, K. A., & Kano, M. (2018). Biodiesel production from palm oil, its by-products, and mill effluent: A review. *Energies*, 11(8), 1–25.
- Zainal, A., Yaakob, Z., Takriff, M. S., Rajkumar, R., & Ghani, J. A. (2012). Phycoremediation in anaerobically digested palm oil mill effluent using cyanobacterium, *Spirulina platensis*. *Journal of Biobased Materials and Bioenergy*, 6(6), 704–709.
- Zainal, N. H., Jalani, N. F., Mamat, R., & Astimar, A. A. (2017). A review on the development of palm oil mill effluent (POME) final discharge polishing treatments. *Journal of Oil Palm Research*, 29(4), 528–540.
- Zanella, L., & Vianello, F. (2020). Microalgae of the genus *Nannochloropsis*: Chemical composition and functional implications for human nutrition. *Journal of Functional Foods*, 68, 1-13.
- Zhang, R., Parniakov, O., Grimi, N., Lebovka, N., Marchal, L., & Vorobiev, E. (2019). Emerging techniques for cell disruption and extraction of valuable bio-molecules

of microalgae *Nannochloropsis* sp. *Bioprocess and Biosystems Engineering*, 42(2), 173–186.

Zhao, Y., Yue, C., Geng, S., Ning, D., Ma, T., & Yu, X. (2019). Role of media composition in biomass and astaxanthin production of *Haematococcus pluvialis* under two-stage cultivation. *Bioprocess and Biosystems Engineering*, 42(4), 593–602.

Zhu, L. D., Li, Z. H., & Hiltunen, E. (2016). Strategies for lipid production improvement in microalgae as a biodiesel feedstock. *BioMed Research International*, 1, 1–8.