

Review

Seaweed organic compounds source of hydrocolloids and sustainable food packaging: properties, application, and future direction

Muhammad Qasim Ali¹ · Mohd Akmal Azhar¹ · Mimi Sakinah Abdul Munaim¹ · Nur Fathin Ruslan¹ · Luay M. Alsubhi² · Noormazlinah Ahmad¹ · Abeer Essam Noman³

Received: 30 March 2024 / Accepted: 9 September 2024

Published online: 03 October 2024

© The Author(s) 2024 [OPEN](#)

Abstract

Seaweed has different biologically active macromolecules, including polyphenols, fiber, proteins, and polysaccharides. Recent developments in seaweed bioactive compounds improved food packaging quality and functional properties and increased food production innovations and sustainability. Seaweed compounds are a good source of gelling, thickening, and emulsifying agents in food industrial products. Further Green Extraction methods are used for the extraction of bioactive compounds, these methods are environment friendly, with less time and high-yield production. Seaweeds incorporate antioxidants that reduce lipid oxidation, thus enhancing food's durability and nutritional value and reducing free radicals' occurrence and retard the growth of bacteria. Seaweed has increased its potential for antimicrobial packaging solutions. The manuscript explores the perspective for advancing seaweed-based films, involving property improvements, increased shelf life, and production scalability. Seaweed-based films offer sustainable packaging for fresh produce, seafood, bakeries, and confectionery products. Seaweed-derived bioactive compounds enhance the quality and safety of packaged food products and seaweed polysaccharides in food packaging are their biodegradability and environmental friendliness.

✉ Noormazlinah Ahmad, mazlinah@umpsa.edu.my; ✉ Abeer Essam Noman, abeernoman168@gmail.com | ¹Faculty of Chemical and Process Engineering Technology, University Malaysia Pahang, Al Sultan Abdullah, 26300 Gambang, Pahang, Malaysia. ²Environment Health Laboratory for Food and Water, Jeddah Municipality, Jeddah, Saudi Arabia. ³Department of Food Science and Nutrition, Faculty of Agriculture, Food and Environment, Sana'a University, Sana'a, Yemen.

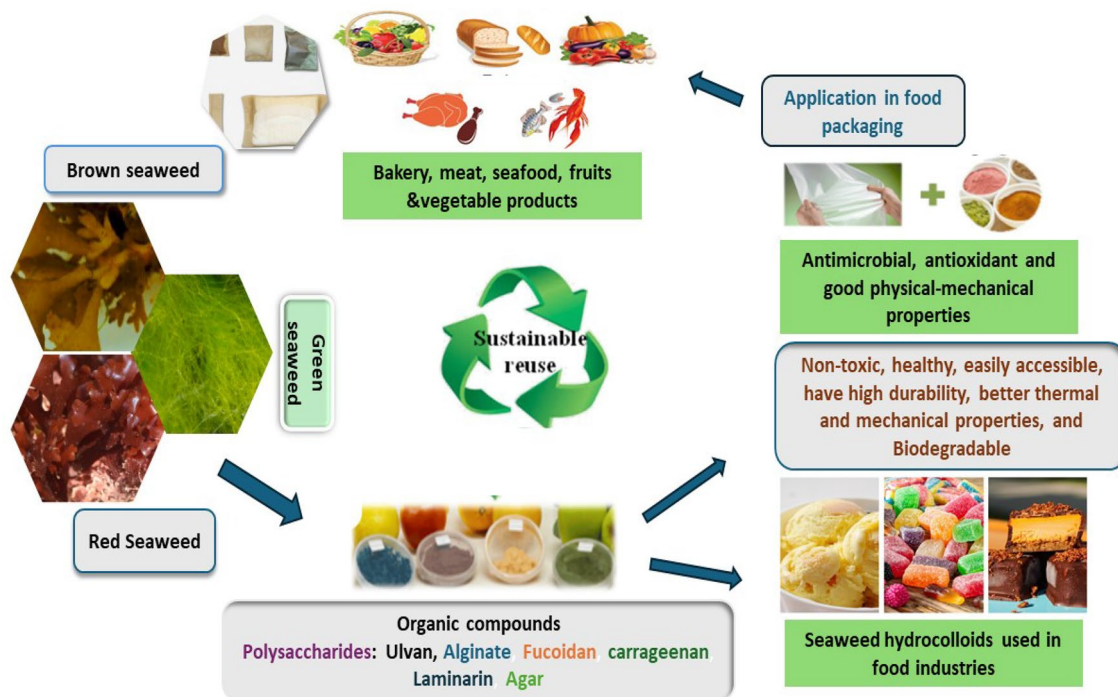


Discover Food

(2024) 4:101

| <https://doi.org/10.1007/s44187-024-00173-w>

Graphical Abstract



Keywords Seaweeds · Bioactive compounds · Polysaccharides · Food packaging · Sustainable packaging

1 Introduction

The demand for sustainable packaging has surged due to environmental concerns like plastic pollution and climate change. The edible films and coatings offer functional and sustainable benefits, protect perishable products from moisture and contamination, and increase their shelf life. This approach addresses the global need for creative packaging solutions that minimize environmental effects [1].

Packaging is a significant component in food industries that helps the essential functions of preserving food and ensuring its overall quality and food safety [2, 3]. The importance of packaging within the broader supply chain framework is not without obstacles. These complications include the potential passage of waste products, cost-effectiveness and energy efficiency considerations, and the imperative of sustainability [4, 5].

Seaweed, rich in polysaccharides and other bioactive compounds, is an efficient source of food packaging materials. Combined with biodegradable polymers, the composite material provides a sustainable, eco-friendly alternative that outclasses packaging materials [6]. Biodegradable polymers, like polyhydroxy butyrate and polylactic acid, are made from renewable resources such as microbial polyesters, proteins, and polysaccharides. Biobased plastics, made from these materials, are particularly useful in food packaging applications like edible coatings and films because they have antimicrobial and antioxidant properties and polysaccharides for packaging that contribute to environmental preservation [2, 7].

Various industrial processes, mainly in the food industry, generate considerable seaweed waste and the significance of reusing these waste materials cannot be emphasized enough, as releasing them into the environment may adversely affect marine ecosystems. Furthermore, this movement underscores algae's immense potential for high-value products as a renewable energy source [8, 9].

Seaweed holds sustainable biomass with numerous possible applications. Research has indicated a high potential for seaweed use in various fields, including the food industry. However, further research is necessary to ensure the successful implementation of this potential [10–12]. Algae's biometabolites have various biological properties like antibacterial,

anti-cancer, anti-fouling, anti-inflammatory, antimutagenic, antiviral, and antibiotic [13, 14]. Recently, marine algae have been identified as a potential source of organic compounds that have advantageous effects on health. The quantity of these beneficial compounds is affected by various factors, such as water temperature, nutrients, and salt content, which control the growth conditions of marine algae [15, 16].

This article explores the use of seaweed polysaccharides in food packaging, highlighting their chemical, mechanical, antioxidant, and antimicrobial properties. Seaweed compounds are a good source of gelling, thickening, and emulsifying agents in food industrial products. The potential for advancements in seaweed-based films and the implications of their use in various food packaging scenarios, including seafood, bakeries, and fresh produce. The review also expects and provides future research pathways in seaweed-based films.

2 Overview of seaweeds

Seaweeds, benthic, macroscopic, and multicellular algae, exhibit greater photosynthesis efficiency than their terrestrial counterparts. Consequently, they grow faster, resulting in rapid biomass accumulation. Seaweed constitutes a significant proportion of marine biomass, contributing to approximately 50% of the earth's primary production. Their versatile applications, which include food and feed production, have made them extensively utilized [17]. The considerable abundance of seaweed has increased demand, leading to the widespread cultivation of this marine resource over the past decade. Seaweed farming and production processes have experienced significant expansion, playing a role in the biotechnological and fishing industries of nations [18]

Seaweed represents a critical living resource for marine biodiversity, with more than 10,000 species worldwide growing on firm substrates, including rocks at depths of up to 180 m. These species were classified into three groups: red (Rhodophyta), brown (Ochrophyta), and green (Chlorophyta) [19]. Seaweeds constitute an outstanding source of diverse bioactive compounds in several fields, including pharmaceuticals and agriculture. Different types of seaweed, red, green, and brown images are displayed in Fig. 1.

Red seaweed stands out from its unique red and blue pigments, namely phycoerythrin, phycocyanin, and chlorophyll a. The variety of red seaweed species is due to their diverse plant forms and life cycles. Prominent examples of red seaweed species are *Poryphyra capensis*, *Notogenia stiriata*, and *Aeodes orbitosa* [21].

Brown seaweed comprises many polysaccharides, including fucoidans, alginate, and laminarin. Brown seaweed exhibits various sizes, species, and overall morphologies. They are generally characterized by their brown colour, which is

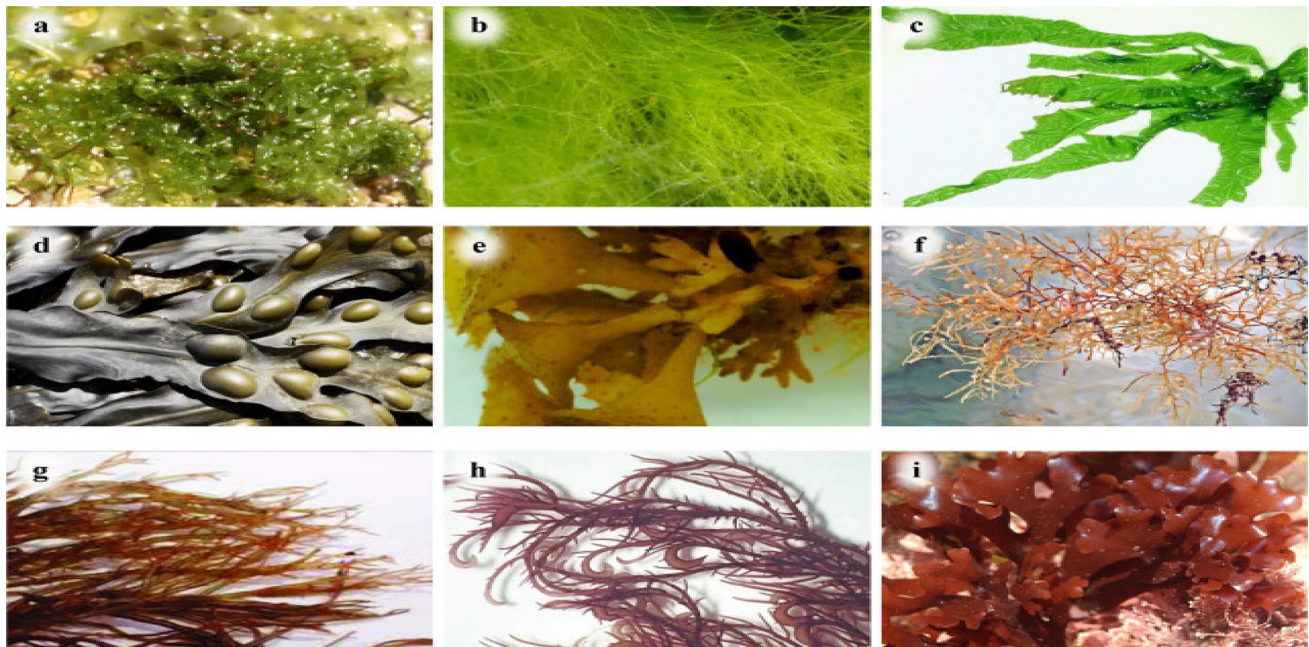


Fig. 1 Seaweeds as green, brown, and red. **a** *Ulva reticulata* Forsskål; **b** *Chaetomorpha linum* Kützinger; **c** *Ulva lactuca* f. *fasciata*; **d** *Fucus vesiculosus* Linnaeus; **e** *Turbinaria turbinata* Kuntze; **f** *Sargassum natans* Gaillon; **g** *Gracilaria edulis*; **h** *Hypnea musciformis*; **i** *Chondrus crispus* [20]

attributed to the occurrence of a photosynthetic pigment called fucoxanthin. Kelp, including species such as *Laminaria pallida*, *fucus*, and *zonaria*, is a widespread type of brown seaweed [22].

Green algae are common in fresh and saltwater environments, and their green colour is due to chlorophyll a and b [23, 24]. Common green seaweed includes the genera *Monostroma*, *Cladophora*, *Ulva*, and *Chaetomorpha* species. These are commonly encountered in marine and brackish water environments. *Chaetomorpha* and *Cladophora* are filamentous green algae often found in intertidal areas, attached to rocks, or floating freely in the water [24, 25].

3 Extraction methods for bioactive components

Seaweed has been an excellent source of diverse bioactive compounds in several fields of food and pharmaceutical industries. For extraction of bioactive compounds from seaweeds in Conventional Extraction such as Solid–Liquid Extraction (SLE), this extraction uses organic solvent with a specified extraction time. The further efficient methods are Green Extraction methods, these methods are environment-friendly with high-yield productions. Green extraction processes such as Ultrasound-assisted extraction (UAE), Microwave-Assisted Extraction (MAE), Supercritical Fluid Extraction (SFE), and Reactive Extrusion Using a small amount of solvent under specific conditions with a fast extraction [26].

3.1 Ultrasound-assisted extraction (UAE)

Ultrasound-assisted extraction is more efficient than conventional extraction and some other green extractions. Ultrasound allows greater solvent penetration into the sample by increasing the contact surface between the solid and liquid phases [27]. Uses ultrasound waves with a frequency between (20 and 100) kHz which causes bubbles due to the created pressure difference. Then, these bubbles collapse and undergo cavitation, which causes the breakdown of the liquid–solid interface near the particles, releasing bioactive compounds into the substance [28].

The ability of ultrasound to stimulate cavitation depends on several factors including ultrasonic frequency and intensity, properties of the medium such as surface tension and viscosity and ambient conditions including temperature and pressure [29]. Ultrasound decreases extraction time, solvent used and processing costs [27]. Parameters that are optimized during this method are frequency, power, temperature, time and sample and solvent ratio [30]. The application of UAE in producing bioactive compounds from seaweed extraction such as phenolic compounds. Some main advantages of extracting phenolic compounds from seaweed using the UAE method are low temperature, short time and a small quantity of solvent [31].

Polysaccharide compounds in the form of *laminaria* obtained by the UAE method show better biological activity, and antioxidant and antimicrobial activity and preserve better antioxidant properties [32]. Another study shows that a high percentage of fucoxanthin and phenolic compounds from (*Padina tetrastromatica*) macroalga [33]. Using UAE for extraction of fucoxanthin from *Sargassum echinocarpum* gets a high yield (2.8–3.9%) and high antioxidant activity between (44.6–98.8 ppm) mostly depending upon the extraction conditions [34]. The research shows that UAE extraction methods are efficient for organic compounds. Seaweed (*Sargassum carpophyllum*) extract has a good total phenolic compound (46.22–65.00 mg) phloroglucinol, ethanol concentration is 50% and ultrasonic power was 200 watts [35].

3.2 Microwave-assisted extraction (MAE)

MAE is an alternative technology that provides an environmentally and economically advantageous option. It allows the production of affordable and high-quality compounds that meet “green” environmental criteria. This method is efficient due to the reduced processing time and amount of solvent used [36]. This technique integrates microwave and conventional solvent extraction, utilizing ionic conduction and dipole rotation to directly affect and occur simultaneously with the molecules [37]. Microwave heating induces energy absorption by polar molecules without any heat loss to the surroundings, while simultaneously disrupting cells. Damaged cells enhance the speed of mass transfer and diffusion out of a solid when the processes of mass and heat transfer work together and in the same direction [38]. Microwave extraction involves using microwave power to heat samples and solvents that are contained in high-quality vessels, with the temperature being carefully regulated [39].

The use of microwave-assisted pressurized hot water extraction (MAPHWE) technology for solubilizing seaweed components, specifically at an initial stage, has been explored in various studies. This method is recognized for its efficiency in extracting valuable compounds from seaweed, such as polysaccharides, phenolic compounds, and other bioactive

substances while maintaining a green and sustainable approach [40]. A study shows that MAPHWE was applied to *Rugulopteryx okamurae* at 180 °C for 10 min with a liquid-to-solid ratio of 30, resulting in the dissolution of over the (40%) initial material. The alginate recovery yield was (3.2%) and the phenolic content of the water-soluble extracts was (2.3%). When distilled water was used as the solvent both were slightly higher. Interestingly, the carbohydrate content in the extract remained consistent at 60% regardless of the solvent used, while the sulphate concentration was higher in samples processed with salt water from the same coastline as the seaweed [41].

Ulva prolifera, a green alga, uses MAE technology to study polysaccharide characteristics and bioactivities. Temperature and acid concentration increased sulphur and decreased molecular weight. The extraction of polysaccharides at 90 °C with 0.05 M HCl is noted for its superior water-holding capacity (41.32 g/g) and oil-holding capacity (15.09 g/g). This suggests that the extraction conditions, including temperature and pH, significantly influence these properties. Regarding the foaming properties, the extraction at 150 °C with 0.05 M HCl is highlighted for its optimal foaming capacity (143% and 113%) and higher antioxidant activity [42]. A study shows that *Carpophyllum flexuosum*, *Carpophyllum plumosum*, and *Ecklonia radiata* brown seaweeds have strong antioxidant activity (62.1 mg gallic acid) and radical scavenging ability [43].

Figure 2 indicates that four kinds of brown algae, namely *Ascophyllum nodosum*, *Fucus vesiculosus*, *Laminaria digitata*, and *Saccharina latissima*, were utilized to extract organic compounds using three different pretreatment methods. Among these methods, swelling had the most stated impact on improving the production of bioactive compounds. By incorporating the UAE and MAE techniques in the processing of seaweed, the production of bioactive substances such as pigments, mannitol, and polyphenols can be increased. This has the potential to improve their abilities to combat free radicals and iron chelating capacity [44].

In this study most suitable conditions for solubilizing *Ascophyllum nodosum* seaweed biomass using microwave-assisted extraction were temperature (120 °C) and a solid-to-liquid ratio (1.03 w: v), a processing time (15 min). which maximized solubilization efficacy while minimizing the applied energy per mass of solubilized seaweed. Antimicrobial tests conducted on *S. aureus* and *E. coli* showed up to 97% inhibition of bacterial growth after 8 h, indicating the presence of antimicrobial characteristics in the extracts obtained through the dedicated microwave extraction of *Ascophyllum nodosum* [45].

3.3 Supercritical fluid extraction (SFE)

Applied generally at both laboratory and industrial levels, SFE is a green extraction process for important non-polar or mid-polar substances such as lipids, essential oils, and carotenoids [46]. Using solvents in the supercritical conditions of this method; so, temperature and pressure are elevated over their critical point. Under those conditions, supercritical

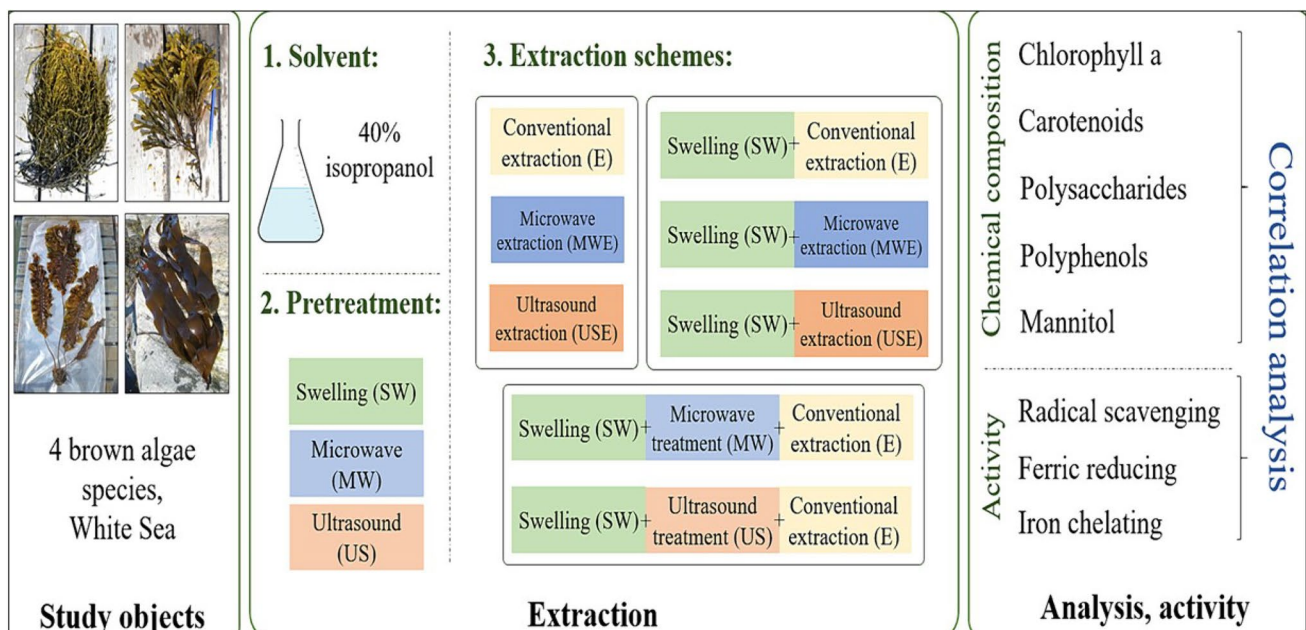


Fig. 2 shows the effects of extraction methods on chemical composition and bioactivity [44]

fluids often acquire higher density but keep similar viscosities and intermediate diffusivities to gas [47]. Carbon dioxide is the recommended solvent for the extraction of bioactive substances from natural sources since other fluids can be employed at supercritical conditions. Among its various benefits are mild critical conditions associated with carbon dioxide. It is cheap, safe, environmentally friendly, non-toxic, non-flammable, non-explosive, easily available [48].

Conditions during the extraction, especially pressure and temperature, influence the solubility and selectivity of the many chemicals in the supercritical fluid [47]. low critical temperature (31.1 °C) and pressure (73.8 bar), carbon dioxide is a great solvent for extracting heat-sensitive bioactive chemicals so preserving them, and no degradative changes can occur [47]. In addition to temperature, pressure, supercritical solvent, proportion, and type of co-solvent, various other factors are associated with and linked to the sample and the extraction process. These factors include water content, particle size, dispersant agent, sample amount, SCF flow rate, mode of extraction, extraction time, and fractionation [49]. The interactions of various components that enhance a specific process are complicated. Therefore, obtaining appropriate extraction conditions is important for the successful isolation of bioactive compounds [30].

The study examined the application of a design of experiment to assess supercritical fluid extraction (SFE) as a sustainable extraction technology. The approach utilized a mixture of CO₂, ethanol, and water. The temperature was regularly varied between 40 and 80 °C while keeping the (300 bar) constant pressure [50]. *Alaria esculenta*, *Laminaria digitata*, and *Ascophyllum nodosum* seaweed were used for analysing organic compounds. The highest level of selectivity was achieved while employing CO₂ with a concentration of only 5 vol% ethanol and no water. This facilitated a significant retrieval of β-carotene and yielded an extract devoid of carbohydrates, proteins, and toxic metals such as arsenic. The most effective methods for extracting the highest amount of the remaining target analytes showed considerable diversity. The analytes with the highest water content, including fucoxanthin and phloroglucinol, also had the lowest relative selectivity [50].

3.4 Advantages and limitations of green extraction methods

All extraction methods, conventional or green have advantages and disadvantages. An extraction process selection must consider their, rate of recovery, cost, time, solvent volume, and scale-up. Under suitable conditions, water extraction is the easiest approach for polysaccharide extraction from plants, microbes, and algae. However, high heat and extensive processing times may degrade polysaccharides and important bioactive chemicals [51]. In contrast, green extraction methods can successfully extract polysaccharides with short treatment time, low solvent consumption, high extraction yields, and other enriched features [52].

Microwave-assisted extraction (MAE) has a brief duration of treatment. Both organic solvents and water are viable options for application. Appropriate for heat-sensitive chemicals, and superior to the traditional Soxhlet process. Only solvents possessing a high dielectric constant and a low dissipation factor are suitable for application. The MAE closed vessel has a significant risk of explosion, particularly its high capital cost [39]. Ultrasound-assisted extraction (UAE) is characterised by a brief treatment duration and reduced solvent usage. Optimal extraction efficiency, Affordable. The effectiveness of extraction is contingent upon the composition of the plant matrix. Preferable solvents are those that have low surface tension, viscosity, and vapour pressure. Overusing sonication can potentially impair the quality of extracts [53].

Supercritical Fluid Extraction (SFE) is a form of green extraction method. Extracts obtained with high levels of purity and without any remaining contaminants. Extracts obtained without the use of solvents and with a minimal extraction duration are an ideal method for extracting thermolabile compounds, however, it does have some drawbacks. Expensive equipment is required for high-pressure applications. Extracting polar molecules may provide challenges. The processing cost and energy usage are high [54]. The green extraction technique is costly because of its high processing expenses and energy consumption. However, it offers efficient extraction with a good yield and requires less time [54–57].

4 Bioactive compounds in seaweed

As living marine resources, seaweed contains abundant macro- and micro-nutrients, including carbohydrates, proteins, fiber, vitamins, and minerals. They serve as natural sources of macro elements, including sodium, calcium, potassium, magnesium, Sulphur, chlorine, and phosphorus, as well as micro aspects like iodine, zinc, copper, selenium, nickel, cobalt, boron, and manganese. Seaweeds contain substantial amounts of iodine, which is crucial in preventing goiter disease

in humans [58, 59]. Red and green seaweed generally exhibits a high protein content, up to 30%, as compared to brown seaweed. However, green seaweeds are typically rich in carbohydrates compared to red and brown seaweed [60].

The green seaweed *Ulva lactuca* and *Enteromorpha intestinalis* have the maximum amount of carbohydrates, i.e., 36.01% and 31.08% [61, 62], and brown seaweed (*Dictyota dichotoma*) contains minimum carbohydrate content, i.e., 09.95% [63]. The fiber content of seaweeds ranges from 35 to 60% of dry mass, higher than other plants [10, 58]. The lipid content of seaweed generally ranges from 5.1% in *Ulva clathrata* to 1.29% in *Enteromorpha intestinalis*. However, studies reported that lipid content in *Utricularia rigida* is 13% and *Kappaphycus alvarezii*, 1.07%, respectively [64, 65].

Various commercially available seaweed species, namely *Undaria pinnatifida*, *Saccharina angustata*, *Pyropia tenera* and *Sargassum fusiforme* along with nine non-commercial seaweed species, specifically *Ecklonia radiata*, *Cystophora polycystidea*, *Hormosira banksii*, *Codium galeatum*, *Durvillaea potatorum*, *Cystophora torulosa*, *Phyllotricha decipiens*, *Laurencia filiformis*, *Phyllospora comosa*, were examined by a group of researchers to determine their functional properties. The protein, total lipid, crude fiber, and ash content of commercially available seaweeds ranged from 32.76 to 350 mg/g, 7.24 to 31.5 mg/g, 28.85 to 46.24 mg/g, and 120.3 to 297.45 mg/g, respectively. In difference, the varieties of non-commercial seaweeds were found to be 31.5 to 157 mg/g, 4.57 to 102.05 mg/g, 31.32 to 219.96 mg/g, and 54.56 to 196.06 mg/g, respectively [66–69]. In addition, seaweeds are a rich source of Fat and water-soluble vitamins, phenolic compounds and essential fatty acids (ω -6 and ω -3 fatty acids), as reported in this study [70]. The bioactivity of the substances present in seaweeds is given in Table 1.

5 Seaweeds polysaccharides

Polysaccharides have different structures and properties, and it is a sulfated (e.g., fucoidans, carrageenans, Galatians, and agars) or non-sulfated (e.g., alginates and laminarin) [79, 80]. These polysaccharides are important in food packaging materials and increase their physical and functional properties. Table 2 shows different associated polysaccharides and their chemical structure. The main types of seaweed and their polysaccharides compounds are presented in Fig. 3.

5.1 Agar and Carrageenan

Sulfated galactans, including carrageenans and agar, are present in seaweeds and extensively utilized as biopolymers in the food industry [60]. Agar and carrageenan are digestible oligosaccharides that are harmless to human teeth and are linked with various beneficial properties, such as prebiotic effects, anti-tumor, antioxidant, and immune-modulating effects [19, 93].

Agar is comprised of agarose and agaropectin, which have structures and functions comparable to carrageenan. Its ability to gel, emulsify, and thicken makes it a valuable substance not just in scientific research but also in different commercial applications, including food and medical productions [59, 60, 94].

Carrageenan solubility is affected through its hydrophilic and anionic properties, with greater sulphate ester levels resulting in higher solubility. Carrageenan is categorized into three classes based on its degree of sulphation, with the Lambda type having the highest sulphation level at 40% (w/w). Lambda carrageenan does not form gels but exhibits thickening properties. Iota carrageenan is less sulphated than Lambda, enabling it to form gels when combined with calcium ions. The kappa family has the lowest possible level of sulphate ester at 20% (w/w) [94]. When exposed to potassium ions, these types of carrageenan form a robust gel and are utilized to produce transparent, cohesive films [94, 95].

Seaweed-derived biopolymers have excellent film-forming capabilities and mechanical and barrier characteristics. Agar generates gel, due to its distinctive molecular structure. The film-forming properties of agar are influenced by factors such as concentration, temperature, and the presence of additional ingredients. Agar films are utilized in food packaging due to their non-toxic properties, a barrier against oxygen and moisture, and their capacity to preserve food quality [96, 97].

Carrageenan, a sulfated polysaccharide, used in the production of gels and films due to its exceptional mechanical properties and flexibility. The addition of carrageenan to other chemicals improves the film's tensile strength and elongation at the point of rupture, hence boosting its appropriateness for a variety of uses [98]. The molecular weight and quantity of sulfate have a major impact on the film-forming properties of carrageenan. Usually, solid films with better

Table 1 The bioactivity of the substances present in seaweeds

Bioactive compounds	Components	Benefits	References
Protein	Have different essential amino acid	Anti-inflammatory capabilities, as well as antimicrobial and antiviral activities, Communication between cells, antioxidative properties	[71]
Fat	Polyunsaturated fatty acids, ω -3- ω -6 fatty acids	Activity promoting health, antimicrobial and antibiotic capabilities, membrane flexibility, oxygen and electron transportation, adaptation to heat	[72]
Polyphenols	Isoflavones, lignans, benzoic Acid, Flavonoids, cinnamic Acid, phenolic acids, Quercetin	Anti-photoaging, anticancer activity, anti-viral, antiobesity, antioxidants, antimicrobial, host defence, antiallergic	[73]
Polysaccharide	Galactans, fucooidan, laminarin, alginates	Good Antioxidants activity, soluble dietary fiber, anti-microbial activity, antitumor and anti-inflammatory	[74]
Minerals	Entirely macro and microelements	Development and health-improving for growth and prevention of goitre disease	[75]
Sterols	Brassica sterol, desmosterol, sitosterol, uco-cholesterol	Decrease cholesterol level of blood serum in humans	[76]
Pigments	Chlorophylls and carotenoids, fucoxanthin and phycobiliproteins	Rich in antioxidants, anti-cancer, anti-angiogenic, exhibiting neuroprotective, anti-obesity, and anti-microbial properties	[77, 78]

Table 2 Chemical structures of seaweed polysaccharides

Polysaccharide	Chemical structure	References
Alginate		[81–83]
Fucoidan		[63, 84, 85]
Laminarin		[84, 86, 87]
Agar		[81, 88, 89]
Kappa-Carrageenan		[81, 90, 91]
Ulvan		[23–25]

barrier properties follow from higher molecular weight and increased sulfate. Carrageenan films show great thermal stability, which makes them quite helpful for uses requiring resistance to strong heat [99, 100].

5.2 Ulvan

Common green seaweed includes the genera *Monostroma*, *Cladophora*, *Ulva*, and *Chaetomorpha*. Ulvan is a sulfated polysaccharide, a primary cell wall component in some green seaweed species, particularly in the *Ulva* genus, and its applications in the agriculture, pharmaceutical, and food industries. [23–25]. Ulvan comprises α - and β -(1,4)-linked monosaccharides, including rhamnose, xylose, glucuronic acid, and iduronic acid. These monosaccharides combine to form characteristic repeating disaccharide units in Ulvan, known as aldoburonic acid, combined with rhamnose and glucuronic acid or iduronic acid [101, 102].

The properties of Ulvan, like those of other seaweed polysaccharides, are heavily influenced by factors such as eco-physiology, the specific species it is derived from, and the methods used for extraction [61]. Ulvan gels exhibit

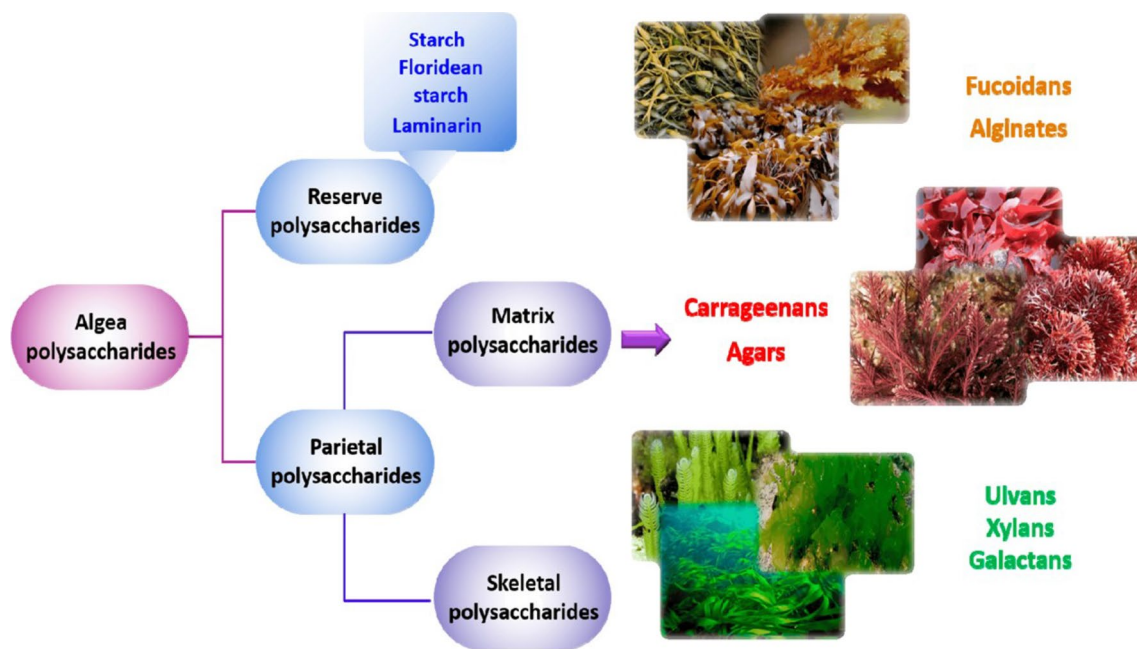


Fig. 3 The types of seaweeds and their polysaccharide compounds [92]

thermo-reversible properties, meaning they can undergo gelation and solation reversibly upon temperature changes. The conformation of Ulvan, and consequently its gel formation, is influenced by factors such as ion concentration and pH variations, with high or low ion concentrations and pH levels playing a role [103].

Furthermore, the bioactivity of Ulvan is also affected by its molecular mass, emphasizing the importance of molecular size in determining its biological effects [11]. Lower molecular weight polysaccharides have demonstrated more significant antioxidant activity and antilipidemic effect on triglycerides and HDL-cholesterol. Additionally, polysaccharides have shown inhibitory effects on various viruses, although the specific outcomes depend on the controlled dosage and the kind of virus [11, 61, 101].

Ulvan-based films for Biodegradable packaging and wound dressings are suitable with desired mechanical and barrier properties, good mechanical strength, and flexibility. Ulvan has a Sulphate group that improves its capacity for film stability by encouraging intermolecular interactions, which are fundamental for preserving film integrity under various environments [104]. Ulvan-based films prevent moisture and gas exchange and extend the shelf life of perishable goods. The study also shows that Ulvan-based films are biodegradable and environment friendly, increasing demand for sustainable packaging solutions [103]. The potential of Ulvan-based films in biomedical applications, such as wound dressings. Due to biocompatibility and its ability to form films providing a protective barrier that supports healing. Ulvan-based wound dressing enhances its suitability for wound care by reducing the risk of infection [105].

5.3 Fucoidans, alginate, and laminarin

Brown seaweed comprises many polysaccharides, including fucoidans, alginate and laminarin. These polysaccharides are essential components of brown seaweed cell walls and have various biological activities and potential applications [10]. Alginates constitute the cellular walls and intracellular matrix found in brown seaweed. They are composed of exchanging blocks of β -D-mannuronic acid and α -L-guluronic acid [106].

Alginates possess diverse properties stemming from their unique monomer sequence. This versatility allows them to serve as a multifunctional material, finding applications as films through interaction with di and trivalent cations, as well as decaying agents in tablets by their water-swelling capabilities [106–108].

The primary polysaccharide for glucose storage in brown seaweed is known as laminarin. This compound is associated with assorted bioactivities, such as antitumor and antioxidant properties [84, 108, 109]. Its potential applications in various fields, the primary utilization of which is in the industrial sector, are specifically related to its role as a ligand. A ligand is a molecule that binds to a receptor, and in this context, it binds to pattern-recognition receptors within the innate immune system [58, 83, 110].

Fucoidan polysaccharides refer to complex sulphated polysaccharides extracted from brown seaweed. These polysaccharides consist of L-fucose and sulphate ester groups and other components such as monosaccharides, proteins, and acetyl. The characteristics and composition of Fucoidan polysaccharides vary depending on conditional factors such as season, geographical origin, extraction methods, and species. Due to their advantageous properties, Fucoidan polysaccharides are extensively utilized in biomedical applications and related fields [58, 110]. Additionally, Brown seaweed contains various other components, including terpenes, halogenated compounds, lipids, sterols, pigments, alkaloids, phenolic compounds [108, 111, 112].

6 Hydrocolloid substances in seaweeds

Seaweed produces a variety of biologically active macromolecules, including polyphenols, diterpenes, and polysaccharides, which exhibit diverse structural and physicochemical properties and serve fascinating functions. Hydrocolloids are commonly utilized due to their inherent physical properties, serving to stabilize emulsions, exhibit viscous characteristics, promote gelation, maintain suspensions and foams, and regulate crystal growth [92].

The composition of hydrocolloids in seaweeds is subject to many biological, physical, and environmental influences. These factors, including yield timing, variety differences, and extraction methods, can intensely affect the functional characteristics of the polysaccharides [113–115]. The preparation method mainly influences the viscosity, with elevated temperatures posing a particularly detrimental effect. Moreover, it is critical to maintain a pH level ranging from 6 to 7 [115]. Seaweeds offer various hydrocolloids utilized by the food and pharmaceutical sectors [116, 117].

Most hydrocolloids (agar, alginates, and carrageenan) in seaweeds are predominantly situated within the cellular structure known as the cell wall [116, 118]. Extraction of this hydrocolloid from the red algae genera (*Gelidium* and *Gracilaria*), [119], is common in Chile, India, Japan, Europe, the Philippines, and the southern United States [115, 117]. Around 25% and 17% of agar are stored in the cell walls of *Gelidium* and *Gracilaria* [115, 117]. Additional genera, such as *Pterocladia*, primarily encountered in (Portugal and New Zealand) and *Gelidiella*, found in (Egypt and India) as valuable agar resources [115, 118, 120].

The primary utilization of agar stems from its remarkable attributes in thickening and gelling. It can retain substantial quantities of soluble solids, including sugars, and elevated melting temperatures and its proficiency in preventing sugar crystallization, agar is widely desirable in the food industry, particularly for the preparation of icings and bakery glazes [81, 88].

The formation of a low gel strength matrix by agar is a distinctive property that renders it highly versatile for various applications. It finds practical utility in multiple domains, including liquid and spreadable foods and soft-textured confectionery [121, 122]. Agar serves multiple functions beyond its primary applications. It can be employed as a substitute for fats, as a cryoprotectant that mitigates damage during freezing and thawing processes, and as a material for producing edible films [123–125].

Alginates, alginic acid, or algin are the most synthesized algal polysaccharides. The compounds are obtained from the cellular walls of brown algae [126, 127]. The main constituents of alginate are 1,4- β -D-mannuronic acid and α -L-guluronic acid, which comprises a linear polymer. The composition of *A. nodosum* includes 60% mannuronic acid and 40% guluronic acid [128–130].

Alginates can produce cross-linked gels that do not undertake melting, in contrast to agar. Therefore, they are utilized to produce restructured meat, vegetables, and baked products. Alginate is used independently or in combination with other hydrocolloids in the production of frozen desserts and reduced-fat products to enhance the stability of mixtures, elevate viscosity, prolong melting time, and improve organoleptic characteristics [123, 127, 129].

According to research, alginate exhibits potential as an appetite regulator and may have utility as a dietary supplement. Nevertheless, the integration of the substance mentioned above into breakfast bars does not show significant variances as a means of reducing appetite in comparison to the control group [81, 114, 124, 129]. The efficacy of alginate as a coating film in cooked chicken nuggets was observed, resulting in enhanced heat distribution and reduced cooking duration [82, 131, 132].

Applying sodium alginate as a coating agent on bream yielded promising outcomes, including enhanced antioxidant capacity. This attribute is believed to inhibit fish spoilage and prolong its shelf-life effectively [133]. Alginate is a coating agent that enhances sensory characteristics and decreases water loss in food products. It has antimicrobial agents and anti-browning agents, such as ascorbic acid and citric acid, to maintain the colour of freshly cut Kent mangoes and enhance their antioxidant capability [74, 115, 127, 134].

The carrageenans are characterized by the number and position of sulphate groups on the repeating galactose units. Carrageenan's capacity to establish linkages with milk proteins, yet when present in low concentrations of 0.01%, represents a highly advantageous characteristic [135–137].

Carrageenans and semi-refined carrageenans are utilized in the food industry because they gel, emulsify, thicken, and stabilize. Carrageenans are frequently utilized as a constituent in dairy commodities, particularly in frozen desserts like ice cream [58, 81, 90] and milk-based products [74, 115]. According to the findings of some other studies, the shelf life of fresh chicken breasts increased by utilizing carrageenan as an encompass film or in bakery products like bread [138]. Table 3. shows some physicochemical and functional properties and applications in different industries and food and beverage products [139]

7 Properties of seaweed-based films

Seaweed is commonly utilized in packing, culinary, and biomedical applications because they have biocompatibility, absorbability, and biodegradability. Hydrocolloid polysaccharides are derived from seaweeds and utilized as biopolymeric films [149]. These compounds have antimicrobial properties that prevent food packaging from microbial spoilage. Seaweed films are flexible and can be modified by merging with adding additives and other biopolymers. This flexibility allows the film's modification for specific needs like strength, permeability, and moisture resistance. Overall, seaweed films provide an eco-friendly alternative to traditional packaging materials [150, 151]. Their biodegradability, flexibility, and versatility make them a favorable solution for businesses seeking to reduce plastic waste and moderate its harmful impact on the environment [152].

The toxicity of chemicals in food packaging can be relieved by using natural compounds like seaweed, which contains organic compounds. Seaweed, with its high organic content, can be an effective active agent or raw material. When combined with biodegradable polymers, these materials offer enhanced sustainability, functionality, and sensory properties. Seaweeds produced environmentally friendly and biodegradable packaging materials, as shown in Fig. 4 [74].

Seaweed-based films have biodegradability, and distinct visual characteristics, particularly transparency are essential for applications in food packaging and coating. Agar and carrageenan are derived films due to their gel strength, low turbidity, high transparency and good appearance [153, 154]. The molecular structure of these polysaccharides allows for the formation of films with minimal light scattering, enhancing transparency. Casting and extrusion techniques can influence the film's microstructure, affecting its optical clarity the film's thickness and uniformity, which are crucial for achieving high transparency [155, 156].

During film formation, drying conditions such as temperature and humidity, can impact the film's surface smoothness and transparency [157]. Additives and plasticizers are incorporated into seaweed-based films to enhance their mechanical properties and flexibility and affect appearance [158]. A smooth surface is essential for achieving high transparency and is achieved through control of the film-forming process and appropriate additives [159]. The colour of the film can be modified by specific seaweed or natural pigments, which can enhance the visual of the packaging material and maintain transparency [160].

7.1 Mechanical properties

At present, diverse potential implementations exist for packaging that incorporates seaweed and other composite materials [161]. Comprising chitosan and κ -carrageenan (κ -CG) determined the mechanical and structural properties of the blend film. The results indicate that integrating kappa-carrageenan into the chitosan film improved its flexibility and achieved a uniform and smooth surface. Furthermore, it was found that incorporating κ -CG reduced water solubility and extension at break. So increased moisture affinity, enhanced tensile strength, and improved hydrophobicity [162] Another investigation was of a film composed of kappa-carrageenan and chitosan combined with allyl isothiocyanate. The polysaccharide film with opposite charges displayed favorable coating characteristics and improved gas barrier performance. Moreover, it delayed the release of bioactive substances incorporated into the film. The Chitosan/ kappa-carrageenan complex film demonstrated enhanced antimicrobial effectiveness against *Bacillus subtilis* and *Bacillus cereus*, as supported by the results [74]

A nanocomposite film was formed using polylactic acid and laminated agar/ kappa-carrageenan /clay bio-nanocomposite. The film showed improved properties, such as increased tensile strength, water vapour permeability, water uptake ratio, water solubility, and enhanced thermal stability. kappa-carrageenan and polydextrose matrix plays an essential

Table 3 Functional properties of hydrocolloids and their applications

Seaweeds hydrocolloids	Functional and Physicochemical properties	industries	Food & beverages	References
Carrageenan	Thermo-reversible, good gelling (Gell Strength: 100–350 g/cm ² , Gelling Point: 30–50 °C, Melting Point: 50–70 °C, Appearance: yellowish powder) thickening (Viscosity: 30–300 cP) stabilizing, emulsifying, and water holding properties	Thickenings for toothpaste, eye drops, beads for the controlled released system, cosmetics, suspending agents in antacids, lotion and creams, suppositories, Cosmetic things, and the paint industry	Puddings, milkshakes, canned food, ice cream, desserts, vegan alternatives to gelatine, processed meat, cream thickener, chocolate milk, fruit juices, beer, jam, pet food, sauces, and gravies,	[139–142]
Alginate	Fast absorption of water (Soluble in water and insoluble in ethanol and ether), Excellent gelling, stabilizing, thickening agent (viscosity is high and Appearance is yellowish powder), Thermo-irreversible (with the presence of calcium, Ca ²⁺), melting point is > 300 °C	Dental impression, filling agents, matrices to drug delivery systems, additives in dehydrated products, wound dressing, paper and textile industry, medicine for rheumatoid arthritis, emulsifiers for paint and plastic industry, antacids, prosthetic devices, Heavy metal absorption and purifier for wastewater, material for artificial fiber, biodegradable sutures, lubricant, refining	Thickeners in drinks, cold prepared bakery cream, pie and pastry filling, ice cream, jelly, restructured foods, dessert gels, instant pudding,	[139, 141, 142]
Agar	Thermo-reversible, Excellent gelling agent (gelling point: 32–45 °C, melting point: 85–95 °C, viscosity: 10–100 cP, Appearance: yellowish powder), high temperature resistant	Thickeners for toothpaste, beads for the controlled released system, Inhibitor of Papilloma, suppositories, Cosmetic and paint industry, dengue and herpes virus, cosmetics, lotion, and cream, suspending agents in antacids, and eye drops	Puddings, milkshakes, desserts, cream thickener, chocolate milk, vegan alternatives to gelatines, tofu, ice cream, processed meat, canned food, beer, sauces and gravies, fruit juices, jam, pet foods	[141–144]
Ulvan	Structural, complexity, and bioactivity viscosity is 10–100 cp, slightly granular, appearance: powder is pale yellow to light green	Used in cosmetics, biomaterials, fibres, scaffolds, pharmaceutical gels, fuel, antibacterial, and drug delivery systems	Food packaging, food preservation, beverages, juices, healthy drinks, food and feed products	[145–148]

Seaweed	Characteristics	Applications
Red Seaweed <i>Poryphyra capensis</i> <i>Aeodes orbitosa</i> Agar Carrageen 	Sustainable Source Eco-friendly and Biodegradable Reduces carbon emission	 → Sustainable Packaging  → Bioactive Plastic
	Active Functions Antioxidant Antimicrobial agent	
Green Seaweed <i>Ulva</i> <i>Monostroma</i> <i>Cladophora rupestris</i> <i>Codium tomentosum</i> 	Shelf Life Enhanced low water content Increased stability	 → Active Packaging  → Edible Packaging
	Health Nutraceutical Nutritious	
Brown Seaweed <i>Laminaria</i> Kelp Fucus, <i>Sargassum muticum</i> 	Market Value Colourful Flavourful	 → Sachet Packaging
	Plasticizers Cross-linking agent	

Fig. 4 Different types of Seaweed and in their applications in food packaging [74]

role in the stability and quality control, enabling the diffusional mobility of α -lionic acid. An electrostatic attraction was observed among the components of the polylysine, kappa-carrageenan, and pectin film, forming a robust complex that acts as an effective antimicrobial delivery system [151, 163].

A study investigated the reinforcement of nanocrystalline cellulose in alginate-based biodegradable nanocomposites. The findings exhibited improvements in tension strength, modulus of elasticity, thermal stability, and structural and physical characteristics of the barrier [164]. They also reported decreased swelling properties, water vapour permeability, and extension at the discontinuity of the nanocomposites [164]. The researcher prepared biopolymer films using thymol-loaded nanostructured lipid carriers and nanoemulsions added to Ca-alginate solutions. It increased the films' porosity, surface roughness, thickness, and water vapour permeability and decreased the films' water contact angle, mechanical strength, and swelling ratio [165].

A study found that agar and nanocrystalline cellulose decreased water solubility, water vapour permeability, tensile strength, and elastic modulus. This unique packaging technique improved food safety, shelf life, water contact angle, swelling ratio, viscosity, and elongation at break [89]. Table 4 shows the Modern Application of seaweed polysaccharides in (edible film, coating, and active) packaging systems.

7.2 Antimicrobial properties

Active packaging relies heavily on antimicrobial packaging as it plays a crucial role in addressing the prominent food safety issue within the food industry [74]. When aiming for sustainable and secure packaging, it is essential to consider the inclusion of natural antimicrobial substances like chitosan [182]. Through agar diffusion tests, multiple research studies have demonstrated the antimicrobial activity of crude seaweed extracts that contain a combination of polysaccharides in a survey conducted by [183].

The biochemical characteristics and polysaccharide composition of brown seaweed *Chaetomorpha antennina* were assessed by extracting its polysaccharides using both traditional and alternative methods. Mixed fucoidans extracted using microwave and subcritical water techniques prevented *Escherichia coli* growth. Enzyme-ultrasound, ultrasound microwave, and subcritical water extraction techniques showed antibacterial activity against *Staphylococcus aureus*. All fucoidans had an antiviral impact on HSV-2 infections, regardless of extraction method [184].

Multiple studies have demonstrated that dietary seaweed preparations have a therapeutic effect on infectious diseases in fish [185]. The therapeutic effect of orally administered seaweed polysaccharide preparation, such as *Ascophyllum nodosum*, is a bacterial infection model involving mammals. Previous studies have explored the incorporation of pure

Table 4 Recent Application of Seaweed Organic Compounds in (edible film, coating and active) Packaging System

Compounds	Packaging Matrix	Properties of materials	References
Carrageenan	Chitosan	Preserve the longan fruit while minimizing alterations in quality and quantity, resulting in reduced water loss, weight loss, and respiratory rate in coated fruits	[166]
Fucus vesiculosus	Whey protein	It effectively prevents lipid oxidation in packaged poultry meat, maintaining this protection for at least 25 days during storage while fortifying the thickness, tensile strength, and elastic modulus	[167]
Sodium Alginate	Carboxymethyl cellulose, CaCl ₂ , Pyrogalllic acid	It demonstrates efficacy against <i>E. coli</i> and <i>S. aureus</i> , increasing moisture content, water vapour permeability, oxygen permeability, and excellent elongation at break	[168]
Sodium Alginate	Lemongrass oil	The films successfully suppressed the proliferation of <i>E. coli</i> and <i>L. monocytogenes</i> bacteria and achieved a controlled release of lemongrass oil through microencapsulation	[169]
Furcellaran	Gelatin hydrolysate/Rosemary extract	They included heightened antioxidant activity, elevated thickness, increased water content, enhanced tensile strength, and variations in color at different pH levels	[170]
K and I carrageenan, and Alginate	Glycerol	Alginate contributes to film uniformity and transparency. I-carrageenan influences the film's opacity, while k-carrageenan enhances moisture resistance and tensile properties	[171]
Sodium Alginate	Sodium carboxymethylcellulose, collagen, and Lactococcus lactis	It successfully hinders the growth of <i>S. aureus</i> over seven days, but when <i>Lactococcus lactis</i> is added, it decreases gloss and transparency	[172]
Alginate	Pullulan and Capsaicin	They demonstrate vigorous antibacterial activity against <i>E. coli</i> and <i>S. aureus</i> and improve tensile strength, water vapor permeability, and surface contact angle	[173]
Sodium Alginate	Gelatin-tea polyphenols	Raising the levels of tea polyphenols leads to enhanced antioxidant properties and physical characteristics. However, this also results in compact extension at break and decreased water vapor permeability. On the positive side, it boosts tensile strength, contact angle, and cross-linking	[174]
Sodium Alginate	Ferulic acid	There was no observed inhibition zone against <i>Klebsiella pneumoniae</i> and <i>Salmonella enterica</i> . However, an increase in the concentration of ferulic acid resulted in heightened antioxidant activity	[175]
Agar	Nano-bacterial cellulose	Including bacterial cellulose, which reduces moisture content, water solubility, and water vapour permeability while increasing tensile strength from 22.10 to 44.51 MPa. Additionally, it led to improved crystallinity and enhanced thermal stability	[176]
Agar	Starch and Maltodextrin	Enhanced film-forming capability and hydrophobic characteristics are achieved. Including maltodextrin leads to the formation of highly compatible and flexible starch-agar films. Furthermore, solubility increases with higher maltodextrin concentration	[177]

Table 4 (continued)

Compounds	Packaging Matrix	Properties of materials	References
Semi-refined carrageenan and Ulvan	Glycerol	Semi-refined carrageenan and Ulvan films exhibit a notable capacity for chelating metal ions. Ulvan polysaccharide-based films display robust hydroxyl radical scavenging activity. Films with lower molecular weights demonstrate superior antioxidant activity	[178]
Sodium Alginate	Pomegranate peel extract, Chitosan	Effectively maintained the nutritional properties, delayed senescence, and enhanced the visual characteristics of guava	[179]
Sodium Alginate	Oregano essential oil (1.5–2.5% w/w), mandarin fiber (0.5% w/w)	Efficiently removes pathogens like <i>Staphylococcus aureus</i> and prolongs the shelf life of low-fat sliced cheese	[180]
k-carrageenan	Chitosan	There is a decrease in weight loss and a positive impact on reducing disease infection in dragon fruit stored at 10 °C and 90–95% relative humidity. Additionally, the freshness and bract chlorophyll content are maintained for 30 days	[181]

saccharides like carrageenan and alginate into polymer films; however, these films have demonstrated limited and no detrimental impact against *Escherichia coli* and *Listeria monocytogenes* [186].

However, a film production including the extracts derived from seaweed was developed, and the findings demonstrated a remarkable area of inhibition against gram-positive bacteria. This finding illustrates that the extract can sustain its antibacterial activity. Research on crude seaweed extract shows significant potential for antimicrobial properties of algae polysaccharides [86]. Studies on polysaccharide films have shown their versatility in incorporating different materials, including polymers, nanoclays, and copper nanoparticles. This adaptability enables the production of active antimicrobial packaging that might inhibit microbe growth, thereby increasing the safety and durability of packed products [74, 187, 188].

To investigate the development of films using alginate-extracted seaweed biomass. In their research, control films made of durian starch and carrageenan did not exhibit any inhibitory effect on *Staphylococcus aureus*, as observed in the disk diffusion assay. However, upon incorporating ten weight percent (wt%) carvacrol into the film and subjecting it to rediffusion for 24 and 48 h, a notable and comprehensive suppression of the growth of gram-positive bacteria was observed [132].

Similarly, the composite film based solely on pure carrageenan did not demonstrate antibacterial effects against *E. coli* or *Listeria monocytogenes*. This lack of antibacterial activity persisted even after incorporating nano-clay or nano-silver into the film [138]. An agar blend was studied for its antimicrobial properties against food-borne pathogenic bacteria. Initially, the blend presented no significant antimicrobial activity against gram-positive and harmful bacteria. However, the combination acquired antimicrobial properties upon adding seaweed nanoparticles, which possessed robust antimicrobial activity [188, 189]. A study of the antimicrobial activity of an aqueous extract of *Kappaphycus alvarezii* found that the raw extract was effective against gram-positive bacteria, like *Staphylococcus aureus* and *Bacillus cereus*. The inhibition of *Staphylococcus aureus* was more significant than that of *Bacillus cereus* [190].

7.3 Antioxidant properties

Lipid oxidation significantly influences the quality of food and food products. The formation of agents, including free radicals, during oxidation, can have cytotoxic properties, mutagenic and carcinogenic, leading to serious health issues. Furthermore, oxidation negatively impacts food's shelf life and nutritional value, decreasing product quality [191]. Antioxidant packaging prevents the degradation of lipids and proteins in packaged food and products. Active packaging effectively counteracts oxidation by incorporating or coating antioxidants onto the packaging film. Natural antioxidants have recently gained popularity due to a desire to reduce synthetic ingredients in packaging. Biological additives are considered safe to consume and provide additional health benefits, making them appealing to the market [132, 189, 191].

Gracilaria lemaneiformis exhibited less antioxidant activity due to its non-sulfated polysaccharide nature. On the other hand, green seaweed polysaccharides demonstrated high antioxidant activity, and brown seaweed polysaccharides also displayed a more reducing power [192]. The antioxidant effects were higher than those observed in methanolic or ethanolic seaweed extracts. Incorporating the extract into a Poly vinyl Alcohol film showed a substantial enhancement in radical scavenging activity, surpassing the effects of pure Poly vinyl Alcohol films and those with seaweed extracts [193–195].

More studies have revealed Lambda-carrageenan to have significant antioxidant activity. They provided evidence that incorporating antioxidant-rich polysaccharides can effectively prevent the oxidative deterioration of food [193]. A DPPH experiment assessed the antioxidant capacity of a carrageenan and mulberry extract film. The results showed that carrageenan had an antioxidant activity that depended on the quantity employed and that mulberry extract further increased this activity [193]. The DPPH assay revealed that the pure κ -carrageenan film exhibited moderate antioxidant activity due to its inherent natural polyphenols, nevertheless, including plant essential oils enhanced these results [196].

The results suggest that polysaccharides exhibit varying antioxidant activity, rendering them appropriate for deployment in antioxidant packaging. However, the incorporation of constituents such as essential oils derived from plants and extracts of mulberry can be utilized to amplify the antioxidant characteristics, thereby ensuring efficient mitigation of lipid oxidation and augmenting food safety. Additionally, additional investigation is necessary to comprehensively understand the antioxidant properties exhibited by diverse seaweed polysaccharides and extracts. This will maximize seaweed's antioxidant properties in active food packaging [138].

The use of active components in antioxidant and antimicrobial packaging films to achieve the desired effects on the product. It further emphasizes the need to determine the feasibility of producing such films by examining the active agent diffusivity and release rate into the food [197]. The release of the component is crucial, as it should neither be

extremely rapid, which could cause migration about the internal part of the food, nor excessively slow, which would delay attaining the required inhibitory concentration [12].

A study investigated the migration of natamycin from an antimicrobial film made of an alginate/pectin blend. The researchers measured natamycin concentration in water and determined parameters related to its release. Results showed that natamycin release in pure pectin film occurred within 30 h, composite film over 70 h, and pure alginate film over 800 h. Adding alginate reduced the natamycin diffusion rate, suggesting higher compatibility between natamycin and alginate than pectin [198].

A study analyzed the movement of sorbic acid from a film made of polylactic acid and fucus. The release of sorbic acid was slower in 95 wt% ethanol at 40 °C compared to pure polylactic acid film. However, the pure polylactic acid film's release was significantly lower in 10 wt% ethanol. The polylactic acid/fucus blend showed slightly higher diffusion coefficients, possibly due to weaker immobilization of the antimicrobial agent [199].

The study examined calcium release from alginate films when exposed to ethanol, acetic acid, and oleic acid. It found that the concentration of nanoparticles significantly impacted the release rate, with lower concentrations resulting in slower release due to stronger interactions. Conversely, higher nanoparticle concentrations led to particle aggregation, reducing chemical interaction and accelerating release. Moreover, the acidic nature of acetic acid hastened release compared to ethanol. In contrast, the solubility of nanoparticles in oleic acid and the insolubility of the alginate layer further increased release rates in oleic acid [132].

8 Application of seaweed based food packaging

8.1 Fresh produce packaging

Seaweed-based films have great potential for preserving freshness in products. These films create a protective barrier that prevents the exchange of gases and moisture between the packaged produce and its conditions. Altering the atmosphere of fresh fruit packaging effectively slows down the process of deterioration [74].

Moreover, seaweed-based coatings offer barrier properties that minimize water loss from the product, thereby reducing shrinkage and protecting its firmness. This preservation effect significantly prolongs the freshness and appeal of perishable fruits and vegetables over an extended period. Additionally, these films prevent moisture loss, thereby retaining the inherent juiciness of the produce and improving its sensory characteristics [200]. Packaging fresh produce a critical challenge, preserving the desired texture and nutritional post-packaging values. Seaweed-based films present an ideal solution to attempt these challenges effectively. Their formulation advances a microenvironment favourable to maintaining the texture, thereby modifying issues such as browning, wilting, and softening [201].

Seaweed-based films are crucial in preserving the nutritional value of fruits and vegetables by shielding them from light and preventing the degradation of vitamins and antioxidants. These films also contain bioactive components that can be transferred to vegetables, providing additional health benefits. They extend shelf life and maintain texture and nutritional quality by creating a modified environment that reduces respiration rate and moisture loss, enhancing the freshness, appearance, and nutritional value of packed produce, reducing food waste, and increasing customer satisfaction [200].

Figure 5 A study on tomatoes coated with seaweed found that the seaweed extract, specifically *Kappaphycus alvarezii*, had superior antibacterial and antifungal properties compared to *Sargassum tenerrimum*. The study also assessed quality parameters and shelf life, finding that *K. alvarezii* was more effective in preserving tomato fruit quality and enhancing it during storage. The findings suggest that seaweed gel can improve tomato quality and extend shelf life [134].

8.2 Meat and seafood packaging

Using seaweed-based films is advantageous in protecting and upholding the property of meat and seafood products. Acting as protecting barriers, these coatings effectively block the oxygen and moisture, which contribute significantly to the degradation of seafood and meat. Modified atmosphere packaging and seaweed-based films increase the shelf life of products by inhibiting bacterial growth that leads to spoilage and minimizing oxidative reactions, thus preventing the deterioration of the items [151].

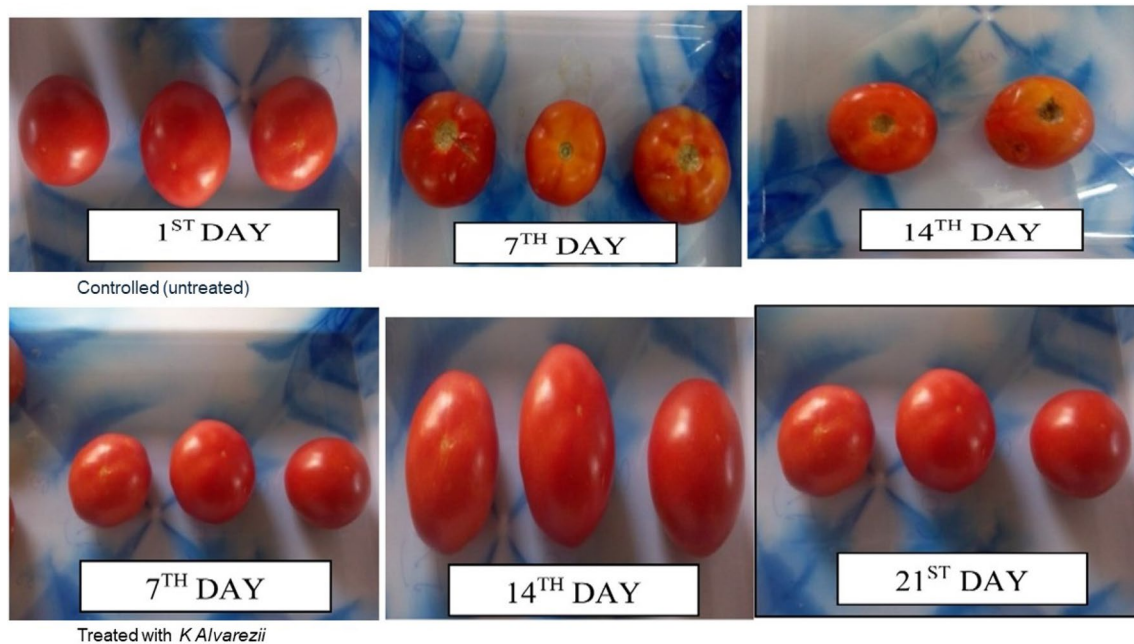


Fig. 5 Shows the effect of texture of seaweed-coated and uncoated tomatoes during storage [134]

These films also possess antibacterial properties due to bioactive compounds found in seaweed, inhibiting harmful bacteria growth, decreasing foodborne disorders, and improving the safety of packaged meat and seafood [202]. Seaweed films have barrier properties that reduce the loss of natural fluids and flavors and preserve the quality of packaged meat and seafood [203].

Seaweed-based films provide barrier properties to maintain the freshness and reliability of packaged items by minimizing humidity loss and resistance against external contamination. They prevent dehydration in meat, ensuring tenderness and juiciness. They retard bacterial growth and microbiological spoilage, extending product shelf life [204].

Seaweed-based films maintain quality and extend shelf life in the highly perishable seafood sector and help to retard deterioration. Preserving seafood in this way enhances the taste, texture, and colour of packaged seafood, reducing the likelihood of food waste [74]. Industries use this packaging for meat and seafood to extend shelf life, reduce food waste, and improve product quality and safety. This sustainable and environmentally friendly method prolongs the freshness of these products, ensuring optimal consumer experience [203]

Figure 6 Shows that the study investigated the impacts of edible coatings made from agar/sodium alginate (AS) and AS combined with ginger essential oil on refrigerated fresh beef quality and shelf life. Results showed that AS + GEO beef maintained enhanced sensory characteristics during storage, proposing potential for further development of antibacterial coating material for preserving frozen beef [205].

8.3 Bakery and confectionary packaging

Seaweed-derived films are used in packaging confectionery to maintain texture and freshness. They act as protective barriers, preventing gas and moisture exchange between products and their environment. This helps keep the desired texture of baked goods by minimizing absorption and reducing product moisture loss, preventing staleness, hardness, or dryness. [200]. Seaweed-based films help preserve baked goods' freshness by creating a microenvironment within the packaging, slowing the staling process, and preventing moisture movement between crust and crumb components, thus keeping their distinct textures and flavours during ageing [203].

These films are essential for preventing staleness and preserving shelf durability in baked goods and confectioneries. They provide a shield against moisture absorption, preventing texture and flavour changes. This helps maintain the crunchiness, fluffiness, and freshness of baked goods like cookies, pastries, and bread [161]. These films preserve

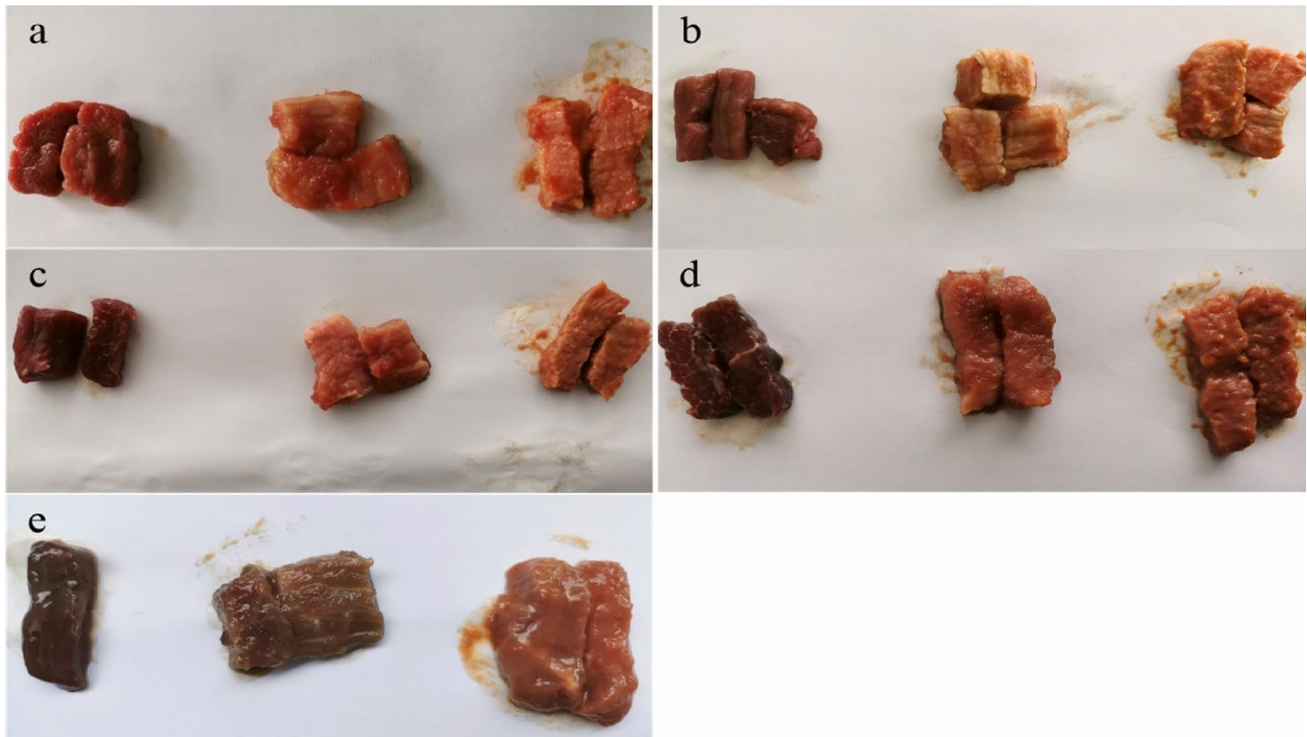


Fig. 6 Images show meat samples texture for different storage times: **a** 3: **b** 6: **c** 9: **d** 15 and **e** 24 days, stored at 4 °C [205]

the shelf-life stability of bread and are sweetened by shielding them from outer effects that accelerate spoilage. Acting as a barrier, these films block light, which can degrade light-sensitive elements such as flavours and colors, etc. Lowering light exposure helps maintain packaged foods' visual appeal, taste, and aroma [206].

9 Conclusions and different challenges

Seaweed contributes essential nutrients, textures, and health-promoting components to food products. Seaweed-based films prefer sustainable food packaging, renewable sourcing, biodegradability, and consumer acceptance. Innovations in seaweed-based films have the potential for sustainable and environmentally friendly packaging solutions, contributing to a more sustainable and circular economy. Seaweed's essential nutrients contribute to gelling agents, colourants, and other food products. Seaweed's biodegradability, transparency, antioxidant, and antimicrobial properties determine its potential in sustainable food packaging. Combining seaweed with biopolymers and additives enhances packaging attributes, yet further research is needed for optimal performance. Incorporating seaweed into polymers aligns with sustainability goals and requires examination of specific effects. Ensuring safety and tailoring mechanical properties through polymer blends or natural additives shows promise for eco-friendly packaging.

Enhancing mechanical strength and durability is crucial for ensuring suitability for packaging and transportation. Research is needed to improve stability and usability over extended periods by mitigating degradation factors like moisture absorption and oxidation. Developing effective and eco-friendly mass production and handling methods is essential for increasing demands. We focus on researching competent and cost-effective techniques to enhance economic competitiveness in production and processing. Furthermore, we are exploring composite materials and novel formulations to enhance functional properties. Detailed studies on seaweed strength, antibacterial properties, and controlled release are needed. Regulatory standards and addressing production costs are crucial for commercial viability. Adapting seaweed-based films for various applications through polymer blends is significant for eco-friendly packaging.

Acknowledgements The Faculty of Chemical and Process Engineering Technology, University of Malaysia Pahang Al Sultan Abdullah, financially supported (Grant No RDU222802 & UIC220818) this work as well as for the additional financial support under PGRS grant PGRS 220350 and Doctoral research scheme (DRS). The author thanks Dr Noormazlinah Binti Ahmad for contributing to this work.

Author contributions Muhammad Qasim Ali: writing original draft, graph, Table, Nur Fathin Ruslan: graph and table: Noormazlinah Ahmad, Mohd Akmal Azhar, Mimi Sakinah Abdul Munim: Luay M Alsubhi, Abeer Essam Noman: Editing and Review of the Article.

Data availability No datasets were generated or analysed during the current study.

Declarations

Competing interests The authors declare no competing interests.

Open Access This article is licensed under a Creative Commons Attribution-NonCommercial-NoDerivatives 4.0 International License, which permits any non-commercial use, sharing, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if you modified the licensed material. You do not have permission under this licence to share adapted material derived from this article or parts of it. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit <http://creativecommons.org/licenses/by-nc-nd/4.0/>.

References

1. Al Mahmud MZ, Mobarak MH, Hossain N. Emerging trends in biomaterials for sustainable food packaging: a comprehensive review. *Heliyon*. 2024;10:e24122. <https://doi.org/10.1016/j.heliyon.2024.e24122>.
2. Nešić A, Cabrera-Barjas G, Dimitrijević-Branković S, Davidović S, Radovanović N, Delattre C. Prospect of polysaccharide-based materials as advanced food packaging. *Molecules*. 2019;25:135. <https://doi.org/10.3390/molecules25010135>.
3. Yan MR, Hsieh S, Ricacho N. Innovative food packaging, food quality and safety, and consumer perspectives. *Processes*. 2022;10:747. <https://doi.org/10.3390/pr10040747>.
4. Karmaus AL, Osborn R, Krishan M. Scientific advances and challenges in safety evaluation of food packaging materials: workshop proceedings. *Regul Toxicol Pharmacol*. 2018;98:80–7. <https://doi.org/10.1016/j.yrtph.2018.07.017>.
5. Asim Z, Shamsi IR, Wahaj M, Raza A, Abul Hasan S, Siddiqui SA, Aladresi A, Sorooshian S, Seng TT. Significance of sustainable packaging: a case-study from a supply chain perspective. *Appl Syst Innov*. 2022;5:117. <https://doi.org/10.3390/asi5060117>.
6. Lomartire S, Marques JC, Gonçalves AMM. An overview of the alternative use of seaweeds to produce safe and sustainable bio-packaging. *Appl Sci*. 2022;12:3123. <https://doi.org/10.3390/app12063123>.
7. Cheng J, Gao R, Zhu Y, Lin Q. Applications of biodegradable materials in food packaging: a review. *Alex Eng J*. 2024;91:70–83. <https://doi.org/10.1016/j.aej.2024.01.080>.
8. Park E, Yu H, Lim J-H, Hee Choi J, Park K-J, Lee J. Seaweed metabolomics: a review on its nutrients, bioactive compounds and changes in climate change. *Food Res Int*. 2023;163: 112221. <https://doi.org/10.1016/j.foodres.2022.112221>.
9. Qasim Ali M, Rana K, Khaledur Rahman Bhuiyan M, Miah S, Zahid S, Ahmed A, Waheed H, Ghosh Chowdhury P, Shanzid Hasana M, Rezwan Ahmed Mahedi M. Current Status and Prospects of Microalgae Bioactive Compounds for Anticancer and Antiviral Actions. *Biomater J*. 2022; 1, 28–36. <https://doi.org/10.5281/zenodo.582940>
10. López-Hortas L, Domínguez H, Torres MD. Valorisation of edible brown seaweeds by the recovery of bioactive compounds from aqueous phase using MHG to develop innovative hydrogels. *Process Biochem*. 2019;78:100–7. <https://doi.org/10.1016/j.procbio.2019.01.010>.
11. Brain-Isasi S, Carú C, Lienqueo ME. Valorization of the green seaweed *Ulva rigida* for production of fungal biomass protein using a hypercellulolytic terrestrial fungus. *Algal Res*. 2021. <https://doi.org/10.1016/j.algal.2021.102457>.
12. Kaushalya KGD, Gunathilake KDPP. Encapsulation of phlorotannins from edible brown seaweed in chitosan: effect of fortification on bioactivity and stability in functional foods. *Food Chem*. 2022. <https://doi.org/10.1016/j.foodchem.2021.132012>.
13. Selvin J, Huxley AJ, Lipton AP. Immunomodulatory potential of marine secondary metabolites against bacterial diseases of shrimp. *Aquaculture*. 2004;230:241–8. [https://doi.org/10.1016/S0044-8486\(03\)00427-7](https://doi.org/10.1016/S0044-8486(03)00427-7).
14. Qasim Ali M, Hasan S, Ahmed Mahedi R, Rahman Bhuiyan K, Afrin S, Bahadur Khadka R, Ore Areche F, Reza R, Miah S, Cruz-Sosa F, Pereira L. A descriptive review on microalgae and microalgal compounds with their interactions. *Rev Article Plant Cell Biotechnol Mol Biol*. 2022;23:56–69.
15. Kadam SU, Tiwari BK, O'Donnell CP. Application of novel extraction technologies for bioactives from marine algae. *J Agric Food Chem*. 2013. <https://doi.org/10.1021/jf400819p>.
16. Quitério E, Soares C, Ferraz R, Delerue-Matos C, Grosso C. Marine health-promoting compounds: recent trends for their characterization and human applications. *Foods*. 2021;10:3100. <https://doi.org/10.3390/foods10123100>.
17. Govaerts F, Ottar Olsen S. Consumers' values, attitudes and behaviours towards consuming seaweed food products: the effects of perceived naturalness, uniqueness, and behavioural control. *Food Res Int*. 2022. <https://doi.org/10.1016/J.FOODRES.2022.112417>.
18. Cai J. Global status of seaweed production, trade and utilization. 2021.

19. Hossain MdS, Sifat SA, Hossain MA, Salleh S, Hossain M, Akter S, Hossain MB. Comparative assessment of bioactive compounds, antioxidant capacity and nutritional quality of red seaweeds and water spinach. *Reg Stud Mar Sci.* 2021;46: 101878. <https://doi.org/10.1016/j.rsm.2021.101878>.
20. Pradhan B, Bhuyan PP, Patra S, Nayak R, Behera PK, Behera C, Behera AK, Ki J-S, Jena M. Beneficial effects of seaweeds and seaweed-derived bioactive compounds: current evidence and future prospective. *Biocatal Agric Biotechnol.* 2022;39: 102242. <https://doi.org/10.1016/j.bcab.2021.102242>.
21. Zhang Q, Qi H, Zhao T, Deslandes E, Ismaeli NM, Molloy F, Critchley AT. Chemical characteristics of a polysaccharide from *Porphyra capensis* (Rhodophyta). *Carbohydr Res.* 2005;340:2447–50. <https://doi.org/10.1016/J.CARRES.2005.08.009>.
22. Nkurunziza D, Sivagnanam SP, Park JS, Cho YJ, Chun BS. Effect of wall materials on the spray drying encapsulation of brown seaweed bioactive compounds obtained by subcritical water extraction. *Algal Res.* 2021. <https://doi.org/10.1016/j.algal.2021.102381>.
23. Robic A, Gaillard C, Sassi JF, Leral Y, Lahaye M. Ultrastructure of Ulvan: a polysaccharide from green seaweeds. *Biopolymers.* 2009;91:652–64. <https://doi.org/10.1002/bip.21195>.
24. Lakshmi DS, Sankaranarayanan S, Gajaria TK, Li G, Kujawski W, Kujawa J, Navia R. A short review on the valorization of green seaweeds and ulvan: feedstock for chemicals and biomaterials. *Biomolecules.* 2020;10:1–20. <https://doi.org/10.3390/biom10070991>.
25. Lahaye M, Robic A. Structure and function properties of Ulvan, a polysaccharide from green seaweeds. *Biomacromol.* 2007;8:1765–74. <https://doi.org/10.1021/bm061185q>.
26. Dulanlebit YH, Hernani H. Overview of extraction methods for extracting seaweed and its applications. *J P enelitian Pendidikan IPA.* 2023;9:817–24. <https://doi.org/10.29303/jppipa.v9i2.3053>.
27. Kumar K, Srivastav S, Sharanagat VS. Ultrasound assisted extraction (UAE) of bioactive compounds from fruit and vegetable processing by-products: a review. *Ultrason Sonochem.* 2021;70: 105325. <https://doi.org/10.1016/j.ultsonch.2020.105325>.
28. Singla M, Sit N. Application of ultrasound in combination with other technologies in food processing: A review. *Ultrason Sonochem.* 2021;73: 105506. <https://doi.org/10.1016/j.ultsonch.2021.105506>.
29. Chavan P, Sharma P, Sharma SR, Mittal TC, Jaiswal AK. Application of high-intensity ultrasound to improve food processing efficiency: a review. *Foods.* 2022;11:122. <https://doi.org/10.3390/foods11010122>.
30. Gil-Martín E, Forbes-Hernández T, Romero A, Cianciosi D, Giampieri F, Battino M. Influence of the extraction method on the recovery of bioactive phenolic compounds from food industry by-products. *Food Chem.* 2022;378: 131918. <https://doi.org/10.1016/j.foodchem.2021.131918>.
31. Maskur M, Sayuti M, Sumandiarsa IK. Environmental-friendly extraction methods to produce bioactive compounds in seaweed. *Res J Chem Environ.* 2023;27:114–21. <https://doi.org/10.25303/2711rjce1140121>.
32. Yin D, Sun X, Li N, Guo Y, Tian Y, Wang L. Structural properties and antioxidant activity of polysaccharides extracted from *Laminaria japonica* using various methods. *Process Biochem.* 2021;111:201–9. <https://doi.org/10.1016/j.procbio.2021.10.019>.
33. Raguraman V, MubarakAli D, Narendrakumar G, Thirugnanasambandam R, Kirubakaran R, Thajuddin N. Unraveling rapid extraction of fucoxanthin from *Padina tetrastomatica*: purification, characterization and biomedical application. *Process Biochem.* 2018;73:211–9. <https://doi.org/10.1016/j.procbio.2018.08.006>.
34. Laeliocattleya RA, Yuniarta Y, Risjani Y, Wulan SN. Characterization of 'novel fucoidan' extracted from brown seaweed (*Sargassum echinocarpum* J Ag) using ultrasound-assisted extraction (UAE) and its potential antioxidant activity. *Nat Prod Res.* 2023. <https://doi.org/10.1080/14786419.2023.2282111>.
35. Liu X, Yin Q, Chen X, Sun P, Liu Y. Ultrasound-assisted extraction of phenolics from *Sargassum carpophyllum*—a kinetics study. *J Phycol.* 2024. <https://doi.org/10.1111/jpy.13477>.
36. Şahin S, Samli R, Tan ASB, Barba FJ, Chemat F, Cravotto G, Lorenzo JM. Solvent-free microwave-assisted extraction of polyphenols from olive tree leaves: antioxidant and antimicrobial properties. *Molecules.* 2017;22:1056. <https://doi.org/10.3390/molecules22071056>.
37. Cikoš A-M, Jokić S, Šubarić D, Jerković I. Overview on the application of modern methods for the extraction of bioactive compounds from marine macroalgae. *Mar Drugs.* 2018;16:348. <https://doi.org/10.3390/md16100348>.
38. Seoane PR, Flórez-Fernández N, Piñeiro EC, González HD. Microwave-assisted water extraction. In: Seoane PR, editor. *Water extraction of bioactive compounds.* Amsterdam: Elsevier; 2017. p. 163–98.
39. López-Salazar H, Camacho-Díaz BH, Ocampo MA, Jiménez-Aparicio AR. Microwave-assisted extraction of functional compounds from plants: a review. *BioResources.* 2023. <https://doi.org/10.15376/biores.18.3.Lopez-Salazar>.
40. Álvarez-Viñas M, Rivas S, Torres MD, Domínguez H. Microwave-assisted extraction of carrageenan from *Sarcopeltis scottsbergii*. *Mar Drugs.* 2023;21:83. <https://doi.org/10.3390/md21020083>.
41. Ferreira-Anta T, Flórez-Fernández N, Torres MD, Mazón J, Domínguez H. Microwave-assisted hydrothermal processing of *rugulopteryx okamurae*. *Mar Drugs.* 2023;21:319. <https://doi.org/10.3390/md21060319>.
42. Yuan Y, Xu X, Jing C, Zou P, Zhang C, Li Y. Microwave assisted hydrothermal extraction of polysaccharides from *Ulva prolifera*: functional properties and bioactivities. *Carbohydr Polym.* 2018;181:902–10. <https://doi.org/10.1016/j.carbpol.2017.11.061>.
43. Zhang R, Yuen AKL, Magnusson M, Wright JT, de Nys R, Masters AF, Maschmeyer T. A comparative assessment of the activity and structure of phlorotannins from the brown seaweed *Carpophyllum flexuosum*. *Algal Res.* 2018;29:130–41. <https://doi.org/10.1016/j.algal.2017.11.027>.
44. Bogolitsyn K, Parshina A, Mamatmyrodov K, Popov N. Recovery of bioactive complex from brown algae by alternative extraction approaches. *Biores Technol Rep.* 2024;26: 101810. <https://doi.org/10.1016/j.biteb.2024.101810>.
45. Verelst L, Sweygers N, Horvath J, Potters G, Dewil R, Appels L. Dedicated microwave extraction allows uniform energy distribution to efficiently solubilize *Ascophyllum nodosum*. *J Environ Chem Eng.* 2024;12: 112773. <https://doi.org/10.1016/j.jece.2024.112773>.
46. Gallego R, Bueno M, Herrero M. Sub- and supercritical fluid extraction of bioactive compounds from plants, food-by-products, seaweeds and microalgae—an update. *Trends Anal Chem.* 2019;116:198–213. <https://doi.org/10.1016/j.trac.2019.04.030>.
47. Uwineza PA, Waśkiewicz A. Recent advances in supercritical fluid extraction of natural bioactive compounds from natural plant materials. *Molecules.* 2020;25:3847. <https://doi.org/10.3390/molecules25173847>.
48. Dias ALB, de Aguiar AC, Rostagno MA. Extraction of natural products using supercritical fluids and pressurized liquids assisted by ultrasound: current status and trends. *Ultrason Sonochem.* 2021;74: 105584. <https://doi.org/10.1016/j.ultsonch.2021.105584>.

49. Misra SK, Pathak K. Supercritical fluid technology for solubilization of poorly water soluble drugs via micro- and nanosized particle generation. *ADMET DMPK*. 2020. <https://doi.org/10.5599/admet.811>.
50. Gondo TF, Jönsson M, Karlsson EN, Sandahl M, Turner C. Extractability, selectivity, and comprehensiveness in supercritical fluid extraction of seaweed using ternary mixtures of carbon dioxide, ethanol, and water. *J Chromatogr A*. 2023;1706: 464267. <https://doi.org/10.1016/j.chroma.2023.464267>.
51. Ahmad MM, Chatha SAS, Iqbal Y, Hussain AI, Khan I, Xie F. Recent trends in extraction, purification, and antioxidant activity evaluation of plant leaf-extract polysaccharides. *Biofuels, Bioprod Biorefin*. 2022;16:1820–48. <https://doi.org/10.1002/bbb.2405>.
52. Putra NR, Yustisia Y, Heryanto RB, Asmalayah A, Miswanti M, Rizkiyah DN, Yunus MAC, Irianto I, Qomariyah L, Rohman GAN. Advancements and challenges in green extraction techniques for Indonesian natural products: a review. *S Afr J Chem Eng*. 2023;46:88–98. <https://doi.org/10.1016/j.sajce.2023.08.002>.
53. Shen L, Pang S, Zhong M, Sun Y, Qayum A, Liu Y, Rashid A, Xu B, Liang Q, Ma H, Ren X. A comprehensive review of ultrasonic assisted extraction (UAE) for bioactive components: principles, advantages, equipment, and combined technologies. *Ultrason Sonochem*. 2023;101: 106646. <https://doi.org/10.1016/j.ultsonch.2023.106646>.
54. Cannavacciuolo C, Pagliari S, Celano R, Campone L, Rastrelli L. Critical analysis of green extraction techniques used for botanicals: trends, priorities, and optimization strategies—a review. *Trends Anal Chem*. 2024;173: 117627. <https://doi.org/10.1016/j.trac.2024.117627>.
55. Usman M, Nakagawa M, Cheng S. Emerging trends in green extraction techniques for bioactive natural products. *Processes*. 2023;11:3444. <https://doi.org/10.3390/pr11123444>.
56. Chemat F, Abert-Vian M, Fabiano-Tixier AS, Strube J, Uhlénbrock L, Gunjević V, Cravotto G. Green extraction of natural products. Origins, current status, and future challenges. *Trends Anal Chem*. 2019;118:248–63. <https://doi.org/10.1016/j.trac.2019.05.037>.
57. Martins R, Barbosa A, Advinha B, Sales H, Pontes R, Nunes J. Green extraction techniques of bioactive compounds: a state-of-the-art review. *Processes*. 2023;11:2255. <https://doi.org/10.3390/pr11082255>.
58. Pradhan B, Bhuyan PP, Patra S, Nayak R, Behera PK, Behera C, Behera AK, Ki JS, Jena M. Beneficial effects of seaweeds and seaweed-derived bioactive compounds: Current evidence and future prospective. *Biocatal Agric Biotechnol*. 2022. <https://doi.org/10.1016/j.bcab.2021.102242>.
59. Rengasamy KR, Mahomoodally MF, Aumeeruddy MZ, Zengin G, Xiao J, Kim DH. Bioactive compounds in seaweeds: an overview of their biological properties and safety. *Food Chem Toxicol*. 2020. <https://doi.org/10.1016/j.fct.2019.111013>.
60. Gullón B, Gagaoua M, Barba FJ, Gullón P, Zhang W, Lorenzo JM. Seaweeds as promising resource of bioactive compounds: overview of novel extraction strategies and design of tailored meat products. *Trends Food Sci Technol*. 2020;100:1–18. <https://doi.org/10.1016/j.tifs.2020.03.039>.
61. Amin MA, Chondra U, Mostafa E, Alam MM. Green seaweed *Ulva lactuca*, a potential source of bioactive peptides revealed by in silico analysis. *Inform Med Unlocked*. 2022. <https://doi.org/10.1016/j.imu.2022.101099>.
62. Kim DH, Lee SB, Jeong GT. Production of reducing sugar from *Enteromorpha intestinalis* by hydrothermal and enzymatic hydrolysis. *Bioresour Technol*. 2014;161:348–53. <https://doi.org/10.1016/j.biortech.2014.03.078>.
63. Rabanal M, Ponce NMA, Navarro DA, Gómez RM, Stortz CA. The system of fucoidans from the brown seaweed *Dictyota dichotoma*: chemical analysis and antiviral activity. *Carbohydr Polym*. 2014;101:804–11. <https://doi.org/10.1016/j.carbpol.2013.10.019>.
64. Susanto E, Fahmi AS, Abe M, Hosokawa M, Miyashita K. Lipids, fatty acids, and fucoxanthin content from temperate and tropical brown seaweeds. *Aquat Procedia*. 2016;7:66–75. <https://doi.org/10.1016/j.aqpro.2016.07.009>.
65. El-Sheekh MM, Bases EA, El-Shenody RA, El Shafay SM. Lipid extraction from some seaweeds and evaluation of its biodiesel production. *Biocatal Agric Biotechnol*. 2021. <https://doi.org/10.1016/j.bcab.2021.102087>.
66. Luo Y, Feng L, Yang G, Mu J. The role of *Ulva fasciata* in the evolution of the microbial community and antibiotic resistance genes in maricultural sediments. *Mar Pollut Bull*. 2021. <https://doi.org/10.1016/j.marpolbul.2020.111940>.
67. Liu Z, Wang Q, Zou D, Yang Y. Effects of selenite on growth, photosynthesis and antioxidant system in seaweeds, *Ulva fasciata* (Chlorophyta) and *Gracilaria lemaneiformis* (Rhodophyta). *Algal Res*. 2018;36:115–24. <https://doi.org/10.1016/j.algal.2018.10.004>.
68. Pliego-Cortés H, Wijesekera I, Lang M, Bourgougnon N, Bedoux G. Current knowledge and challenges in extraction, characterization and bioactivity of seaweed protein and seaweed-derived proteins. *Adv Bot Res*. 2020;95:289–326. <https://doi.org/10.1016/bs.abr.2019.11.008>.
69. Yang Y, Zhang M, Alalawy AI, Almutairi FM, Al-Duais MA, Wang J, Salama ES. Identification and characterization of marine seaweeds for biocompounds production. *Environ Technol Innov*. 2021. <https://doi.org/10.1016/j.eti.2021.101848>.
70. López-López I, Cofrades S, Ruiz-Capillas C, Jiménez-Colmenero F. Design and nutritional properties of potential functional frankfurters based on lipid formulation, added seaweed and low salt content. *Meat Sci*. 2009;83:255–62. <https://doi.org/10.1016/j.meatsci.2009.05.014>.
71. Niemi C, Mortensen AM, Rautenberger R, Matsson S, Gorzsás A, Gentili FG. Rapid and accurate determination of protein content in North Atlantic seaweed by NIR and FTIR spectroscopies. *Food Chem*. 2023. <https://doi.org/10.1016/j.foodchem.2022.134700>.
72. Barta DG, Coman V, Vodnar DC. Microalgae as sources of omega-3 polyunsaturated fatty acids: biotechnological aspects. *Algal Res*. 2021. <https://doi.org/10.1016/j.algal.2021.102410>.
73. Gisbert M, Sineiro J, Moreira R. Polyphenol extraction kinetics from *Ascophyllum nodosum* seaweed employing water and saltwater: Effect of ultrasound sonication. *Algal Res*. 2022. <https://doi.org/10.1016/j.algal.2022.102773>.
74. Carina D, Sharma S, Jaiswal AK, Jaiswal S. Seaweeds polysaccharides in active food packaging: a review of recent progress. *Trends Food Sci Technol*. 2021;110:559–72. <https://doi.org/10.1016/j.tifs.2021.02.022>.
75. Qin N, Pétursdóttir ÁH, Humphries DJ, Desnica N, Newton EE, Vanhatalo A, Halmemies-Beauchet-Filleau A, Bell L, Givens DI, Juniper DT, Gunnlaugsdóttir H, Stergiadis S. Mineral concentrations in milk from cows fed seaweed (*Saccharina latissima*) under different basal protein supplementation. *Food Chem*. 2023. <https://doi.org/10.1016/j.foodchem.2022.134315>.
76. Mouritsen OG, Bagatolli LA, Duelund L, Garvik O, Ipsen JH, Simonsen AC. Effects of seaweed sterols fucosterol and desmosterol on lipid membranes. *Chem Phys Lipids*. 2017;205:1–10. <https://doi.org/10.1016/j.chemphyslip.2017.03.010>.

77. Aryee AN, Agyei D, Akanbi TO. Recovery and utilization of seaweed pigments in food processing. *Curr Opin Food Sci.* 2018;19:113–9. <https://doi.org/10.1016/j.cofs.2018.03.013>.
78. Rubiño S, Peteiro C, Aymerich T, Hortós M. Major lipophilic pigments in Atlantic seaweeds as valuable food ingredients: analysis and assessment of quantification methods. *Food Res Int.* 2022. <https://doi.org/10.1016/j.foodres.2022.111609>.
79. Bouissil S, Pierre G, Alaoui-Talibi ZE, Michaud P, El Modafar C, Delattre C. Applications of algal polysaccharides and derivatives in therapeutic and agricultural fields. *Curr Pharm Des.* 2019;25:1187–99. <https://doi.org/10.2174/1381612825666190425162729>.
80. Nagahawatta DP, Liyanage NM, Jayawardena TU, Yang F, Jayawardena HH, Kurera MJ, Wang F, Fu X, Jeon YJ, Nagahawatta DP, Liyanage NM. Functions and values of sulfated polysaccharides from seaweed. *Algae.* 2023;38:217–40. <https://doi.org/10.4490/algae.2023.38.12.1>.
81. Alba K, Kontogiorgos V. Seaweed polysaccharides (agar, alginate carrageenan). *Encyclopedia Food Chem.* 2018. <https://doi.org/10.1016/B978-0-08-100596-5.21587-4>.
82. Rhein-Knudsen N, Guan C, Mathiesen G, Horn SJ. Expression and production of thermophilic alginate lyases in *Bacillus* and direct application of culture supernatant for seaweed saccharification. *Algal Res.* 2021. <https://doi.org/10.1016/j.algal.2021.102512>.
83. Kim EJ, Fathoni A, Jeong GT, Do Jeong H, Nam TJ, Kong IS, Kim JK. *Microbacterium oxydans*, a novel alginate- and laminarin-degrading bacterium for the reutilization of brown-seaweed waste. *J Environ Manage.* 2013;130:153–9. <https://doi.org/10.1016/j.jenvman.2013.08.064>.
84. Mohd Fauzief NA, Chang LS, Wan Mustapha WA, Md Nor AR, Lim SJ. Functional polysaccharides of fucoidan, laminaran and alginate from Malaysian brown seaweeds (*Sargassum polycystum*, *Turbinaria ornata* and *Padina boryana*). *Int J Biol Macromol.* 2021;167:1135–45. <https://doi.org/10.1016/j.ijbiomac.2020.11.067>.
85. Ermakova S, Sokolova R, Kim SM, Um BH, Isakov V, Zvyagintseva T. Fucoidans from brown seaweeds *Sargassum hornery*, *Eclonia cava*, *Costaria costata*: structural characteristics and anticancer activity. *Appl Biochem Biotechnol.* 2011;164:841–50. <https://doi.org/10.1007/s12010-011-9178-2>.
86. Doh H, Dunno KD, Whiteside WS. Preparation of novel seaweed nanocomposite film from brown seaweeds *Laminaria japonica* and *Sargassum natans*. *Food Hydrocoll.* 2020. <https://doi.org/10.1016/j.foodhyd.2020.105744>.
87. Sterner M, Gröndahl F. Extraction of laminarin from *Saccharina latissima* seaweed using cross-flow filtration. *J Appl Phycol.* 2021;33:1825–44. <https://doi.org/10.1007/s10811-021-02398-Z/TABLES/5>.
88. Mendes M, Fortunato D, Cotas J, Pacheco D, Morais T, Pereira L. Agar content of estuarine seaweed *Gracilaria* using different cultivation methods. *Appl Food Res.* 2022. <https://doi.org/10.1016/j.afres.2022.100209>.
89. Mostafavi FS, Zaeim D. Agar-based edible films for food packaging applications—a review. *Int J Biol Macromol.* 2020;159:1165–76. <https://doi.org/10.1016/j.ijbiomac.2020.05.123>.
90. Guo Z, Wei Y, Zhang Y, Xu Y, Zheng L, Zhu B, Yao Z. Carrageenan oligosaccharides: a comprehensive review of preparation, isolation, purification, structure, biological activities and applications. *Algal Res.* 2022. <https://doi.org/10.1016/j.algal.2021.102593>.
91. Shafie MH, Kamal ML, Zulkiflee FF, Hasan S, Uyup NH, Abdullah S, Mohamed Hussin NA, Tan YC, Zafarina Z. Application of Carrageenan extract from red seaweed (Rhodophyta) in cosmetic products: a review. *J Ind Chem Soc.* 2022. <https://doi.org/10.1016/j.jics.2022.100613>.
92. Hentati F, Tounsi L, Djomdi D, Pierre G, Delattre C, Ursu AV, Fendri I, Abdelkafi S, Michaud P. Bioactive polysaccharides from seaweeds. *Molecules.* 2020;25:3152. <https://doi.org/10.3390/molecules25143152>.
93. Sakthivel R, Devi KP. Antioxidant, anti-inflammatory and anticancer potential of natural bioactive compounds from seaweeds. *Stud Nat Prod Chem.* 2019;63:113–60. <https://doi.org/10.1016/B978-0-12-817901-7.00005-8>.
94. Prajapati VD, Maheriya PM, Jani GK, Solanki HK. Carrageenan: a natural seaweed polysaccharide and its applications. *Carbohydr Polym.* 2014;105:97–112. <https://doi.org/10.1016/j.carbpol.2014.01.067>.
95. Borsani B, De Santis R, Perico V, Penagini F, Pendezza E, Dilillo D, Bosetti A, Zuccotti GV, D’auria E. The role of carrageenan in inflammatory bowel diseases and allergic reactions: where do we stand? *Nutrients.* 2021. <https://doi.org/10.3390/nu13103402>.
96. Ramli NA, Kamaluddin NNA, Adam F. Mechanical, structural and physical properties of carrageenan-gum arabic biocomposite film for hard capsule application. *Solid State Phenom.* 2022;340:11–8. <https://doi.org/10.4028/p-c2wejl>.
97. Wang X, Guo C, Guo H. Progress of Carrageenan-based films and coatings for food packaging applications. *Packag Technol Sci.* 2024;37:533–50. <https://doi.org/10.1002/pts.2806>.
98. Fathiraja P, Gopalrajan S, Karunanithi M, Nagarajan M, Obaiah MC, Durairaj S, Neethirajan N. Response surface methodology model to optimize concentration of agar, alginate and carrageenan for the improved properties of biopolymer film. *Polym Bull.* 2022;79:6211–37. <https://doi.org/10.1007/s00289-021-03797-5>.
99. Zhang J, Yang Y, Zhang J, Shi J, Liu L, Huang X, Song W, Li Z, Zou X, Povey M. High-stability bi-layer films incorporated with liposomes @ anthocyanin/carrageenan/agar for shrimp freshness monitoring. *Foods.* 2023;12:732. <https://doi.org/10.3390/foods12040732>.
100. Giyatmi G, Poetri TA, Irianto HE, Fransiska D, Agusman A. Effect of alginate and polyethylene glycol addition on physical and mechanical characteristics of k-carrageenan-based edible film. *Squalen Bull Marine Fish Postharvest Biotechnol.* 2020;15:41. <https://doi.org/10.15578/squalen.v15i1.418>.
101. Bentil JA, Thygesen A, Lange L, Mensah M, Meyer AS. Green seaweeds (*Ulva fasciata* sp.) as nitrogen source for fungal cellulase production. *World J Microbiol Biotechnol.* 2019. <https://doi.org/10.1007/s11274-019-2658-1>.
102. Ciancia M, Quintana I, Cerezo AS. Overview of anticoagulant activity of sulfated polysaccharides from seaweeds in relation to their structures, focusing on those of green seaweeds. *Curr Med Chem.* 2010;17:2503–29. <https://doi.org/10.2174/092986710791556069>.
103. Robic A, Gaillard C, Sassi J, Lerat Y, Lahaye M. Ultrastructure of ulvan: a polysaccharide from green seaweeds. *Biopolymers.* 2009;91:652–64. <https://doi.org/10.1002/bip.21195>.
104. Amin HH. Safe Ulvan silver nanoparticles composite films for active food packaging. *Am J Biochem Biotechnol.* 2021;17:28–39. <https://doi.org/10.3844/ajbbsp.2021.28.39>.
105. Barakat KM, Ismail MM, Abou El Hassayeb HE, El Sersy NA, Elshobary ME. Chemical characterization and biological activities of ulvan extracted from *Ulva fasciata* (Chlorophyta). *Rend Lincei Sci Fis Nat.* 2022;33:829–41. <https://doi.org/10.1007/s12210-022-01103-7>.

106. Nogueira MT, Chica LR, Yamashita C, Nunes NSS, Moraes ICF, Branco CCZ, Branco IG. Optimal conditions for alkaline treatment of alginate extraction from the brown seaweed *Sargassum cymosum* C. Agardh by response surface methodology. *Appl Food Res*. 2022. <https://doi.org/10.1016/j.afres.2022.100141>.
107. Abka Khajouei R, Keramat J, Hamdami N, Ursu AV, Delattre C, Gardarin C, Lecerf D, Desbrières J, Djelveh G, Michaud P. Effect of high voltage electrode discharge on the physicochemical characteristics of alginate extracted from an Iranian brown seaweed (*Nizimuddiniana zanardini*). *Algal Res*. 2021. <https://doi.org/10.1016/j.algal.2021.102326>.
108. Sapatinha M, Oliveira A, Costa S, Pedro S, Gonçalves A, Mendes R, Bandarra NM, Pires C. Red and brown seaweeds extracts: a source of biologically active compounds. *Food Chem*. 2022. <https://doi.org/10.1016/j.foodchem.2022.133453>.
109. Benslima A, Sellimi S, Hamdi M, Nasri R, Jridi M, Cot D, Li S, Nasri M, Zouari N. The brown seaweed *Cystoseira schiffneri* as a source of sodium alginate: chemical and structural characterization, and antioxidant activities. *Food Biosci*. 2021. <https://doi.org/10.1016/j.fbio.2020.100873>.
110. Flórez-Fernández N, Domínguez H, Torres MD. A green approach for alginate extraction from *Sargassum muticum* brown seaweed using ultrasound-assisted technique. *Int J Biol Macromol*. 2019;124:451–9. <https://doi.org/10.1016/j.ijbiomac.2018.11.232>.
111. Thanh TTT, Quach TTM, Tran VTT, Nguyen TV, Suzuki S, Kitamura S, Yuguchi Y. Structural characteristics and biological activity of different alginate blocks extracted from brown seaweed *Turbinaria ornata*. *J Carbohydr Chem*. 2021;40:97–114. <https://doi.org/10.1080/07328303.2021.1928155>.
112. Pádua D, Rocha E, Gargiulo D, Ramos AA. Bioactive compounds from brown seaweeds: Phloroglucinol, fucoxanthin and fucoidan as promising therapeutic agents against breast cancer. *Phytochem Lett*. 2015;14:91–8. <https://doi.org/10.1016/j.phytol.2015.09.007>.
113. Sultana F, Wahab MA, Nahiduzzaman M, Mohiuddin M, Iqbal MZ, Shakil A, Mamun AA, Khan MS, Wong L, Asaduzzaman M. Seaweed farming for food and nutritional security, climate change mitigation and adaptation, and women empowerment: a review. *Aquac Fish*. 2022. <https://doi.org/10.1016/j.aaf.2022.09.001>.
114. Raja K, Kadirvel V, Subramanian T. Seaweeds, an aquatic plant-based protein for sustainable nutrition—a review. *Future Foods*. 2022. <https://doi.org/10.1016/j.fufo.2022.100142>.
115. Zhang D, Zhang M, Gu X. Seaweed-derived hydrocolloids as food coating and encapsulation agents. In: Zhang D, editor. *Bioactive seaweeds for food applications natural ingredients for healthy diets*. Amsterdam: Elsevier; 2018. p. 153–75. <https://doi.org/10.1016/B978-0-12-813312-5.00008-X>.
116. Baghel RS. Developments in seaweed biorefinery research: a comprehensive review. *Chem Eng J*. 2023;454: 140177. <https://doi.org/10.1016/j.cej.2022.140177>.
117. Qin Y. Production of seaweed-derived food hydrocolloids. In: Qin Y, editor. *Bioactive seaweeds for food applications: natural ingredients for healthy diets*. Amsterdam: Elsevier; 2018. p. 53–69. <https://doi.org/10.1016/B978-0-12-813312-5.00003-0>.
118. Baghel RS, Reddy CRK, Singh RP. Seaweed-based cellulose: applications, and future perspectives. *Carbohydr Polym*. 2021. <https://doi.org/10.1016/j.carbpol.2021.118241>.
119. Rhein-Knudsen N, Meyer AS. Chemistry, gelation, and enzymatic modification of seaweed food hydrocolloids. *Trends Food Sci Technol*. 2021;109:608–21. <https://doi.org/10.1016/j.tifs.2021.01.052>.
120. Gomez LP, Alvarez C, Zhao M, Tiwari U, Curtin J, Garcia-Vaquero M, Tiwari BK. Innovative processing strategies and technologies to obtain hydrocolloids from macroalgae for food applications. *Carbohydr Polym*. 2020. <https://doi.org/10.1016/j.carbpol.2020.116784>.
121. Lee WK, Lim YY, Leow AT, Namasivayam P, Abdullah JO, Ho CL. Biosynthesis of agar in red seaweeds: a review. *Carbohydr Polym*. 2017;164:23–30. <https://doi.org/10.1016/j.carbpol.2017.01.078>.
122. Rhein-Knudsen N, Ale MT, Ajallouei F, Yu L, Meyer AS. Rheological properties of agar and carrageenan from Ghanaian red seaweeds. *Food Hydrocoll*. 2017;63:50–8. <https://doi.org/10.1016/j.foodhyd.2016.08.023>.
123. Albertos I, Martín-Diana AB, Burón M, Rico D. Development of functional bio-based seaweed (*Himantalia elongata* and *Palmaria palmata*) edible films for extending the shelflife of fresh fish burgers. *Food Packag Shelf Life*. 2019. <https://doi.org/10.1016/j.fpsl.2019.100382>.
124. Cian RE, Salgado PR, Drago SR, González RJ, Mauri AN. Development of naturally activated edible films with antioxidant properties prepared from red seaweed *Porphyra columbina* biopolymers. *Food Chem*. 2014;146:6–14. <https://doi.org/10.1016/j.foodchem.2013.08.133>.
125. López-Pérez O, del Olmo A, Picon A, Nuñez M. Volatile compounds and odour characteristics of five edible seaweeds preserved by high pressure processing: changes during refrigerated storage. *Algal Res*. 2021. <https://doi.org/10.1016/j.algal.2020.102137>.
126. Goma M, Hifney AF, Fawzy MA, Abdel-Gawad KM. Use of seaweed and filamentous fungus derived polysaccharides in the development of alginate-chitosan edible films containing fucoidan: study of moisture sorption, polyphenol release and antioxidant properties. *Food Hydrocoll*. 2018;82:239–47. <https://doi.org/10.1016/j.foodhyd.2018.03.056>.
127. Balti R, Mansour MB, Zayoud N, Le Bal'ch R, Brodud N, Arhaliass A, Masse A. Active exopolysaccharides based edible coatings enriched with red seaweed (*Gracilaria gracilis*) extract to improve shrimp preservation during refrigerated storage. *Food Biosci*. 2020. <https://doi.org/10.1016/j.fbio.2019.100522>.
128. Blanco-Pascual N, Montero MP, Gómez-Guillén MC. Antioxidant film development from unrefined extracts of brown seaweeds *Laminaria digitata* and *Ascophyllum nodosum*. *Food Hydrocoll*. 2014;37:100–10. <https://doi.org/10.1016/j.foodhyd.2013.10.021>.
129. Santiago A, Moreira R. Drying of edible seaweeds. *Sustain Seaweed Technol*. 2020. <https://doi.org/10.1016/B978-0-12-817943-7.00004-4>.
130. Agregán R, Franco D, Carballo J, Tomasevic I, Barba FJ, Gómez B, Muchenje V, Lorenzato JM. Shelf life study of healthy pork liver pâté with added seaweed extracts from *Ascophyllum nodosum*, *Fucus vesiculosus* and *Bifurcaria bifurcata*. *Food Res Int*. 2018;112:400–11. <https://doi.org/10.1016/j.foodres.2018.06.063>.
131. Rhein-Knudsen N, Reyes-Weiss D, Horn SJ. Extraction of high purity fucoidans from brown seaweeds using cellulases and alginate lyases. *Int J Biol Macromol*. 2023;229:199–209. <https://doi.org/10.1016/j.IJBIOMAC.2022.12.261>.
132. Cebrián-Lloret V, Metz M, Martínez-Abad A, Knutsen SH, Ballance S, López-Rubio A, Martínez-Sanz M. Valorization of alginate-extracted seaweed biomass for the development of cellulose-based packaging films. *Algal Res*. 2022. <https://doi.org/10.1016/j.algal.2021.102576>.
133. Yamashita C, Freitas Moraes IC, Ferreira AG, Zanini Branco CC, Branco IG. Multi-response optimization of alginate bleaching technology extracted from brown seaweeds by an eco-friendly agent. *Carbohydr Polym*. 2021. <https://doi.org/10.1016/j.carbpol.2020.116992>.

134. Banu AT, Ramani PS, Murugan A. Effect of seaweed coating on quality characteristics and shelf life of tomato (*Lycopersicon esculentum* mill). *Food Sci Hum Wellness*. 2020;9:176–83. <https://doi.org/10.1016/j.fshw.2020.03.002>.
135. Lakshmi DS, Saxena M, Radha KS, Dass LA. Effect of sulfated seaweed polysaccharide on flat sheet polymer (Polysulfone) membrane properties. *Chem Eng J Adv*. 2022. <https://doi.org/10.1016/j.cej.2022.100314>.
136. Rodrigues-Souza I, Pessatti JB, da Silva LR, de Lima BD, de Souza IR, Cestari MM, de Assis HC, Rocha HA, Simas FF, da Silva TE, Leme DM. Protective potential of sulfated polysaccharides from tropical seaweeds against alkylating- and oxidizing-induced genotoxicity. *Int J Biol Macromol*. 2022;211:524–34. <https://doi.org/10.1016/j.ijbiomac.2022.05.077>.
137. Zaitseva OO, Sergushkina MI, Khudyakov AN, Polezhaeva TV, Solomina ON. Seaweed sulfated polysaccharides and their medicinal properties. *Algal Res*. 2022. <https://doi.org/10.1016/j.algal.2022.102885>.
138. Sudhakar MP, Venkatnarayanan S, Dharani G. Fabrication and characterization of bio-nanocomposite films using κ -Carrageenan and *Kappaphycus alvarezii* seaweed for multiple industrial applications. *Int J Biol Macromol*. 2022;219:138–49. <https://doi.org/10.1016/j.ijbiomac.2022.07.230>.
139. Khalil HP, Lai TK, Tye YY, Rizal S, Chong EW, Yap SW, Hamzah AA, Fazita MR, Paridah MT. A review of extractions of seaweed hydrocolloids: properties and applications. *Express Polym Lett*. 2018;12:296–317. <https://doi.org/10.3144/expresspolymlett.2018.27>.
140. Necas J, Bartosikova L. Carrageenan: a review. *Vet Med*. 2013;58:187–205. <https://doi.org/10.17221/6758-VETMED>.
141. Jayakody MM, Vanniarachchy MPG, Wijesekara I. Seaweed derived alginate, agar, and carrageenan based edible coatings and films for the food industry: a review. *J Food Measure Characteriz*. 2022;16:1195–227. <https://doi.org/10.1007/s11694-021-01277-y>.
142. Bisht B, Lohani UC, Kumar V, Gururani P, Sinhmar R. Edible hydrocolloids as sustainable substitute for non-biodegradable materials. *Crit Rev Food Sci Nutr*. 2022;62:693–725. <https://doi.org/10.1080/10408398.2020.1827219>.
143. Wüstenberg T. Cellulose and cellulose derivatives in the food industry. Hoboken: Wiley; 2014.
144. Banerjee S, Bhattacharya S. Food gels: gelling process and new applications. *Crit Rev Food Sci Nutr*. 2012;52:334–46. <https://doi.org/10.1080/10408398.2010.500234>.
145. Shah S, Famta P, Shahrukh S, Jain N, Vambhurkar G, Srinivasarao DA, Raghuvanshi RS, Singh SB, Srivastava S. Multifaceted applications of ulvan polysaccharides: Insights on biopharmaceutical avenues. *Int J Biol Macromol*. 2023;234: 123669. <https://doi.org/10.1016/j.ijbiomac.2023.123669>.
146. Lakshmi DS, Sankaranarayanan S, Gajaria TK, Li G, Kujawski W, Kujawa J, Navia R. A short review on the valorization of green seaweeds and Ulvan: FEEDSTOCK for chemicals and biomaterials. *Biomolecules*. 2020;10:991. <https://doi.org/10.3390/biom10070991>.
147. Wang H, Cao Z, Yao L, Feng T, Song S, Sun M. Insights into the edible and biodegradable Ulvan-based films and coatings for food packaging. *Foods*. 2023;12:1622. <https://doi.org/10.3390/foods12081622>.
148. Amin HH. Ulvan as a new trend in agriculture, food processing and medicine. *Asian J Fish Aquatic Res*. 2020. <https://doi.org/10.9734/ajfar/2020/v6i430105>.
149. Senturk Parreidt T, Müller K, Schmid M. Alginate-based edible films and coatings for food packaging applications. *Foods*. 2018;7:170. <https://doi.org/10.3390/foods7100170>.
150. Rhim J-W. Physical and mechanical properties of water resistant sodium alginate films. *LWT Food Sci Technol*. 2004;37:323–30. <https://doi.org/10.1016/j.lwt.2003.09.008>.
151. Perera KY, Jaiswal AK, Jaiswal S. Biopolymer-based sustainable food packaging materials: challenges, solutions, and applications. *Foods*. 2023;12:2422. <https://doi.org/10.3390/foods12122422>.
152. Shaikh S, Yaqoob M, Aggarwal P. An overview of biodegradable packaging in food industry. *Curr Res Food Sci*. 2021;4:503–20. <https://doi.org/10.1016/j.crfs.2021.07.005>.
153. Pereira L, Cotas J. Seaweed: a sustainable solution for greening drug manufacturing in the pursuit of sustainable healthcare. *Explor Drug Sci*. 2024;2:50–84. <https://doi.org/10.37349/eds.2024.00036>.
154. Sathiamurthy J, Geetha RV, Gopal RK. Production and evaluation of a seaweed-based bioplastic sheet for food packaging. *Uttar Pradesh J Zool*. 2024;45:108–15. <https://doi.org/10.56557/upjz/2024/v45i144184>.
155. El-Sheekh MM, Alwaleed EA, Ibrahim A, Saber H. Preparation and characterization of bioplastic film from the green seaweed *Halimeda opuntia*. *Int J Biol Macromol*. 2024;259: 129307. <https://doi.org/10.1016/j.ijbiomac.2024.129307>.
156. Kajla P, Chaudhary V, Dewan A, Bangar SP, Ramniwas S, Rustagi S, Pandiselvam R. Seaweed-based biopolymers for food packaging: a sustainable approach for a cleaner tomorrow. *Int J Biol Macromol*. 2024;274: 133166. <https://doi.org/10.1016/j.ijbiomac.2024.133166>.
157. Sari WM, Supartono W. Suharno: characterization of biodegradable films from raw seaweed (*Kappaphycus Alvarezii*) and glycerol. *IOP Conf Ser Earth Environ Sci*. 2024;1364: 012081. <https://doi.org/10.1088/1755-1315/1364/1/012081>.
158. Eslami Z, Elkoun S, Robert M, Adjallé K. A review of the effect of plasticizers on the physical and mechanical properties of alginate-based films. *Molecules*. 2023;28:6637. <https://doi.org/10.3390/molecules28186637>.
159. Majeed T. Seaweed selection: exploring best type for bioplastic production. *Natl J Biol Sci*. 2023. <https://doi.org/10.37605/v4i2/3>.
160. Sharma U, Jadaun S, Khapudaang R, Siddiqui S. Seaweed—a sustainable food source in the food industry. Presented at the (2024)
161. Waseem M, Khan MU, Majeed Y, Ntsefong GN, Kirichenko I, Klopova A, Trushov P, Lodygin A. Seaweed-based films for sustainable food packaging: properties, incorporation of essential oils, applications, and future directions. *Potravinarstvo Slovak J Food Sci*. 2023;17:899–917. <https://doi.org/10.5219/1908>.
162. Shahbazi M, Ettelaie R, Rajabzadeh G. Physico-mechanical analysis data in support of compatibility of chitosan/ κ -carrageenan polyelectrolyte films achieved by ascorbic acid, and the thermal degradation theory of κ -carrageenan influencing the properties of its blends. *Data Brief*. 2016;9:648–60. <https://doi.org/10.1016/j.dib.2016.09.039>.
163. Rhim JW, Park HM, Ha CS. Bio-nanocomposites for food packaging applications. *Prog Polym Sci*. 2013;38:1629–52. <https://doi.org/10.1016/j.progpolymsci.2013.05.008>.
164. Rhein-Knudsen N, Reyes-Weiss D, Horn SJ, Horn SJ. Extraction of high purity fucoidans from brown seaweeds using cellulases and alginate lyases. *Int J Biol Macromol*. 2022. <https://doi.org/10.1016/j.ijbiomac.2022.12.261>.
165. Karimi-Khorrami N, Radi M, Amiri S, Abedi E, McClements DJ. Fabrication, characterization, and performance of antimicrobial alginate-based films containing thymol-loaded lipid nanoparticles: comparison of nanoemulsions and nanostructured lipid carriers. *Int J Biol Macromol*. 2022;207:801–12. <https://doi.org/10.1016/j.ijbiomac.2022.03.149>.

166. Lin MG, Lasekan O, Saari N, Khairunniza-Bejo S. Effect of chitosan and carrageenan-based edible coatings on post-harvested longan (*Dimocarpus longan*) fruits. *CyTA J Food*. 2018;16:490–7. <https://doi.org/10.1080/19476337.2017.1414078>.
167. Andrade MA, Barbosa CH, Souza VGL, Coelho IM, Reboleira J, Bernardino S, Ganhão R, Mendes S, Fernando AL, Vilarinho F, Sanches Silva A, Ramos F. Novel active food packaging films based on whey protein incorporated with seaweed extract: development, characterization, and application in fresh poultry meat. *Coatings*. 2021;11:229. <https://doi.org/10.3390/coatings11020229>.
168. Han Y, Wang L. Sodium alginate/carboxymethyl cellulose films containing pyrogallol: physical and antibacterial properties. *J Sci Food Agric*. 2017;97:1295–301. <https://doi.org/10.1002/jsfa.7863>.
169. Bustos CRO, Alberti RFV, Matiacevich SB. Edible antimicrobial films based on microencapsulated lemongrass oil. *J Food Sci Technol*. 2016;53:832–9. <https://doi.org/10.1007/s13197-015-2027-5>.
170. Jancikova S, Jámroz E, Kulawik P, Tkaczewska J, Dordevic D. Furcellaran/gelatin hydrolysate/rosemary extract composite films as active and intelligent packaging materials. *Int J Biol Macromol*. 2019;131:19–28. <https://doi.org/10.1016/j.ijbiomac.2019.03.050>.
171. Paula GA, Benevides NMB, Cunha AP, de Oliveira AV, Pinto AMB, Morais JPS, Azeredo HMC. Development and characterization of edible films from mixtures of κ -carrageenan, ι -carrageenan, and alginate. *Food Hydrocoll*. 2015;47:140–5. <https://doi.org/10.1016/j.foodhyd.2015.01.004>.
172. Ma D, Jiang Y, Ahmed S, Qin W, Liu Y. Physical and antimicrobial properties of edible films containing *Lactococcus lactis*. *Int J Biol Macromol*. 2019;141:378–86. <https://doi.org/10.1016/j.ijbiomac.2019.09.006>.
173. Zhang S, Wei F, Han X. An edible film of sodium alginate/pullulan incorporated with capsaicin. *New J Chem*. 2018;42:17756–61. <https://doi.org/10.1039/C8NJ04249G>.
174. Dou L, Li B, Zhang K, Chu X, Hou H. Physical properties and antioxidant activity of gelatin-sodium alginate edible films with tea polyphenols. *Int J Biol Macromol*. 2018;118:1377–83. <https://doi.org/10.1016/j.ijbiomac.2018.06.121>.
175. Yerramathi BB, Kola M, Annem Muniraj B, Aluru R, Thirumanyam M, Zyryanov GV. Structural studies and bioactivity of sodium alginate edible films fabricated through ferulic acid crosslinking mechanism. *J Food Eng*. 2021;301: 110566. <https://doi.org/10.1016/j.jfoodeng.2021.110566>.
176. Wang X, Guo C, Hao W, Ullah N, Chen L, Li Z, Feng X. Development and characterization of agar-based edible films reinforced with nanobacterial cellulose. *Int J Biol Macromol*. 2018;118:722–30. <https://doi.org/10.1016/j.ijbiomac.2018.06.089>.
177. Wongphan P, Harnkarnsujarit N. Characterization of starch, agar and maltodextrin blends for controlled dissolution of edible films. *Int J Biol Macromol*. 2020;156:80–93. <https://doi.org/10.1016/j.ijbiomac.2020.04.056>.
178. Ramu Ganesan A, Shanmugam M, Bhat R. Producing novel edible films from semi refined carrageenan (SRC) and ulvan polysaccharides for potential food applications. *Int J Biol Macromol*. 2018;112:1164–70. <https://doi.org/10.1016/j.ijbiomac.2018.02.089>.
179. Nair MS, Tomar M, Punia S, Kukula-Koch W, Kumar M. Enhancing the functionality of chitosan- and alginate-based active edible coatings/films for the preservation of fruits and vegetables: a review. *Int J Biol Macromol*. 2020;164:304–20. <https://doi.org/10.1016/j.ijbiomac.2020.07.083>.
180. Artiga-Artigas M, Acevedo-Fani A, Martín-Belloso O. Improving the shelf life of low-fat cut cheese using nanoemulsion-based edible coatings containing oregano essential oil and mandarin fiber. *Food Control*. 2017;76:1–12. <https://doi.org/10.1016/j.foodcont.2017.01.001>.
181. Nguyen HT, Boonyariththongchai P, Buanong M, Supapvanich S, Wongs-Aree C. Chitosan- and κ -carrageenan-based composite coating on dragon fruit (*Hylocereus undatus*) pretreated with plant growth regulators maintains bract chlorophyll and fruit edibility. *Sci Hortic*. 2021;281: 109916. <https://doi.org/10.1016/j.scienta.2021.109916>.
182. Marzban A, Mirzaei SZ, Karkhane H, Ghotekar SK, Danesh A. Biogenesis of copper nanoparticles assisted with seaweed polysaccharide with antibacterial and antibiofilm properties against methicillin-resistant *Staphylococcus aureus*. *J Drug Deliv Sci Technol*. 2022. <https://doi.org/10.1016/j.jddst.2022.103499>.
183. Aluta UP, Aderolu AZ, Ishola IO, Alyassin M, Morris GA, Olajide OA. Polysaccharides from tropical green seaweed *Chaetomorpha antennina* induces non-specific immune responses and improves antioxidative activities in common carp (*Cyprinus carpio*) leukocyte culture cell line. *Algal Res*. 2022. <https://doi.org/10.1016/j.algal.2022.102872>.
184. Zhang B, Liu Y, Wang H, Liu W, Cheong KL, Teng B. Characterization of seaweed polysaccharide-based bilayer films containing essential oils with antibacterial activity. *LWT*. 2021. <https://doi.org/10.1016/j.lwt.2021.111961>.
185. Gora AH, Sahu NP, Sahoo S, Rehman S, Dar SA, Ahmad I, Agarwal D. Effect of dietary *Sargassum wightii* and its fucoidan-rich extract on growth, immunity, disease resistance and antimicrobial peptide gene expression in *Labeo rohita*. *Int Aquat Res*. 2018;10:115–31. <https://doi.org/10.1007/S40071-018-0193-6/TABLES/5>.
186. Okimura T, Jiang Z, Komatsubara H, Hirasaka K, Oda T. Therapeutic effects of an orally administered edible seaweed-derived polysaccharide preparation, ascophyllan HS, on a streptococcus pneumoniae infection mouse model. *Int J Biol Macromol*. 2020;154:1116–22. <https://doi.org/10.1016/j.ijbiomac.2019.11.053>.
187. Thanigavel S, Vickram S, Saranya V, Ali H, Alarifi S, Modigunta JKR, Anbarasu K, Lakshmipathy R, Rohini K. Seaweed polysaccharide mediated synthesis of silver nanoparticles and its enhanced disease resistance in *Oreochromis mossambicus*. *J King Saud Univ Sci*. 2022. <https://doi.org/10.1016/j.jksus.2021.101771>.
188. Santinon C, Ochi D, Beppu MM, Vieira MGA. Chemical modifications in the structure of seaweed polysaccharides as a viable antimicrobial application: a current overview and future perspectives. *Algal Res*. 2022. <https://doi.org/10.1016/j.algal.2022.102796>.
189. Joshi CG, Danagoudar A, Poyya J, Kudva AK, Dhananjaya BL. Biogenic synthesis of gold nanoparticles by marine endophytic fungus *Cladosporium cladosporioides* isolated from seaweed and evaluation of their antioxidant and antimicrobial properties. *Process Biochem*. 2017;63:137–44. <https://doi.org/10.1016/j.procbio.2017.09.008>.
190. Kanatt SR, Lahare P, Chawla SP, Sharma A. *Kappaphycus alvarezii*: its antioxidant potential and use in bioactive packaging films. *J Microbiol Biotechnol Food Sci*. 2015;5:1–6. <https://doi.org/10.15414/jmbfs.2015.5.1.1-6>.
191. Singh S, Gaikwad KK, Lee YS. Antimicrobial and antioxidant properties of polyvinyl alcohol bio composite films containing seaweed extracted cellulose nano-crystal and basil leaves extract. *Int J Biol Macromol*. 2018;107:1879–87. <https://doi.org/10.1016/j.ijbiomac.2017.10.057>.

192. Burgos-Díaz C, Opazo-Navarrete M, Palacios JL, Verdugo L, Anguita-Barrales F, Bustamante M. Food-grade bioactive ingredient obtained from the *Durvillaea incurvata* brown seaweed: antibacterial activity and antioxidant activity. *Algal Res.* 2022. <https://doi.org/10.1016/j.algal.2022.102880>.
193. Rudke AR, da Silva M, de Andrade CJ, Vitali L, Ferreira SRS. Green extraction of phenolic compounds and carrageenan from the red alga *Kappaphycus alvarezii*. *Algal Res.* 2022. <https://doi.org/10.1016/j.algal.2022.102866>.
194. Kumar KS, Ganesan K, Rao PVS. Antioxidant potential of solvent extracts of *Kappaphycus alvarezii* (Doty) doty—an edible seaweed. *Food Chem.* 2008;107:289–95. <https://doi.org/10.1016/j.foodchem.2007.08.016>.
195. Vaghela P, Das AK, Trivedi K, Anand KGV, Shinde P, Ghosh A. Characterization and metabolomics profiling of *Kappaphycus alvarezii* seaweed extract. *Algal Res.* 2022. <https://doi.org/10.1016/j.algal.2022.102774>.
196. Saluri K, Tuvikene R. Anticoagulant and antioxidant activity of lambda- and theta-carrageenans of different molecular weights. *Bioactive Carbohydr Dietary Fibre.* 2020. <https://doi.org/10.1016/j.bcdf.2020.100243>.
197. Lim SJ, Chang LS, Fazry S, Wan Mustapha WA, Babji AS. Functional food & ingredients from seaweed, edible bird's nest and tropical fruits: a translational research. *LWT.* 2021. <https://doi.org/10.1016/j.lwt.2021.112164>.
198. Bierhalz ACK, Da Silva MA, Kieckbusch TG. Natamycin release from alginate/pectin films for food packaging applications. *J Food Eng.* 2012;110:18–25. <https://doi.org/10.1016/j.jfoodeng.2011.12.016>.
199. Jipa IM, Stoica-Guzun A, Stroescu M. Controlled release of sorbic acid from bacterial cellulose based mono and multilayer antimicrobial films. *LWT.* 2012;47:400–6. <https://doi.org/10.1016/j.lwt.2012.01.039>.
200. Khalil HA, Saurabh CK, Tye YY, Lai TK, Easa AM, Rosamah E, Fazita MR, Syakir MI, Adnan AS, Fizree HM, Aprilia NA. Seaweed based sustainable films and composites for food and pharmaceutical applications: a review. *Renew Sustain Energy Rev.* 2017;77:353–62. <https://doi.org/10.1016/j.rser.2017.04.025>.
201. Berry TM, Defraeye T, Shrivastava C, Ambaw A, Coetzee C, Opara UL. Designing ventilated packaging for the fresh produce cold chain. *Food Bioprod Processing.* 2022;134:121–49. <https://doi.org/10.1016/j.fbp.2022.04.005>.
202. Rengasamy KRR, Mahomoodally MF, Aumeeruddy MZ, Zengin G, Xiao J, Kim DH. Bioactive compounds in seaweeds: An overview of their biological properties and safety. *Food Chem Toxicol.* 2020;135:111013. <https://doi.org/10.1016/j.fct.2019.111013>.
203. Ebrahimzadeh S, Biswas D, Roy S, McClements DJ. Incorporation of essential oils in edible seaweed-based films: a comprehensive review. *Trends Food Sci Technol.* 2023;135:43–56. <https://doi.org/10.1016/j.tifs.2023.03.015>.
204. Albertos I, Martin-Diana AB, Burón M, Rico D. Development of functional bio-based seaweed (*Himantalia elongata* and *Palmaria palmata*) edible films for extending the shelflife of fresh fish burgers. *Food Packag Shelf Life.* 2019;22: 100382. <https://doi.org/10.1016/j.fpsl.2019.100382>.
205. Zhang B, Liu Y, Wang H, Liu W, Cheong K, Teng B. Effect of sodium alginate-agar coating containing ginger essential oil on the shelf life and quality of beef. *Food Control.* 2021;130: 108216. <https://doi.org/10.1016/j.foodcont.2021.108216>.
206. Trindade MA, Nunes C, Coimbra MA, Gonçalves FJ, Marques JC, Gonçalves AM. Sustainable and biodegradable active films based on seaweed compounds to improve shelf life of food products. In: Rao AR, editor. *Sustainable global resources of seaweeds*, vol. 2. Cham: Springer International Publishing; 2022. p. 235–52.

Publisher's Note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.