

EFFECTS OF TRANSMEMBRANE PRESSURE ON THE  
PERFORMANCE OF FLAT SHEET MEMBRANE ON MILK PROTEIN  
SEPARATION

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A thesis submitted in fulfillment  
of the requirements for the award of the degree of  
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May 2009

I declare that this thesis entitled “Effect of Transmembrane Pressure on the performance of Flat Sheet Membrane on milk protein separation” is the result of my own research except as cited in references. The thesis has not been accepted for any degree and is not concurrently submitted in candidature of any other degree.

Signature :.....

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Date : May 2009

*Special Dedication to my family members,  
my friends, my fellow colleague and all faculty members  
For all your care, support and believe in me.*

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## ABSTRACT

This research is to study the effect of transmembrane pressure on cross flow filtration on protein milk separation. The different transmembrane pressure will be effect the performance of cross flow filtration. By this research, the optimum operation condition of the cross flow filtration will get especially when comes to protein milk separation. In other word, the suitable parameter will be known in order to separate protein milk. Protein milk can be separated from other composition in the skim milk using cross flow filtration. Before using filtration, skim milk should be centrifuged to remove fat and undesired composition to reduce membrane faultiness or membrane blockage. Protein compositions in both permeate and retentate analyzed using Lowry method. From the result, the optimum operation condition for milk protein separation is at TMP 25 psi and flux recorded is 94 ml/ (m<sup>2</sup>.min) and 78 ml/ (m<sup>2</sup>.min). This is experiment using 10K membrane cassette in batch mode.

## ABSTRAK

Kajian ini adalah untuk mengetahui kesan tekanan transmembran pada prestasi penyaringan aliran bersilang terutamanya apabila ingin memisahkan protein susu. Pelbagai nilai tekanan transmembrane akan mempengaruhi prestasi penyaringan aliran bersilang. Dengan adanya kajian ini, nilai keadaan optima operasi dapat diketahui terutamanya yang berkaitan dengan pemisahan protein susu daripada susu sejat. Protein susu dapat dipisahkan daripada lain – lain komposisi menggunakan kaedah penyaringan membrane. Sebelum menggunakan kaedah penyaringan, susu sejat hendaklah dilakukan pengemparan untuk memisahkan lemak dan komponen lain yang tidak diinginkan. Selain itu, ia dapat mengurangkan keadaan kegagalan atau membrane tersumbat. Kandungan protein yang melalui membrane dan yang tidak melalui membrane di analisa menggunakan kaedah Lowry. mengikut kajian yang dijalankan, pemisahan protein susu adalah terbaik pada TMP 25 yang mencatatkan nilai flux 94 ml/ (m<sup>2</sup>.min) dan 87 ml/ (m<sup>2</sup>.min). kajian ini menggunakan 10K membrane kaset dalam keadaan ultrafiltration.

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**LIST OF ABBREVIATIONS**

|                 |                               |
|-----------------|-------------------------------|
| CFF             | Cross flow filtration         |
| Da              | Dalton                        |
| FDA's           | Food & Drug Administration    |
| MF              | Microfiltration               |
| MW              | Molecular weight              |
| MWCO            | Molecular Weight Cut Off      |
| NF              | Nano filtration               |
| NFF             | Normal Flow Filtration        |
| OD              | Optical density               |
| PV              | Permeate volume               |
| RO              | Reverse Osmosis               |
| RV              | Retentate volume              |
| TMP             | Transmembrane Pressure        |
| UF              | Ultrafiltration               |
| UTMP            | Uniform Transmembran pressure |
| V <sub>cf</sub> | Cross flow velocity           |

**LIST OF SYMBOLS**

|                 |   |                                      |
|-----------------|---|--------------------------------------|
| $\Delta P$      | - | Differnce of pressure                |
| $\Delta P_{TM}$ | - | Difference of transmembrane pressure |
| JC              | - | Cleaning flux                        |
| JT              | - | Total flux                           |
| JW              | - | Water flux                           |
| L               | - | Liter                                |
| $\alpha$        | - | Alpha                                |
| $\beta$         | - | Beta                                 |

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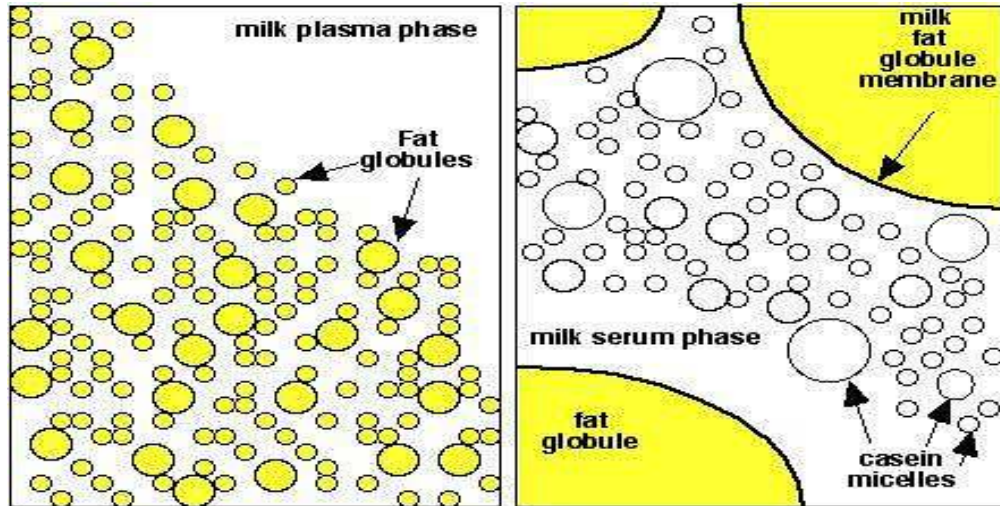
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## **CHAPTER 1**

### **INTRODUCTION**

#### **1.1 Milk protein**

The word protein can be defined as any group of complex organic compounds, consisting essentially of combinations of amino acids in peptide linkages, which contain carbon, hydrogen, oxygen, nitrogen and usually sulfur. Widely distributed in plants and animals, proteins are principal constituent of the protoplasm of all cells and are essential to life. Protein is derived from Greek word meaning 'first' or 'primary', because of the fundamental role of proteins in sustaining life (Morris, 1992). Milk is an emulsion of fat globules, and a suspension of casein micelles (casein, calcium, phosphorous), all of which are suspended in an aqueous phase that contains solublized lactose, whey proteins, and some minerals. Leukocytes in milk are part of the suspended phase. This can be shown clearly as in Figure1.



**Figure 1:** Milk component diagram

Application of the membrane technology in processing milk and dairy product have been shown successfully since 1970 (Y.Poulit, 2008) and this widely used nowadays. Membrane filtrations commonly use to concentrate milk protein from skim milk. The membrane separation is base on molecular cut off (MWCO) of the targeted product. In protein separation technology, membrane technology is more popular than other protein separation method.

## 1.2 Problem statement

The identification of the problem is to determine operating condition to investigate the effectiveness of cross flow filtration in protein milk separation through identification of the optimum parameter condition. The optimum parameters are varies with other operation condition. Transmembrane pressure one of parameters that influence the performance of cross flow filtration and need to studied and identified. The way to identify which tranmembrane pressure is required in protein milk separation is by conducting an experiment. By using different transmembrane pressure, the correct value of transmembrane pressure may be can be



determined. Beside that other parameter such as velocity and pressure in both permeate and retentate can be identified.

### **1.3 Objectives**

The objective of this study is to optimize the usage of cross flow filtration system in milk protein separation and to determine the effects of transmembrane pressure (TMP) on cross flow filtration performance.

### **1.4 Scope of study**

The objective can be achieved by varying the transmembrane pressure until get the optimum separation on milk protein separation. This study will be prepared by using cross flow flat sheet membrane. The other parameter concerned on this experiment is cross flow velocity and pressure both permeate and retentate stream by using 10 K membrane cassette.

## CHAPTER 2

### LITERATURE REVIEW

#### 2.1 Milk protein component

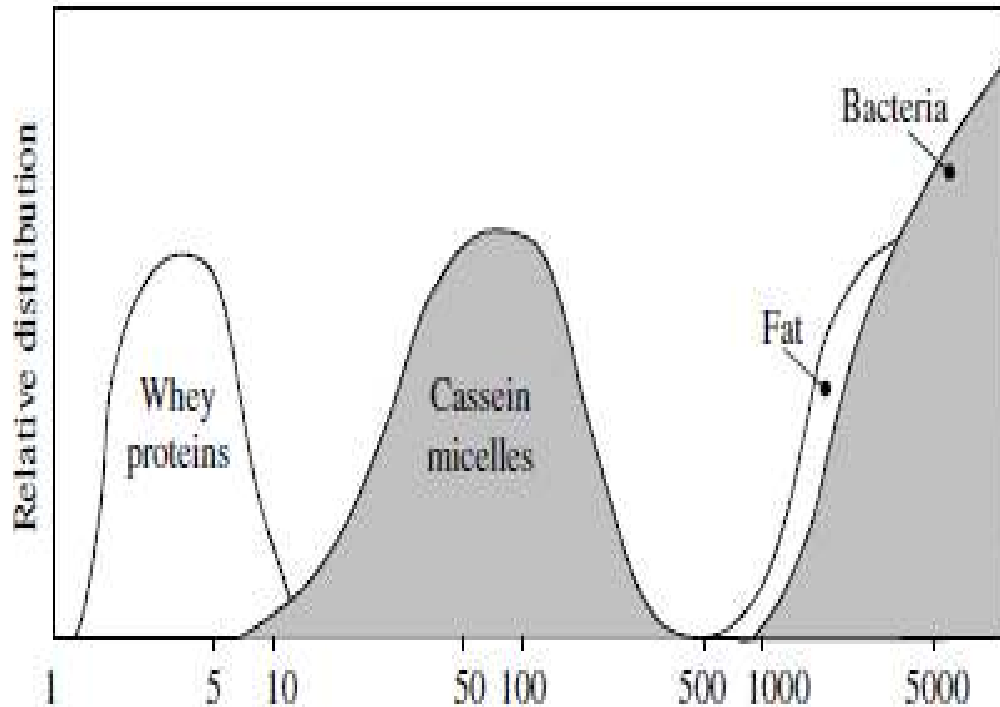
The total protein component of milk is composed of numerous specific proteins. The primary group of milk proteins is the caseins. There are 3 or 4 caseins in the milk of most species, the different caseins are distinct molecules but are similar in structure. All other proteins found in milk are grouped together under the name of whey proteins. The major whey proteins in cow milk are  $\beta$ -lactoglobulin and  $\alpha$ -lactalbumin. The major milk proteins, including the caseins,  $\beta$ -lactoglobulin and  $\alpha$ -lactalbumin, are synthesized in the mammary epithelial cells and are only produced by the mammary gland. The immunoglobulin and serum albumin in milk are not synthesized by the epithelial cells. Instead, they are absorbed from the blood (both serum albumin and the immunoglobulins). An exception to this is that a limited amount of immunoglobulin is synthesized by lymphocytes which reside in the mammary tissue (called plasma cells). These latter cells provide the mammary gland with local immunity. Caseins have an appropriate amino acid composition that is important for growth and development of the nursing young. This high quality protein in cow milk is one of the key reasons why milk is such an important human food.

Caseins are highly digestible in the intestine and are a high quality source of amino acids. Most whey proteins are relatively less digestible in the intestine, although all of them are digested to some degree. When substantial whey protein is not digested fully in the intestine, some of the intact protein may stimulate a localized intestinal or a systemic immune response. This is sometimes referred to as milk protein allergy and is most often thought to be caused by  $\beta$ -lactoglobulin. Properties of protein milk can be seen in table 1. Casein is the highest percentage protein in skim milk (3% -55%) compared to other proteins such as alpha-Lactalbumin (2% - 5%).

**Table 1:** Biochemical characteristics of milk proteins (RD Bremel, University of Wisconsin)

| Protein                   | Approx.% of skim milk protein | Isoelectric point | Molecular weight |
|---------------------------|-------------------------------|-------------------|------------------|
| alpha-Casein              | 45-55                         | 4.1               | 23,000           |
| kappa-Casein              | 8-15                          | 4.1               | 19,000           |
| beta-Casein               | 25-35                         | 4.5               | 24,000           |
| gamma-Casein              | 3-7                           | 5.8-6.0           | --               |
| alpha-Lactalbumin         | 2-5                           | 5.1               | 14,437           |
| beta-Lactoglobulin        | 7-12                          | 5.3               | 18,000           |
| Blood serum albumin       | 0.7-1.3                       | 4.7               | 68,000           |
| Lactoferrin               | 0.2-0.8                       | --                | 87,000           |
| Immunoglobulins:          | --                            | --                | --               |
| IgG1                      | 1-2                           | --                | 160,000          |
| IgG2                      | 0.2-0.5                       | --                | 160,000          |
| IgM                       | 0.1-0.2                       | --                | ~1,000,000       |
| IgA                       | 0.05-0.10                     | --                | ~400,000         |
| Proteose peptone fraction | 2-6                           | 3.3-3.7           | 4,100 to 200,000 |

Major size distribution of milk component (Cheryan and Alvarez, 1995) can be interpreted as in figure 2. Most components such as whey proteins, casein micelles, fat and bacteria can be see clearly the size of each component. Milk protein is in range 1 to 500 while fat and bacteria are having size greater than 500.



**Figure 2:** Size distribution of major milk components

Another component and size distribution can be see in Table 2. Both chloride ion and calcium ion have a same diameter (0.4nm) and water 0.3nm in diameter. The protein (casein) has diameter size 25-130 nm while fat is 2000-10000nm. In protein milk separation (filtration) it is necessary to remove fat first before undergoes filtration process. This is due to minimizing the layer forming at the membrane surface during filtration.

**Table 2:** Molecular size of milk component

| Component                                  | Molecular weight | Diameter (nm) |
|--|------------------|---------------|
| Water                                      | 18               | 0.3           |
| Chloride ion                               | 35               | 0.4           |
| Calcium ion                                | 40               | 0.4           |
| Lactose                                    | 342              | 0.8           |
| a-lactalbumin                              | 14500            | 3             |
| b-lactalbumin                              | 36000            | 4             |
| Blood serum albumin                        | 69000            | 5             |
| Casein micelles (milk protein in solution) | 107–109          | 25–130        |
| Fat  | -                | 2000–10000    |

## 2.2 Membrane protein separation

Membrane filtration becomes more popular bioseparation application especially among dairy product. The principle of membrane separation is utilized in the dairy industry serve as a different purpose. Membrane filtration separation technique can be classified as Reverse osmosis, Nanofiltration, Ultrafiltration and Microfiltration base on pore size. Reverse osmosis use for dehydration of whey. Nanofiltration usually used when partial desalination of whey. While Ultrafiltration is typically used to concentrate of milk proteins in milk and whey and for protein standardization of milk intended for cheese, yoghurt and some other products. For Microfiltration, basically used for reduction of bacteria in skim milk, whey and brine, besides defatting of whey (Goude-dranche, Fauquant, &maubois, 2000) intended for whey protein concentrate and for protein fractionation. RO, NF, UF, and MF cover entire separation domain of milk constituents, from casein micelles to monovalent

ions (Y.Pouliot, 2008). Beside protein separation, UF and MF also used to concentrate juice (B. Sarkar et al, 2008). To separate protein milk, the type of filtration has to determine. Figure 2.1 is spectrum diagram of membrane. By using spectrum diagram, the separation process can be decided.

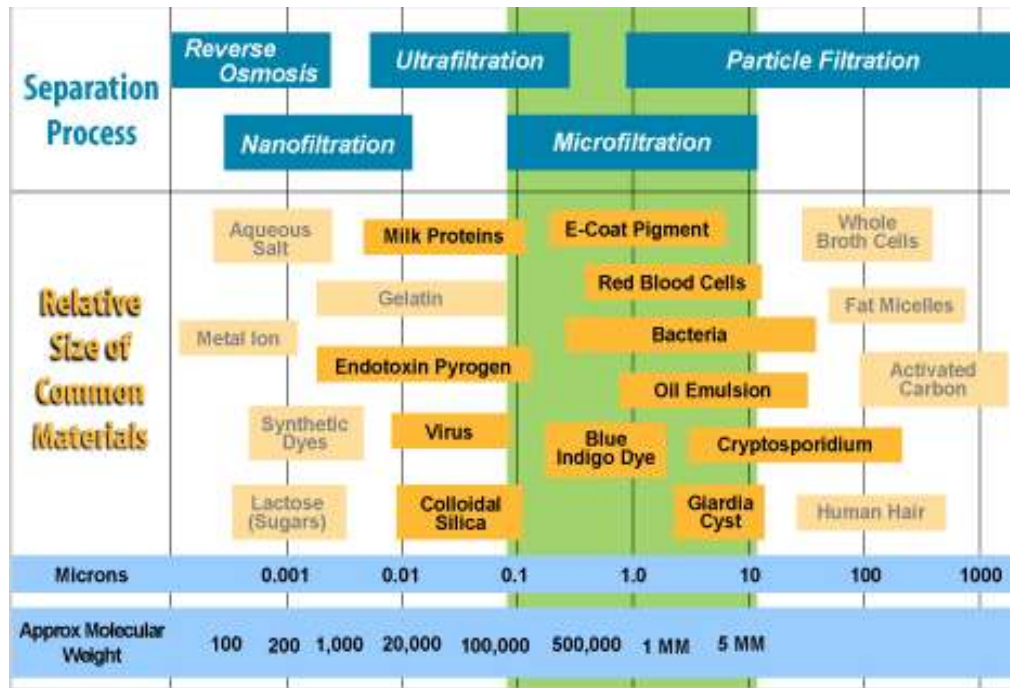


Figure 2.1: Spectrum diagram of membrane

### 2.2.1 Ultrafiltration (UF)

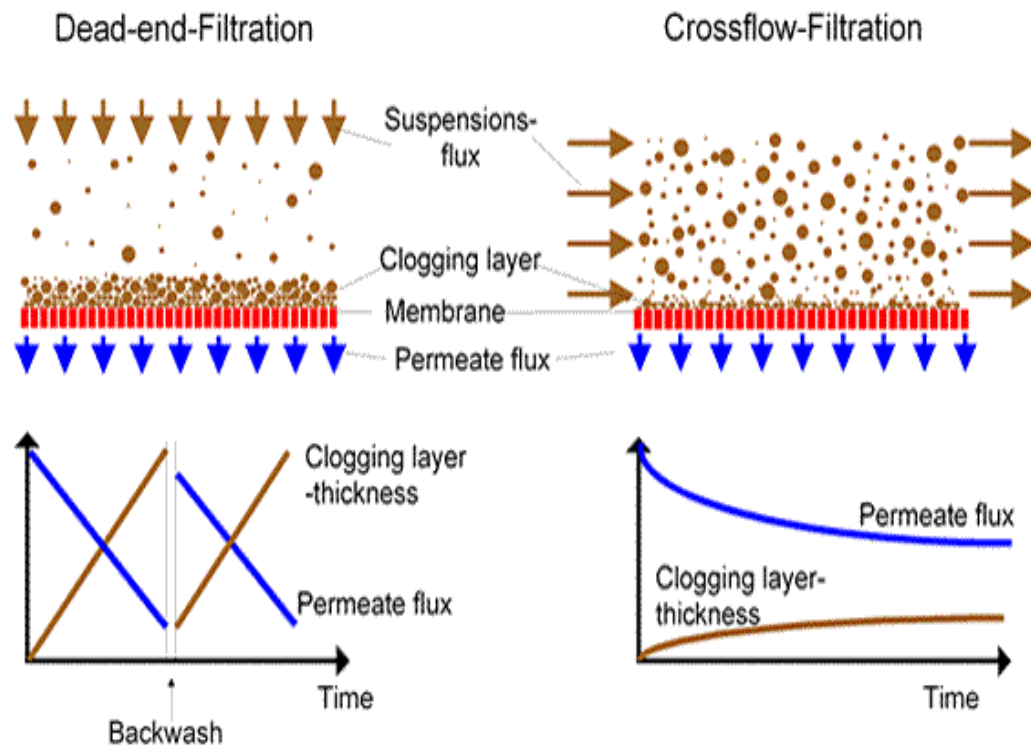
Ultrafiltration is a separation process using membranes with pore sizes in the range of 0.1 to 0.001 micron as be seeing in Figure 2.1. Typically, ultrafiltration will remove high molecular-weight substances, colloidal materials, and organic and inorganic polymeric molecules. Low molecular-weight organics and ions such as sodium, calcium, magnesium chloride, and sulfate are not removed. Because only high-molecular weight species are removed, the osmotic pressure differential across the membrane surface is negligible. Low applied pressures are therefore sufficient to achieve high flux rates from an ultrafiltration membrane. UF also use in whey treatment to recover cheese manufacturing byproduct (M.S. Yorgun *et al.* 2007).

### **2.2.2 Microfiltration**

Microfiltration is a filtration process which removes contaminants or undesired substances from a fluid (liquid & gas) by passage through a microporous membrane. A typical microfiltration membrane pore size range is 0.1 to 10 micrometres ( $\mu\text{m}$ ). Microfiltration is not fundamentally different from reverse osmosis, ultrafiltration or nanofiltration, except in terms of the size of the molecules it retains. The use of microfiltration not only standardize of protein content but also the casein to total protein ratio (Johnson & Lucy, 2006). Developed by Professor Sigmond University of Goettingen, Germany, in 1935, membrane filters were first commercially produced by Sartorius GmbH a few years later. Membrane filters found immediate application in the field of microbiology and in particular in assessment of safe drinking water. Further development of microfilters in the mid 1970's lead by the FDA's requirement for non-fibre releasing filters to be used in the production of injectable solutions. Microporous membranes are used by the micro-electronics industry as an integral part of water production. Membrane filters are widely used in biotechnology and food and beverage applications where sterile product is required (Yves Pouliot, 2008).

### **2.3 Mode of operation**

Membrane flow system can be divided by two systems, dead end filtration and cross flow filtration or tangential flow filtration. For many reason, the cross flow filtration is most widely used in dairy product industry due to the advantages of the system. Figure 2.2 shows that the differences of dead end and cross flow filtration. In dead end filtration, clogging layer rapidly increase and need backwash frequently compare to cross flow filtration.



**Figure 2.2** Dead end filtration Vs Cross flow filtration

Both normal flow and cross flow technologies purify bioprocess solutions by removing contaminants with a fixed porous medium, yet each format has unique advantages. Generally, normal flow filters (NFF) or dead end filtration are used where clarification and/or bio-burden reduction is desired in relatively low solid streams, for protecting or enhancing downstream operations, or when final polishing is required to achieve sterility. Cross flow filters (CFF) are best suited for higher solids, more viscous feed solutions, and/or where concentration or purification of cells or target species is desired.

### 2.3.1 Cross flow filtration

In cross-flow filtration, a feed liquor to be filtered flows through an overflow channel, whereby the feed is directed against the surface of the filter element tangentially. The filter splits the feed into a concentrate (retentate) and a filtrate