# ENZYMATIC EXTRACTION OF *FICUS* DELTOIDEA FOR BETTER ENHANCEMENT OF EXTRACTED YIELD

HAZWANI BINTI SIDEK HARON

BACHELOR OF CHEMICAL ENGINEERING UNIVERSITI MALAYSIA PAHANG

©HAZWANI SIDEK HARON (2014)



# **Thesis Access Form**

No	Location				
Author :					
Title :					
Status of acce	ss OPEN / RESTRICTED / CONFI	DENTIAL			
Moratorium p	Moratorium period: years, ending /200				
Conditions of	access proved by (CAPITALS): DI	R AHMAD ZIAD	BIN SULAIMAN		
Supervisor (S	ignature)				
Faculty:					
Author's Decl	aration: I agree the following condu	itions:			
OPEN access work shall be made available (in the University and externally) and reproduced as necessary at the discretion of the University Librarian or Head of Department. It may also be copied by the British Library in microfilm or other form for supply to requesting libraries or individuals, subject to an indication of intended use for non-publishing purposes in the following form, placed on the copy and on any covering document or label.					
The statement	itself shall apply to <b>ALL</b> copies:				
This copy has quotation fro	s been supplied on the understand m the thesis may be published wi	ling that it is cop thout proper ack	yright material and that no mowledgement.		
<b>Restricted/confidential work:</b> All access and any photocopying shall be strictly subject to written permission from the University Head of Department and any external sponsor, if any.					
Author's signatureDate:					
users declaration: for signature during any Moratorium period (Not Open work): <i>I undertake to uphold the above conditions:</i>					
Date	Name (CAPITALS)	Signature	Address		

# ENZYMATIC EXTRACTION OF *FICUS DELTOIDEA* FOR BETTER ENHANCEMENT OF EXTRACTED YIELD

# HAZWANI BINTI SIDEK HARON

Thesis submitted in partial fulfilment of the requirements for the award of the degree of Bachelor of Chemical Engineering

#### Faculty of Chemical & Natural Resources Engineering UNIVERSITI MALAYSIA PAHANG

JUNE 2014

©HAZWANI SIDEK HARON (2014)

#### SUPERVISOR'S DECLARATION

We hereby declare that we have checked this thesis and in our opinion, this thesis is adequate in terms of scope and quality for the award of the degree of Bachelor of Chemical Engineering.

Signature	:
Name of main supervisor	: DR. AHMAD ZIAD BIN SULAIMAN
Position	: SENIOR LECTURER
Date	:

#### **STUDENT'S DECLARATION**

I hereby declare that the work in this thesis is my own except for quotations and summaries which have been duly acknowledged. The thesis has not been accepted for any degree and is not concurrently submitted for award of other degree.

Signature:Name: HAZWANI BINTI SIDEK HARONID Number: KA10031Date: JUNE 2014

# Dedication

To my parents for their support

## ACKNOWLEDGEMENT

I would like to express my gratitude and sincere appreciation to my supervisor, Dr. Ahmad Ziad Bin Sulaiman for his guidance, support and advice throughout the preparation and realisation of this project.

My gratitude also goes to my family and friends who gives their support and help me to finish my project.

Finally, above all, thanks to Almighty God for giving me a good health, strength and perseverance to complete this project.

#### ABSTRACT

This thesis reported the enzymatic extraction of *Ficus deltoidea* (mas cotek) for better enhancement of extracted yield. F. deltoidea from Moraceae family which also known as "mas cotek", "serapat angin" and "telinga beruk" in Malaysia, "tabat barito" in Indonesia and "kangkalibang" in Africa. This plant has many used in traditional medication such as to help uterus contraction after deliver, improve blood circulation, reducing cholesterol, treat diabetes and also can use as aphrodisiac. Previous research had shown that there were several active compounds in F.deltoidea which are flavonoids, phenol, tannis, riterpenoids, isovitexin, vitexin, proanthocyanidins, flavan-3ol monomers and flavones glycosides. Since the uses of active compound of this plant have been known and have the high demand, there are several extraction technique have been research. However, the ideal technique is not reported yet. Hence, this work aims to study the most ideal extraction technique of F. deltoidea. The F. deltoidea ground leaves are added into the water to form suspension. The ground leaves are extracted by using two methods which is hydro distillation which is extraction by using water only and enzyme cellulase extraction which is the extraction wit enzyme assist. For the first enzymatic extraction, the different concentrations of enzyme cellulase but same temperature which is  $50^{\circ}$ C are used to get the optimum concentration. For the second enzymatic extraction used different temperature from the optimum concentration. The suspension is separated to oil-rich and emulsion phase by using centrifuge. The extraction is withdrawn by using micropipette. Then, the extracted active compound is analyzed by using high performance liquid chromatography (HPLC) to identifying and purifying the active compounds of the mixture. The isovitexin is used as the standard which is the active compound that will be detected by HPLC. According to the HPLC analysis, the retention time of isovitexin detected is around 13 min to 14 min for the standard solution. The optimum concentration for enzyme extraction is 1 mg/L. The yield of the enzymatic assist is higher than the control extraction which is without enzyme.

#### ABSTRAK

Tesis ini melaporkan tentang pengekstrakan Ficus deltoidea (mas cotek) dengan menggunakan enzim bagi meningkatkan lagi hasil diekstrak. F. deltoidea tergolong dari kumpulan Moraceae yang juga dikenali sebagai "mas cotek", "serapat angin" dan "telinga beruk" di Malaysia, "tabat barito" di Indonesia dan "kangkalibang" di Afrika. Tumbuhan ini banyak digunakan dalam ubat-ubatan tradisional seperti membantu pengecutan rahim selepas bersalin, meningkatkan peredaran darah, mengurangkan tahap kolesterol, merawat kencing manis dan juga boleh digunakan sebagai afrodisiak. Kajian sebelum ini telah menunjukkan bahawa terdapat beberapa sebatian aktif yang terkandung di dalam F. deltoidea iaitu flavonoid, fenol, tanis, riterpenoids, isovitexin, vitexin, proanthocyanidins, monomer flavan-3-ol dan flavon glikosida. Sejak kegunaan sebatian aktif tumbuhan ini diketahui ramai disamping mempunyai permintaan yang tinggi, beberapa penyelidikan tentang pengekstrakan telah dijalankan. Walau bagaimanapun, teknik penegkstrakan yang ideal belum dilaporkan lagi. Oleh itu, kajian ini bertujuan untuk mengkaji teknik pengekstrakan yang paling sesuai untuk mengekstrak daun F. deltoidea. Daun F. deltoidea yang telah dikeringkan dan telah dikisar dimasukkan kedalam air untuk menghasilkan campuran. Daun ini diekstrak dengan menggunakan dua kaedah iaitu hydro-penyulingan iaitu pengekstrakan yang menggunakan air sahaja dan pengekstrakan menggunakan bantuan enzim. Pengekstrakan menggunakan enzim yang pertama ialah menggunakan kepekatan enzim yang berbeza tetapi suhu yang sama iaitu  $50^{\circ}$ C dan mendapatkan kepekatan yang optimum. Yang keduanya ialah menggunakan kepekatan enzim yang optimum tetapi suhu yang berlainan. Campuran dipisahkan menggunakan centrifuge. Pengekstrakan itu diambil menggunakan micropippete. Sebatian aktif yang diekstrak kemudiannya dianalisis menggunakan kromatografi cecair prestasi tinggi (HPLC) untuk mengenal pasti sebatian aktif. Isovitexin digunakan sebagai standard yang merupakan sebatian aktif yang akan dikesan oleh HPLC. Menurut analisi HPLC, masa untuk mengesan isovitexin adalah dalam sekitar 13 min ke 14 min. kepekatan optimum untuk pengekstrakan adalah 1 mg/L. pengekstrakan menggunakan bantuan enzim adalah lebih berkesan daripada tanpa bantuan enzim.

# TABLE OF CONTENTS

SUPERVISOR'S DECLARATION	IV
STUDENT'S DECLARATION	V
Dedication	VI
ACKNOWLEDGEMENT	VII
ABSTRACT	VIII
ABSTRAK	IX
TABLE OF CONTENTS	X
LIST OF FIGURES	XII
LIST OF TABLES	XIII
LIST OF ABBREVIATIONS	
LIST OF ABBREVIATIONS	XV
1 INTRODUCTION	
1.1 Background	
1.2 Problem statement	
1.3 Objectives	
1.4 Scope of this research	4
1.5 Organisation of this thesis	4
	-
2 LITERATURE REVIEW	6
2.1 Overview	
2.2 Ficus Deltoidea	6
2.2.1 <i>History, habitat and industrial development</i>	6
2.2.2 Benefits of Ficus deltoidea	
2.2.3 Traditional uses of F. deltoidea	
2.3 Antioxidants	
2.4 Extraction of active compound	
	10
2.4.1 water extraction	
2.4.2 Enzyme assisted extraction	
2.5 Summary	
3 MATERIALS AND METHODS	
3.1 Overview	
3.2 Introduction	
3.3 Chemicals	
3.4 Ficus deltoidea leaves	
3.5 Hydrodistillation extraction	
3.6 Enzymatic extraction	
3.7 HPLC analysis	
3.8 Summary	
	10
4 RESULT AND DISCUSSION	
4.1 Stanuaru Canoration	
4.2 Engumetic extraction	
4.5 Enzymatic extraction extraction and enzymatic extraction	
4.4 Comparison of water extraction and enzymatic extraction	
4.5 Summary	
5 CONCLUSION	
5.1 Conclusion	

5.2	Recommendation	24
REFRE	NCES	.25
APPEN	DICES	.27

# LIST OF FIGURES

Figure 1-1: Global Market Value of Herbal Industry according to year 1
Figure 1-2: <i>Ficus deltoidea</i> tree 2
Figure 2-1: Comparison between male and female leaves of <i>Ficus deltoidea</i> 7
Figure 2-2: Malaysian natural product industry in 2007
Figure 2-3: <i>F. deltoidea</i> based product
Figure 2-4: Basic structure of flavonoids 11
Figure 3-1: Process flow 14
Figure 3-2: Hydrodistillation process 15
Figure 3-3: Incubator shaker 16
Figure 3-4: High performance liquid chromatography (HPLC) 17
Figure 4-1: Calibration curve of isovitexin 18
Figure 4-2: Percentage of yield of isovitexin for hydrodistillation process with different duration
Figure 4-3: Percentage of yield of isovitexin for different concentration of enzyme 21
Figure 4-4: Percentage of yield of isovitexin different temperature for enzymatic extraction
Figure 4-5: Comparison of percentage of yield of water extraction and enzyme extraction

# **LIST OF TABLES**

Table 2-1: Chemical and nutritional constituents of F. deltoidea	. 9
Table 4-1: Peak area and yield percentage for different water extraction duration	19
Table 4-2: Peak area and percentage of yield for enzymatic extraction	20

# LIST OF ABBREVIATIONS

F.d	ficus deltoidea
t <sub>R</sub>	retention time
V	volume
v/v	volume per volume
W	weight of sample
w/v	weight per volume
у	yield

# LIST OF ABBREVIATIONS

Acetonitrile ACN

HPLC

High performance liquid chromatography Malaysia Agriculture Research and Development Institute MARDI

Revolutions per minutes rpm

# **1 INTRODUCTION**

#### 1.1 Background

Medicinal and herbal plants are very important nowadays and also have been used since a long time ago in Malaysia. The demands for the herbal plant are always increase due to the increasing in the usage. Basically, there is high antioxidant activity that contained in herbal and vegetable plant (Asmah *et al.*, 2003). It is very important in inhibiting and scavenging radicals and also provide shield to humans against treat such as infections and degenerative diseases. The effective antioxidants from natural sources are strongly needed as alternatives to synthetic food additives in order to prevent deterioration of foods, drugs, and cosmetics for their role in the maintenance and improvement of health and wellness. In this research, there are also antioxidant activities in the herbal plant which is Mas Cotek.



Global Market Value of Herbal Industry

Figure 1-1: Global Market Value of Herbal Industry according to year

Mas Cotek or known as *Ficus deltoidea* in the scientific are from Moraceae family (Mahmood *et al*, 2010). The name of Mas Cotek is well known in Malaysia and there also several others name such as '*serapat angin*' and '*telinga beruk*'. While, in Indonesia they known as '*tabat barito*' and in Africa known as '*kangkalibang*'. Unlike the *Labisia pumilla* (kacip Fatimah) which specifically used for woman and *Eurycomo longifolia* (tongkat ali) which specifically used for man, *F. deltoidea* didn't have the specification of uses whether for both man and woman (Hakiman and Maziah, 2009).



Figure 1-2: Ficus deltoidea tree

According to the research by Malaysia Agriculture Research and Development Institute (MARDI), there are several active compounds that contain in *F. deltoidea* and the main four are flavonoid, tannis, triterpenoids, and phenols. The herbal plant can be used in many industries such as flavor and fragrance, pharmaceuticals and herbal remedies. Since there is strong antioxidant that contain in *F.deltoidea* which is in phenols, it can be used in all of those industry but mostly *F.deltoidea* are used in pharmaceuticals and herbal remedies.

Plant cell wall consists of cellulose, hemi-cellulose and pectin which act as the barrier for the intracellular substance to be release. Degradation of the cell wall is the basic step to release the active compounds. Usually, the active compounds are extracted by using extraction process which is transferring the solutes from a solid medium to a solvent. There are various methods that been used to assist the extraction of the active compound from plant such as ultrasonication, supercritical fluids, microwave, membrane adsorption and molecular imprinting. However, there still some active compound that still retain in the polysaccharide-lignin network by hydrogen or hydrophobic bonding, which cannot be easily extracted by solvent. Thus, Chen *et al.* (2011) suggests that enzyme-assisted extraction, in which the extraction is improved the cell wall degradation. This method also can increase the solubility of the target compounds in the extractant.

Extraction of natural functional compounds from plant by using enzyme-assisted is widely investigated in recent years. The advantages for this method are can provide high efficiency, easily operated, and environment friendly. The enzymes breaking down the cell walls and the membrane of the plant and this will enable the extraction of active compound to better release. This method also can reduce the usage of solvent and can lower the energy consumption. Thus, enzymatic-assisted extraction has been represented as an alternative method to extract the natural product.

In past, the conventional extraction method of *F. deltoidea* required long extraction time and high energy consumption with low extracted yield. Therefore, to increase the extracted yield, the ideal extraction method should be introduced.

#### 1.2 Problem statement

In increased of health problem and rising health concern in modern world has promoted various medicinal and natural products. In Malaysia especially, there are many causes of death and the major cases are heart attack and the major sickness for Malaysian are diabetes. The active compound that contain in *Ficus deltoidea* can be used to reduce the cholesterol in the body and also will increase the blood circulate. Thus, the demand for the *F. deltoidea* extraction increased due to increasing the health concern in human.

The active compound in *F. deltoidea* can be better release by breaking down the cell wall of the plant. *F. deltoidea* which is the lignocellulosic material that contain high ligning and hemicellulosic that cannot be break easily. Thus, the enzyme is used to assist the solvent to break down the cell wall and the membrane of the plant.

Besides, by using enzymatic extraction will reduce the cost of the process and the time to produce the oil yield. The other method of extraction such as using water and solvent extraction need more process cost and also takes the long extraction time to produce the oil.

## 1.3 Objectives

The following are the objectives of this research:

- $\circ$  To extract active compound of *F. deltoidea* using enzymatic extraction as the main method.
- To compare the result which in process time, yield and extraction productivity for extraction with other extraction methods.

## 1.4 Scope of this research

In order to achieve the objective, the following scopes have been identified and to be applied:

- i) Optimization study of enzymes for Ficus deltoidea extraction.
- ii) Use enzymatic extraction to extract active compound of Ficus deltoidea.
- iii) Analyze the active compound constituents from the method using HPLC.
- iv) Compare the yield of active compound of F. deltoidea from the method.

## 1.5 Organisation of this thesis

The structure of the reminder of the thesis is outlined as follow:

In chapter 2, it is involves about the overall explanation about the *Ficus deltoidea* including history, habitat, tree and industrial development. Besides, the uses and the active compound that contain in *F. deltoidea* also mentioned in this chapter. More than that, the different types of methods of extraction such as water extraction and enzymatic extraction also mentions. Everything about all these and the statement the previous research are also presented.

Chapter 3 gives the explanation about the method used in this research. All the methods involve such as preparation of raw material and extraction process which is using water extraction and enzymatic extraction. Besides, analytical methods are also provided in this chapter. In addition, all the parameters related in this research are also described as it is necessary.

Chapter 4 will be present about the result got from this research. The result gained from this research is explained.

Chapter 5 draws together a summary of the thesis and outlines the recommendation for the future work which might be derived from the model developed in this work.

# **2** LITERATURE REVIEW

#### 2.1 Overview

This chapter are about the finding of the related article from previous researches about the history, habitat, industrial development, usage, and active compound that contain in *Ficus deltoidea*. This chapter also tell about the extraction technique that be used by previous researches.

#### 2.2 Ficus Deltoidea

#### 2.2.1 History, habitat and industrial development

*Ficus deltoidea* or known as Mas Cotek is belongs to Moraceae family. This plants are originated in Asia Minor and can be found throughout Mediterranean, India subcontinent, Latin America, Texas, Southern California, until the far east such as Malesian tropical rain forest which about 750 species (Nashriyah *et al.* 2012). It is one of the largest genres of flowering plants with six traditional subgenera that recognized based on morphology and distribution. Due to the diverse natural habitats, the *Ficus deltoidea* plant successful spreading to the worldwide. According to Nashiriyah *et al.* (2012), *Ficus deltoidea* mistletoe fig is a native of Peninsular Malaysia and been introduced elsewhere which is in Malesia that include Thailand, Indonesia and Malaysia.

According to Mahmood *et al.* (2010), the vernacular name for *F. deltoidea* are '*mas cotek*', '*serapat angin*', '*telinga beruk*' and many more in Malaysia, while in Indonesia known as '*tabat barito*', and in Africa known as '*kangkalibang*'. There are fine spots with gold colour on the surface of each leaves and the vernacular name of Mas Cotek is given in Peninsular Malaysia because of it.

*Ficus deltoidea* plant is commonly found in Peninsular Malaysia which is generally as an epiphyte; depend on another bigger or taller plant (usually 20 feet from the land) to reach their own sunlight resource, in lowlands and mountains but also as a terrestrial shrub on sandy shores, mountain tops and in bogs (Starr *et al.*, 2003). The majority of varieties grow at below 1200 m altitude, however *F.deltoidea* var. *intermedia* can be found in the higher mountain areas above the dipterocarp forest. *Ficus deltoidea* var.

*angustifolia* grows in places by the streams or riversides. In Borneo, the var. *motleyana* is found in the coastal, peat-swamp and sandy heath forest.

*Ficus deltoidea* have two types namely the male and female. The male and female plant can be differentiated by their morphology. The male plant has smaller and elongated leaves size whereas the female plant has larger and more rounded leaves (Naquib & Vivi, 2013). The top of the leaf surface is shiny green in colour while golden in color (rust-red to olive brown) with black dots under the leaf (Starr *et al.*, 2003). Normally, the flowers are found freely in paired or single for each plant. The flower is built in receptacle where the fruits and flowers are appearing inside. The colour of flowers changed from green to red when it became matured.



Figure 2-1: Comparison between male and female leaves of Ficus deltoidea

The herbal extractions have many uses. Their uses can be channeled into four product groups which are herbal remedies, pharmaceuticals, flavor and fragrance and bio pesticides. According to Rasadah & Li (2008), herbal medicinal products have taken on increasing significance as regards medical and economic importance over the last two decades, and have registered as strong comeback in developed countries in the USA, the European Union (EU), Australia and Canada. The global expansion of the herbal industry has a significant impact on the Malaysia market. In Malaysia, the herbal market which was worth RM3.8 billion (USD 1.03 billion) in 2006, is anticipated to grow with an annual growth rate of between 15 and 20 percent.



Figure 2-2: Malaysian natural product industry in 2007

There is a focus on herbs based on traditional systems of medicine and remedies based on folk knowledge due to their access, low cost and faith of the people. Natural products from medicinal and aromatic plants can be applied to a wide range of items such as herbal and traditional medicine, food and beverages, bio-pharmaceuticals, flavours and fragrances, cosmetics and toiletries, dye, detergents, bio-pesticides and special chemicals. Considering their importance, quality is essence and guidelines for quality control at each step of production are critical to the sustainability of the product and its wide acceptance. For *Ficus deltoidea*, it commonly used in herbal and traditional medicine, bio-pharmaceuticals, and cosmetics and toiletries.



Figure 2-3: F. deltoidea based product

## 2.2.2 Benefits of Ficus deltoidea

According to the research that been made by Malaysia Agriculture Research and Development Institute (MARDI), *F. deltoidea* contains antioxidant chemicals such as saponins, flavonoids, tannins, polyphenols, triterphenoids, and proanthocyanins. The analyses of the chemical and nutritional constituents are summarized in Table 2.1 below.

Chemical	Nutritional constituents		
Naringin	An anti-inflammatory for small injury.		
Flavonoid	Acts an antioxidant which affects better blood flow, antiviral		
	and as heart and estrogen hormones protector.		
Tannins Protect the skin and acts as an anti-wrinkle, diarrhea and p			
	the damaged tissue.		
Polyphenols	Young leaves contain high polyphenol that acts as anti-		
	inflammatory and anti-septic.		
Triterpenoids	Important during the nutrient absorption process.		
Proanthocyanins	Protect tissues from damage and it also acts as anti-toxic.		

Table 2-1: Chemical and nutritional constituents of F. deltoidea

Other than that, a researcher from Universiti Malaya reported that a right combination of *Ficus deltoidea* species is capable to neutralize body system. Therefore, *F. deltoidea* can reduce high blood pressure problem, diabetes, gout, migraine, ulcer and other diseases.

According to Adam et al. (2009), F. deltoidea has been claimed to have antidiabetic properties and has been used as remedy to treat the diabetes. This theory is based on the ethnobothancal approaches and the scientific evidence to confirm its efficiency however is still lacking. Zainah et al. (2010) have done the research on antihyperglycemic and glucose tolerance activity of *Ficus dektoidea* on diabetic rats by using ethanolic extraction. In the paper, 70% ethanol are used to extract ground F. deltoidea leaves for 3 days at room temperature. The result show that the F. deltoidea extract can reduce the blood glucose level in the diabetes rats. As the result, while using the 500 mg/kg/b.w of extract F. deltoidea the glucose level in the blood decrease from  $23.81\pm0.98$  mmol/L to 21.21±2.48 mmol/L for 2 hours. Adam et al. (2010) also do the research on the reducing blood glucose and the result shows that the blood glucose reduced at the certain time. Besides, according to Mahmood et al. (2010), Ficus deltoidea extract also can be used to heal the wound. The group rats are taken and treat by using sterile deionized water, blank placebo and placebo containing F. deltoidea. From the result, wound treat with aqueous extract F. deltoidea showed considerable signs of dermal healing and significantly (p<0.05) healed earlier compared to wounds that treat with sterile deionized water or blank placebo.

#### 2.2.3 Traditional uses of F. deltoidea

According to Malaysia natives, the *Ficus deltoidea* plant is usually used by the women that just give birth, regain energy, repair blood flows and associated problem and also can be used as herbal drinking for healthy and beauty. For almost traditional medicine practice, *F. deltoidea* is also capable of preventing and curing certain diseases if it is used rightly. For example, *F. deltoidea* can be used to cure diabetes (Adam *et al.*, 2009), high blood pressure and low libido energy.

According to the natives also, these herbs must be boiled with the water for 45 minutes until all the extracts whether from the stems, leaves, fruits, twigs or roots are obtained. Then the extracts that obtain from boiling are filtered. It can be stored in refrigerator for longer use but must be consumed within 3 days. This drink is suitable for everyone with no limits of age and genders; however it is not suitable for pregnant women.

#### 2.3 Antioxidants

The antioxidant activity and its potency in certain plant depend on the existence of various compounds found in that plant. Some of phenolic compounds (anthocyanidin, catechines, flavones, flavonols and isoflavones), tannins (ellagic acid, gallic acid), phenyl isopropanoids (cafeicacis, coumaric acid, ferulic acid), lignans, catchol and others are antioxidants. The antioxidative and radical scavenging activities of this antioxidant are well studied (Emami *et al.*, 2007).



Flavanone

Figure 2-4: Basic structure of flavonoids

Antioxidant inadequacy is associated with oxidative damage or kills to DNA of the germ line as well as somatic cells. In addition to the reduction to the reduction in cancer rates, high consumption of antioxidants is related to reduce risk of cardiovascular disease including heart attack and coronary heart disease. Although antioxidant supplementation is widely used in order to avoid the development of cancer, it has been suggested that antioxidants may interfere with cancer treatments. High doses of water-soluble antioxidants such as ascorbic and very high doses of some antioxidants are found to be toxic. This may have adverse long-term effects.

#### 2.4 Extraction of active compound

Plant cell wall consists of cellulose, hemi-cellulose and pectin which act as the barrier for the intracellular substance to be release. Degradation of the cell wall is the basic step to release the active compounds. Usually, the active compounds are extracted by using extraction process which is transferring the solutes from a solid medium to a solvent. There are various methods that been used to assist the extraction of the active compound from plant such as ultrasonication, supercritical fluids, microwave, membrane adsorption and molecular imprinting. However, there still some active compound that still retain in the polysaccharide-lignin network by hydrogen or hydrophobic bonding, which cannot be easily extracted by solvent. Thus, Chen *et al.* (2011) suggests that enzyme-assisted extraction, in which the extraction is improved the cell wall degradation. This method also can increase the solubility of the target compounds in the extractant.

#### 2.4.1 Water extraction

The water extraction method is a method that commonly used in chemistry for general extracts of interest extraction. This process used a combination of high pressure water for agitation and hot water to increase reaction rate. The process generally involves taking a mixture of substances; dissolve in warm water and then cooling the mixture. The insoluble compounds precipitate out of the water, while the soluble ones stay dissolve. The solution can be separated by filtration or decantation. This process works by exploiting the differences in solubility (with respect to temperature) of varying substances.

Mostly of the report are used water and solvent to extract the bioactive compound. Misbah *et al.* (2013) and Adam *et al.* (2010) used distilled water to extract the bioactive compounds. While Farsi *et al.* (2011) used solvent to extract the bioactive compound. Some of active compound generally have poor solubility in mild solvents such as ethanol-water solution. Therefore, to increase the extracted yield, the ideal extraction method should be introduced.

#### 2.4.2 Enzyme assisted extraction

Extraction of natural functional compounds from plant by using enzyme-assisted is widely investigated in recent years. The advantages for this method are can provide high efficiency, easily operated, and environment friendly. The enzymes breaking down the cell walls and the membrane of the plant and this will enable the extraction of active compound to better release. This method also can reduce the usage of solvent and can lower the energy consumption. Thus, enzymatic-assisted extraction has been represented as an alternative method to extract the natural product.

There are several reports that study on enzyme-assisted extraction for bioactive compound. Chen *et al.* (2011) has been study on enzyme-assisted extraction of flavonoids from *Ginko biloba* leaves. From the study, found that enzyme was the effectively hydrolyze the active compound. Although the enzyme-assisted approach has largely improved the extraction rate, but the solvent such water still needed to perform the extraction after enzymatic treatment.

#### 2.5 Summary

Nevertheless, regarding toxicological limitations, there is a clear trend in the industry to substitute these organic solvent for alternative nontoxic solvents, with the first option by using water or alcohols. Therefore, large amount of high concentration ethanol-water solution is often used. However, this will leads to high production cost, making it desirable to increase the solubility of active compound without affecting their physiological activities.

# **3 MATERIALS AND METHODS**

#### 3.1 Overview

In this chapter, we are discussing more detail about the method to conduct in this research on enzymatic extraction of *Ficus deltoidea*. The procedures include in this research are, preparing the *F. deltoidea* powder and extraction process by using water extraction and enzyme extraction. There are also including the analytical method in which the method to determine the yield of the extracted essential oil from *F. deltoidea* leaves.

## 3.2 Introduction





The first step of the experiment is preparing the *Ficus deltoidea* powder which is the plant leaves need to be dried at certain temperature and then grind to form the powder. *Ficus deltoidea* need to be dried to remove all the moisture content in the leaves.

After that, the *Ficus deltoidea* powder must be immerse into the solvent to do the extraction process. In this research, there are two types of extraction process which is by using hydro distillation extraction process and enzymatic assisted extraction process. There is another method of extraction that using chemical solvent that known as solvent extraction like methanolic and ethanolic extraction. Hydro distillation is used for this experiment because of its environmental friendly and same goes to enzymatic extraction. There is not chemical as a solvent involve in the extraction of active compound. Other than that, both of these methods are used a lower production cost and very simple extraction method.

After the production of essential oil, high performance liquid chromatography is used for the composition analysis. In this step, the data were collected and the yield will be calculated.

## 3.3 Chemicals

The phosphoric acid and acetonitrile are obtained from Sigma- Aldrich Malaysia. Enzymes which is cellulase from *Aspergillus niger* used in the study are also obtained from Sigma- Aldrich Malaysia. The technique cell free enzyme is conducted during the experiment.

# 3.4 Ficus deltoidea leaves

The original leaves of *F. deltoidea* are obtained from Sungai Tengi Selatan, Selangor, Malaysia. The leaves are dried in the oven at temperature  $45^{\circ}$ C and grind into the fine powder. Then, the powders are stored until experiment.

# 3.5 Hydrodistillation extraction

The powder from *F*. *feltoidea* leaves (100 g) will through hyrodistillation extraction process for 10 hour and the distillate be collected every 2 hours.



Figure 3-2: Hydrodistillation process

#### 3.6 Enzymatic extraction

10 g of ground *F. deltoidea* is immersed into the distilled water to form 100 ml suspension of *F.deltoidea*. The enzyme which is cellulase is added into the suspension. The suspensions were put in the incubator shaker with the speed and the temperature is fixed at 200 rpm and  $50^{\circ}$ C respectively and the cellulase is allowed to act for 2 hour. After that, the suspension is centrifuged 3000 rpm for 15 minutes at  $20^{\circ}$ C to separate the oil-rich and emulsion phase. The oil-rich is withdrawn by using a micropipette. The quantities of enzyme used in the experiments evaluating the effects of the individual parameters are: cellulase: 0.5, 1, 1.5, 2, 2.5 mg/L.

By using the same method, 1 mg/L of cellulase is added into the suspension. The speed of the incubator shaker is fixed at 200 rpm but different temperature. The temperatures that used for enzymatic extraction are: 25, 35, 45, 50, 60 and 70  $^{\circ}$ C.



Figure 3-3: Incubator shaker

#### 3.7 HPLC analysis

*F. deltoidea* solution were dilute to 1:9 dilution factors with ultrapure water and then filtered with the syringe filter (0.22  $\mu$ m). Separations were carried out by high performance liquid chromatography (HPLC) analysis using C-18 column (Prodigy 5  $\mu$  ODS-3 100A). Filtered samples were injected into a column (250 x 4.60 mm). The column equilibrated in 0.1% v/v solvent A (phosphoric acid in ultrapure water) and 100% solvent B (acetonitrile). The flow rate was set at 1.0 ml/min and column

temperature was maintained at room temperature. The peak areas were determined at wavelength 335 nm.



Figure 3-4: High performance liquid chromatography (HPLC)

## 3.8 Summary

There were two methods of extraction that involve in this experiment which the extraction that used water only and extraction by using enzyme assist. For the enzyme assist, there were two parameter involve which concentration and temperature. Then, all the samples were analysed by using HPLC.

# **4 RESULT AND DISCUSSION**

#### 4.1 Standard Calibration

The active compound were identified by means of retention time and peak were quantified by comparing peak areas with the results of calibration series using pure native standards which is apigenin-6-C-glucoside (isovitexin) which obtained from Sigma-Aldrich. Isovitexin calibration standard was prepared by serial dilution using ultrapure water. There was linear response between the active compound and UV absorbance at 335 nm for all standards over the calibration rage studied as shown in Figure 4.1. The standard curves were constructed for the purpose of quantitation using a line fit force through zero and the correlation coefficient was  $r^2 \approx 0.98$  for isovitexin.



Figure 4-1: Calibration curve of isovitexin

#### 4.2 Hydrodistillation

Comparison of HPLC chromatogram was made between the standard and sample of the active compound to quantify the active compound in each sample. The peak corresponding to isovitexin appeard at retention times 13.7. HPLC method is a rapid and accurate qualitative and quantitative analysis of the active compound in F. *deltoidea*. The analysis time as approximately 20 minutes which is faster than previously reported which is 40 minutes in Maizatul et al. (2011). The yield of the active compound was calculated according to the following equation

yield, y (%) = 
$$\frac{\left[\text{concentration } \left(\frac{\text{mg}}{\text{L}}\right)\right] \times \text{volume (L)} \times \text{[distillation factor]}}{\text{weight of sample, w (mg)}} \times 100$$

In Table 4.1 shows the peak area that obtained from HPLC analysis for hydrodistillation process for different duration. Yield of isovitexin were calculated by using the equation and the data are shown in Table 4.1

Table 4-1: Peak area and yield percentage for different water extraction duration

duration, hr	peak area	yield (%)
2	115.48175	0.151
4	121.40926	0.155
6	113.99442	0.150
8	142.56859	0.171
10	101.32638	0.140

The peak area of the isovitexin which detected at  $t_R$  is around 13.7 minutes has the highest value at duration 8h. When the peak areas were increase, the percentages of yield also increase. The percentage of yield of isovitexin against duration can be concluding in the Figure 4.2.



Figure 4-2: Percentage of yield of isovitexin for hydrodistillation process with different duration

From the figure, the yield was increase until 8h duration and then decrease at the 10h duration. At 8h duration has the highest percentage of yield which is the isovitexin content is higher. At 8h duration, the cell wall of the *F. deltoidea* leaves had been

degraded and more active compound released. Since the active compound are almost released at the 8h duration, thus at 10h duration the active compound reduced.

#### 4.3 Enzymatic extraction

There many parameters that can be consider in enzymatic extraction which are enzyme concentration, temperature, pH and retention time. This research is to study the effect of the enzyme concentration and the temperature toward the active compound productions. In Table 4.2 shows the parameters which are enzyme concentration and temperature and the peak area which get from HPLC analysis. The peak area was highest at concentration 1 mg/L which is 167.76. While for the temperature, the highest peak area was detected at 25<sup>o</sup>C which is 306.25 and the lowest peak area was at 50<sup>o</sup>C which is 167.76. The percentage of yield can be concluding in Figure 4.3 for enzyme concentration and Figure 4.4 for temperature different.

Parameter		peak area	yield (%)
Concentration	0.5	138.46167	0.168
(mg/L)	1	167.76205	0.189
	1.5	164.08655	0.187
	2	151.70726	0.178
	2.5	120.98145	0.155
Temperature	25	306.24799	0.291
(°C)	35	283.93219	0.275
	45	288.16373	0.278
	50	167.76205	0.189
	60	244.04507	0.245
	70	189.29407	0.205

Table 4-2: Peak area and percentage of yield for enzymatic extraction

Enzyme-assisted extraction of natural functional compounds from plants was widely investigated recent years for its advantages in easy operation, high efficiency and environment friendly. It also can reduce the extraction time. Enzyme will degrade the cell wall constituents of *F. deltoidea* leaves faster to improve the release of intracellular content.



Figure 4-3: Percentage of yield of isovitexin for different concentration of enzyme

The effect of enzyme concentration on the yield of isovitexin was demonstrated in Figure 4.3. The yield of isovitexin reached maximum when the enzyme concentration was raised to 1.0 mg/L. Indeed, the lowest yield of isovitexin was obtained when 2.5 mg/L of cellulase was used. In this study, the sole variable was the concentration of cellulase enzymes and the cell wall has become the limiting factor. As the enzyme concentration increased, more active sites were available for effective collisions in the formation of enzyme-substrate complex. Hence, the number of active compound isolated was also increased correspondingly. Since the tissues were readily attacked by the enzymes, an increasing enzyme concentration contributed to an increase in the penetration ability of enzymes through multilayer of tightly packed cell in *F. deltoidea* leaves. Therefore, an addition of enzymes per unit volume was unable to further increase the active compound. In contrast, a higher concentration of enzymes has negatively influenced the released of active compound. The reduction in the yield of isovitexin in excess enzyme concentration was probably due to over-digestion of active compound by the enzymes.



Figure 4-4: Percentage of yield of isovitexin different temperature for enzymatic extraction

At fixed concentration which was 1.0 mg/L, isovitexin yield increased with rising temperature as the rate of enzyme-catalysed reactions increased with increasing temperature. However, at high temperatures which at 50  $^{0}$ C, the yield decreased significantly as enzymes which are proteins, became denatured at higher temperatures. Since the enzyme was denatured at temperature 50 $^{0}$ C, there were no enzyme activities that involve at temperature 60 $^{0}$ C and 70 $^{0}$ C. The percentage of yield of isovitexin was increased at temperature 60 $^{0}$ C probably due to the water extraction.



## 4.4 Comparison of water extraction and enzymatic extraction

Figure 4-5: Comparison of percentage of yield of water extraction and enzyme extraction

The comparison of hydrodistillation extraction method and enzyme extraction method at duration 2h were shown in Figure 4.5. This figure shows that percentage of yield of active compound in enzymatic extraction was higher compared to hydrodistillation. Besides, the duration for the water only to degrade the cell wall of the *F. deltoidea* leaf was longer than enzyme assisted extraction. This shows that the enzyme assist extraction was more effective compare the water extraction.

#### 4.5 Summary

The optimum time for the hydrodistillation extraction of the isovitexin was 8h with yield 0.17% and the optimum cellulase concentration for enzymatic extraction was 1.0 mg/L with yield 0.19%.while for the temperature, at 50  $^{0}$ C the yield which 0.19% was contain the lowest active compound compare with the other temperature. Enzymatic extraction is more efficient method compare to the hydrodistillation method. The yield of the optimum concentration of the cellulase was higher compare to the yield of the hydrodistillation extraction.

# **5** CONCLUSION

#### 5.1 Conclusion

*Ficus deltoidea* which from moracheae family, has many uses especially in herbal and traditional medicine. There are many methods that can be used as assist to extract active compound for example ultrasonication, supercritical fluids, microwave, membrane adsorption and molecular imprinting. In this research, the comparisons of two methods which are hydrodistillation method and enzymatic extraction method have been made. The result shows that enzymatic extraction method is more effective compare to the hydrodistillation method. For enzymatic extraction, the optimum concentration of the enzyme used in extraction is 1.0 mg/L and the maximum temperature for enzyme to react is  $50^{\circ}$ C before the enzyme be denatured.

## 5.2 Recommendation

There are a few recommendations for the future study of this research. Firstly, the research should be further with different parameter such as pH, reaction duration, particles size of ground leaf of *Ficus deltoidea* and others. Besides, the research should be study for the mechanism of the degradation of cell wall of the *Ficus deltoidea*'s leaf.

## REFRENCES

- Adam, Z., Khamis, S., Ismail, A., Hamid, M., (2010). Inhibitory properties of Ficus deltoidea on α-glucosidase activity. *Research Journal of Medicinal Plant*. 4(2), 61-75.
- Adam, Z., Hamid, M., Ismail, A., Khamis, S., (2009). Effect of Ficus deltoidea on hepatic basal and insulin-stimulated glucose uptake. *Journal of Biological Sciences*. 9(8), 796-803.
- Alireza, F,. *et al.* (2007). Synthesys and antioxidant properties of substituted
   3-benzylidene-7-alkoxychroman-4-ones. *Bioorganic and Medicinal Chemistry Letters.* Vol 17(24), pg 6764-6769
- Badr, F. H. & Sitohy, M. Z., (1992). Optimizing conditions for enzymatic extraction of sunflower oil. Food Science Department and Biochemistry Department. Faculty of Agriculture, Zagazing Ubiversity. 43, 281-283
- Bateman, D.F., & Basham, H.G., (1976). Degradation of plant cell walls and menbranes by microbial enzymes. *Encyclopedia of Plant Physiology*. Vol. 4, pp 316-355
- Chen, S., Xing, X. H., Huang, J. J., Xu, M. S., (2011). Enzyme-assisted extraction of flavonoids from *Ginko biloba* leaves: Improvement effect of flavonol transglycosylation catalyzed by *Penicillium decumbebs* cellulose. *Enzyme and Microbial Technology*. 48, 100-105.
- Choo, C. Y., Sulong, N. Y., Man, F., Wong, T. W., (2012). Vitexin and isovitexin from the Leaves of Ficus deltoidea with in-vitvo α-glucosidase inhibition. *Journal of Ethnopharmacology*. 142, 776-781
- Farsi, E., et.al., (2011). Correlation between enzymes inhibitory effects and antioxidant activitiestandardized fractions of methanolic extract obtained from Ficus deltoidea leaves. *African Journal of Biotechnology*. Vol. 10(67), pp, 15184-15194.
- Gai, Q-Y., Jiao, J., Wei, F-Y., Meng, L., Wang, W., Zu, Y-G., Fu, Y-J., (2013).
   Enzyme-assisted aqueous extraction of oil from Forsythia suspense seed and its physicochemical property and antioxidant activity. *Industrial Corps and Productions*. 51,274-278
- Hakiman, M. & Maziah, M. (2009). Non-enzymatic and enzymatic antioxidant activities
  in aqueous extract of different *Ficus deltoidea accessions*. J. Medicinal Plant Res. 3(3): 120-131.
- Mahmood, A. A., Khaled, A. A., Faisal, M. A. L., Mazin, M., (2010). Role of Ficus deltoidea extract in the enhancement of wound healing in experimental rats. *Biomedical Research*. 21(3), 241-245
- Maizatul, H. O., William, M., Alan, C., (2011). Identification of Proanthocyanidin Dimers and Trimers, Flavone C-Glycosides, and Antioxidant in Ficus deltoidea, a Malaysian Herbal Tea. *Journal of Agricultural and Food Chemistry*. 59, 1363-1369

- Misbah, H., Azlina, A. A., Norhaniza, A., (2013). Antidiabetic and antioxidant properties of Ficus deltoidea fruit extracts and fractions. *BMC Complementary and Alternative Medicine*. 13, 118
- Murugananthan, G., & Pabbithi, S. C., (2012). Medicinal plant with potent antioxidant constituents. *International Journal Pharmaceutical Sciences and Research*. Vol 3, Issue 05, pp 1268-1273
- Nashiriyah, M., *et al.* (2012). Leaf morphological variations and heterophylly in Ficus deltoidea Jack (Moraceae). *Sains Malaysiana*. 41(5), 527-538
- Oh, M-J., Mariani, A.H., Sulaiman, N., Seo, Y-K., Mohamad, R.S., Park, C.S., (2011). Ficus deltoidea (Mas Cotek) extract exerted anti-melanogenic activity by preventing tyrosinase activity in vitro and by suppressing tyrosinase gene expression in B16F1 melanoma cells. Arch Dermatol Research. 303, 161-170
- Ong, S.L., Ling, A.P.K., Sobri, H., (2008). Establishment and optimization of growth parameters for cell suspension cultures of Ficus deltoidea.. *American-Eurasian Journal of Sustainable Agriculture*. 2(1), 38-49
- Ong, S.L., Ling, A.P.K., Poospooragi, R., Moosa, S., (2011). Production of flavonoid compounds in cell cultures of Ficus deltoidea as influenced by medium composition. *Int. Journal Medical Aroma Plant*. Vol. 1(2). Pp. 62-74.
- Rosenthal, A., Pyle, D.L., Niranjan, K., Gilmour, S., Trinca, L.,(2001). Combined effect of operational variables and enzyme activity on aqueous enzymatic extraction of oil and protein from soybean. *Enzyme and Microbial Technology*. 28, 499-509
- Sulaiman, M.R., Hussain, M.K., Zakaria, Z.A., Somchit, M.N., Moin, S., Mohamad, A.S., Israf, D.A.,(2008). Evaluation of the antinociceptive activity of Ficus deltoidea aqueous extract. *Fitoterapia*. 79, 557- 561.
- Zainah, A., Muhajir, H., Amin, I., Shafii, K., & Norazizah M., (2010). Antihyperglycemic and glucose tolerance activity of Ficus deltoidea ethanolic extract in diabetes rats. *Jurnal Sains Kesihatan Malaysia*. 8(1), 25-30.

# **APPENDICES**









# Hydrodistillation















#### Concentration















# $\frac{\text{Temperature}}{25^{0}\text{C}}$









